

CROSS-TOLERANCE MECHANISM INDUCTION IN MELON SEEDS BY PRIMING PRIOR DRYING

Indução do mecanismo de tolerância cruzada em sementes de melão osmoticamente condicionadas antes da secagem

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ABSTRACT

The loss of benefits after re-drying is one of the drawbacks of the seed priming technique. Different types of stresses have been used before re-drying to preserve the priming benefits. This process may be seen as promoting cross tolerance to increase the defense mechanisms that prevent loss of viability in seeds primed after drying. We tested the effect of some stresses to induce cross-tolerance and different drying conditions with the aim of maintaining priming benefits in melon seeds. The seeds were primed in an aerated KNO₃ solution (0.35M), -1.7MPa, 25 °C, in the dark for six days. The primed seeds were then submitted to slow drying, fast drying, cold shock + slow drying, cold shock + fast drying, heat shock + slow drying, heat shock + fast drying, PEG + slow drying, PEG + fast drying, ABA + slow drying, ABA + fast drying and no drying (planted directly after priming). We evaluated antioxidant enzyme activities (SOD, CAT and APX), germinability, mean time of germination (MTG) and mean rate of germination (MRG). A completely randomized design was used with three repetitions of 50 seeds in each treatment. Data were analyzed by ANOVA and means were compared by the Scott-Knott test ($p \leq 0.05$). ABA increased SOD activity after drying and CAT activity was reduced by priming. APX activity was not observed. The stress submission prior to re-drying improved the MRG and reduced MTG. Therefore, the induction of the cross-tolerance mechanism could be effective to maintain priming benefits in melon seeds.

Index terms: PEG; priming; dry seeds; antioxidant system.

RESUMO

A perda dos benefícios do condicionamento, após a ressecagem, é um dos problemas do condicionamento. Para conservar os benefícios, têm sido utilizados diferentes estresses antes da secagem. Esse processo poderia agir como tolerância cruzada, aumentando os mecanismos de defesa, prevenindo a perda de viabilidade em sementes condicionadas após a secagem. Assim, testou-se o efeito de estresses, na tentativa de induzir tolerância cruzada, após diferentes secagens, mantendo os benefícios do condicionamento em sementes de melão. Dessa forma, as sementes foram condicionadas em solução aerada de KNO₃ (0,35M), -1,7 MPa, 25 °C, no escuro, por seis dias. Depois disso, foram submetidas à secagem lenta, rápida, choque de frio + secagem lenta e rápida, choque de calor + secagem lenta e rápida, PEG + secagem lenta e rápida, ABA + secagem lenta e rápida e sementes sem secagem. As sementes não condicionadas foram o controle. Foi avaliada a atividade das enzimas SOD, CAT e APX, germinabilidade, tempo médio de germinação e taxa média de germinação. O delineamento experimental utilizado foi inteiramente casualizado (DIC) com três repetições, contendo 50 sementes para cada tratamento. Para a análise estatística dos dados, foi utilizado ANOVA e as médias comparadas pelo teste de Scott-Knott ($p \leq 0,05$). A aplicação de ABA aumentou a atividade da SOD após a secagem e o condicionamento reduziu a atividade da CAT. O estresse inicial, antes da secagem, aumentou a e reduziu o tempo médio de germinação. Contudo, a indução do mecanismo de tolerância cruzada pode ser efetiva na manutenção dos benefícios do condicionamento.

Termos para indexação: PEG; condicionamento; sementes secas; sistema antioxidante.

INTRODUCTION

The Brazilian fruit production is characterized by a wide diversity of products, of which only a small part is sold internationally on a large scale. According to data from the Brazilian Institute of Fruits - IBRAF (2013), fruit production in Brazil is approximately 43.6 million, and covers an area of 2.2 million hectares.

Brazil is currently the world's third-largest producer of fruit after China and Turkey. In international markets, Brazil's fruit industry occupies an important position by exporting 693 000 t fresh fruit per year. The melon (*Cucumis melon* L.) is the fresh fruit with highest exported volume (almost 182 thousand tonnes) in Brazil (IBRAF, 2013).

Melon is a species of the Cucurbitaceae family and has low germination and growth at both low temperatures

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(below 13 °C) and high ones (above 40 °C) (Edelstein; Nerson, 2009). This is a problem to melon cultivations in northwestern and southern Brazil. As a result, priming of melon seeds has been used to increase germination and vigor under stressful conditions (e.g., low temperature, high temperature, drought, high salinity, etc.) (Farooq et al., 2007; Nascimento, 2005; Farhoudi; Saedipour; Mohammadreza, 2011).

Priming is a treatment used to improve seed performance under stressful environmental conditions (Ashraf; Foolad, 2005). Seed germination performance (germination rate, germination time and plant uniformity) is very important for successful production of melons in these areas because seeds are occasionally sown under unfavorable temperature conditions due to the local climate.

Priming of melon seeds with KNO₃ (potassium nitrate) was observed to improve germination rate, germination speed, homogeneity of plants and invigoration (Nascimento; Aragão, 2002). However, primed melon seeds cannot be stored due to their high moisture, requiring them to be adequately dried before storage (Butler et al., 2009). But this subsequent drying can reduce or eliminate the priming benefits as a consequence of germination having advanced to a state of lost desiccation tolerance (Sliwinska; Jendrzczak, 2002), making them more susceptible to damage during dry storage (Powell et al., 2000). The temperature and duration of drying after priming affected the vigor and viability of melon seeds (Nascimento; West, 2000).

The effect of priming, particularly with respect to maintenance of benefits and subsequent longevity, can be influenced by the conditions immediately after priming (Butler et al., 2009). Seeds dried at different rates (slow or fast) after priming showed differential gene expression, similar to what occurs with genes that act to protect DNA and promote tolerance to stress (Soeda et al., 2005).

Drying of seeds that have partially lost desiccation tolerance imposes stress, and the resultant injury varies according to drying speed. Primed seeds inevitably endure some injury by the drying to tolerate subsequent stress (Chen; Arora, 2013). Together with drying after priming, low and high temperature, water stress and abscisic acid (ABA) can be used to maintain priming benefits (Lin et al., 2005; Sabehat; Lurie; Weiss, 1998). Drying after priming might work as a cross-tolerance mechanism to activate the seed defense system to subsequent severe stress (Genoud; Metraux, 1999; Kranter et al., 2010).

The antioxidant system is regarded as one of the major defense systems in seeds, operating through enzymatic and non-enzymatic pathways by scavenging reactive oxygen species (ROS) (Chen; Arora; Arora, 2010).

The detoxifying mechanisms play an important role in acquisition of desiccation tolerance, completion of seed germination and seed storability (Bailly, 2004).

Thus, the aim of this study was to evaluate the effects of slow and fast-drying associated or not with low and high temperature, water stress and ABA after priming, to promote cross-tolerance and improve antioxidant system activity and maintain priming benefits in melon seeds.

MATERIAL AND METHODS

Plant material and priming

Melon seeds (*Cucumis melon* cv.), cv. Hales Best Jumbo, were purchased from Feltrin Co. Ltd. and submitted to priming in 0.35M KNO₃. During priming, the seeds were maintained under continuous aeration at 20 °C in the dark for six days (Nascimento, 2004). The primed seeds were then washed twice with distilled water and dried to original moisture content (7.0%) through different treatments before performing germination tests.

Treatments for cross-tolerance induction

The cross-tolerance mechanism was induced using a combination of eight stress conditions: cold shock (CS) + slow drying (SD); cold shock + fast drying (FD); heat shock (HS) + slow drying; heat shock + fast drying; PEG + slow drying; PEG + fast drying; ABA + slow drying; and ABA + fast drying. Besides these combination of factors, as control we formed groups subjected to slow drying, fast drying, no drying as well as unprimed seeds.

Cold shock treatments were at 1 °C, while for heat shock the seeds were submitted to 45 °C, for two hours in both treatments. Drought stress was imposed with PEG (6000), -1.7 MPa, at 5 °C in the dark for three days (Buitink et al., 2003; Faria et al. 2005). Hormo-priming consisted of using 10µM ABA (Vieira et al., 2010).

To slow dry the seeds, we used an open top plastic pot lined with paper towel, at room temperature of 25 °C and 50% RH (relative humidity), to reach the initial seed water content (about 24 hours) of 5-7 gH₂O g DM⁻¹. Fast drying was performed in a flow chamber, temperature of 32 °C, until initial water content (about 2 hours).

Antioxidant activity

SOD activity was measured by the method described by Giannopolitise and Ries (1977), adapted for tomato seed analysis. The assay mix included: 50 mM potassium phosphate buffer (pH = 7.8), 14 mM methionine, 75 µM nitro blue tetrazolium, 2 µM riboflavin, 100 nM EDTA and 20 µl enzyme extract. Reaction was initiated by adding riboflavin and placement under fluorescent light (15 W), and

was allowed to continue for 40 min, when it was ended by switching off the light and covering the tubes with a black cloth. Subsequently, the absorbance of the mixture was measured at 560 nm with a Biotek Epoch spectrophotometer. One unit of SOD activity was defined as the amount of the enzyme required to reach 50% inhibition of the reaction in the “minus enzyme extract” control, which should have a higher absorbance compared to the samples with enzyme extract.

CAT activity was measured using the protocol Havar and McHale (1987). The assay mix included 200 mM potassium phosphate buffer (pH 7.0), 20 μ l enzyme extract, and 120 μ l 12.5 mM H_2O_2 solution. One unit CAT activity was defined as the degradation 1 μ M H_2O_2 during one minute at 240 nm (using extinction coefficient $\epsilon=36\text{ mM}^{-1}\text{ cm}^{-1}$).

APX activity was measured using the protocol of Nakano and Asada (1981) modified for tomato seeds. The assay mix was composed of 50 mM potassium phosphate buffer (pH 7.0), 0.5 mM ascorbic acid (ASA), 0.1 mM EDTA, 20 μ l enzyme extract. H_2O_2 (0.1 mM) was added to initiate the reaction. One unit of APX was defined as the conversion of ASA (1 μ M) into the monodehydroascorbate at 290 nm, during one minute ($\epsilon=2.8\text{ mM}^{-1}\text{ cm}^{-1}$).

Measurements of the germination process

Four replications containing 50 seeds were used as samples. Measurements were performed on paper rolls using Germitest® paper. The rolls were moistened with distilled water and placed in a germination chamber at 25 °C. The number of germinated seeds was recorded daily for seven days. Each seed was considered germinated when its radicle reached 2 mm. At the end of this period, germinability (G%), mean germination time (t), and mean germination rate (v) were calculated (Ranal et al., 2009).

Statistical design

The experiment was completely randomized with four replications and 50 seeds per replicate for each treatment. The data on antioxidant activity and germination were subjected to the Shapiro-Wilk test and analysis of variance (ANOVA). The differences between the means were compared using the Scott-Knott test ($P<0.05$).

RESULTS AND DISCUSSION

Antioxidant activity

The antioxidant activity was evaluated through enzymatic activity. The SOD activities of the re-dried seeds were similar to those of the control seeds. Only priming seeds with ABA increased SOD activity after drying (slow or fast) (Figure 1).

The activity increase in SOD can be seen as a cross-tolerance effect induced by ABA. This phytohormone is involved to signaling the accumulation of stress proteins (LEAs and HSPs) and antioxidant system (Baxter; Mittler; Suzuki, 2013), related to protection of the membrane for desiccation tolerance (Catusse; Job; Job, 2008). Moreover, the priming with ABA can act as a cross-tolerance activating defense system of seeds to drying. Just as in desiccation tolerance, increased activity in SOD triggered by ABA leads to a reduction in ROS levels and consequently less damage to the membrane structure when submitted to water loss (Eisvand et al., 2010). As a result, such seeds should maintain vigor and priming benefits after drying. The priming in melon seeds reduced CAT activity in relation to the control seeds (unprimed) (Figure 2). The lower CAT activity in primed seeds may be due to less ROS production or activation of non-enzymatic antioxidant pathways.

In dry seeds the non-enzymatic antioxidant system plays a more important role than the enzymatic system (Baillly, 2001). Thus, priming associated with stress and ABA could lead to increase of antioxidant molecule synthesis, reducing CAT activity and improving the seed germination performance after re-drying.

The priming with PEG with low osmotic potential (drought simulation) was the only priming treatment that had a smooth increased in CAT activity. This may have been because melon seeds under drought stress increase their antioxidant activity (Farhoudi; Saedipour; Mohammadreza, 2011).

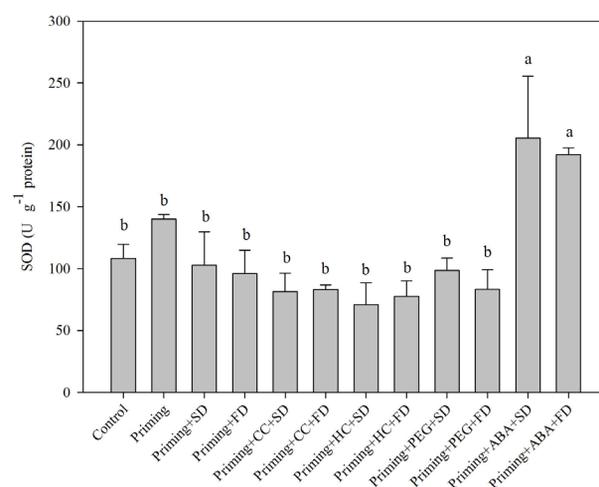


Figure 1: Mean values and standard errors for superoxide dismutase (SOD) activity in melon seeds observed under different conditions. Means were compared by the Scott-Knott test.

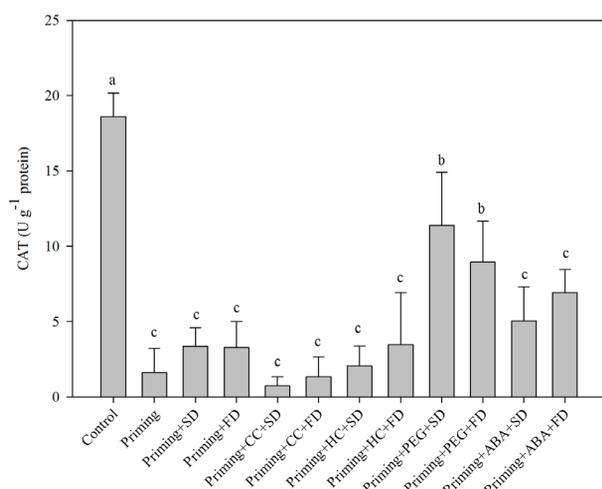


Figure 2: Mean values and standard errors for catalase (CAT) activity in melon seeds observed under different conditions.

Means were compared by the Scott-Knott test.

Of the enzymes tested only APX did not show activity. In some cases, this may have been due to the complex action mechanism of ascorbate–glutathione cycle, which in dry seeds is not activated (Bailly, 2001).

Measurements of the germination process

The priming reduced germinability in relation to control, and cross-tolerance treatment improved germinability (Figure 3). Among the cross-tolerance treatments, only priming + ABA + SD did not improve germinability (G%) when compared with the control.

In seeds with high vigor, priming can reduce germinability (Bray, 1995). For tomato, carrot and asparagus seeds, beneficial effects of priming were observed only for low-vigor seeds (Alvarado; Bradford, 1988a; 1988b; Cantliffe; Elballa, 1994; Bittencourt et al., 2004). Priming of melon seeds increased germination performance in seeds of with low vigor (Nascimento, 2004).

The germinability improvement observed in the cross-tolerance treatment may be related to a possible drop of ROS concentration and consequently lower CAT activity, as observed in the seeds submitted the cross-tolerance treatment. The higher the scavenging capacity of ROS in seeds, the larger the germinability will be (Lara et al., 2014).

The positive effect on the germinability was not observed for the priming + ABA + SD treatment. Although ABA treatment increased SOD activity, treatment with ABA plus slow drying led to reduction in germinability. This can partially be explained by the dormancy effect induced by

ABA (Catusse et al., 2008) and by fewer germinated seeds because of the increased germination time (t) and reduced germination rate (v). However, fast drying appears to have softened the dormancy effect induced by ABA.

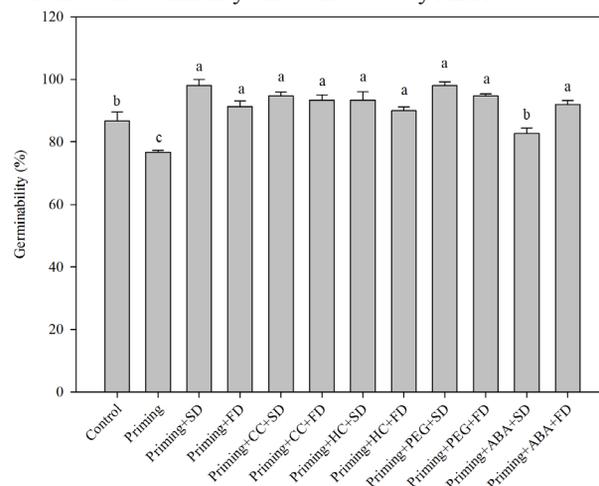


Figure 3: Mean values and standard errors for germinability (%G) of melon seeds observed for different conditions. Means were compared by the Scott-Knott test.

In general, the cross-tolerance treatment reduced mean germination time (t) and increased mean germination rate (v) of melon seeds (Figure 4).

Slow drying appears to have the strongest effects regarding improved germination time (t) and germination rate (v). We observed that slow drying was able to enhance these effects when applied after application of other stresses.

Slow drying after priming in the temperature range 20-30 °C seems not to decrease the activity of antioxidant enzymes such as superoxide dismutase, catalase and peroxidase (Sedghi, 2013). Thus, seeds which were dried slowly remained with a more active antioxidant system. Furthermore, slow drying has been found to be correlated with increases in chaperon protein levels (Lin et al., 2005). However, it seems that drought stress may maintain activity of antioxidant enzymes after fast drying. Considering the desiccation tolerance of seeds, when they were subjected to water stress beforehand, they maintained their viability afterward (Buitink et al., 2003).

As previously shown, ABA improved SOD activity and maintained germinability. Nevertheless, melon seeds with ABA cross-tolerance treatment had longer mean germination time and lower mean germination rate. These reactions may have been due to the dormancy effect of ABA, leading to slower germination.

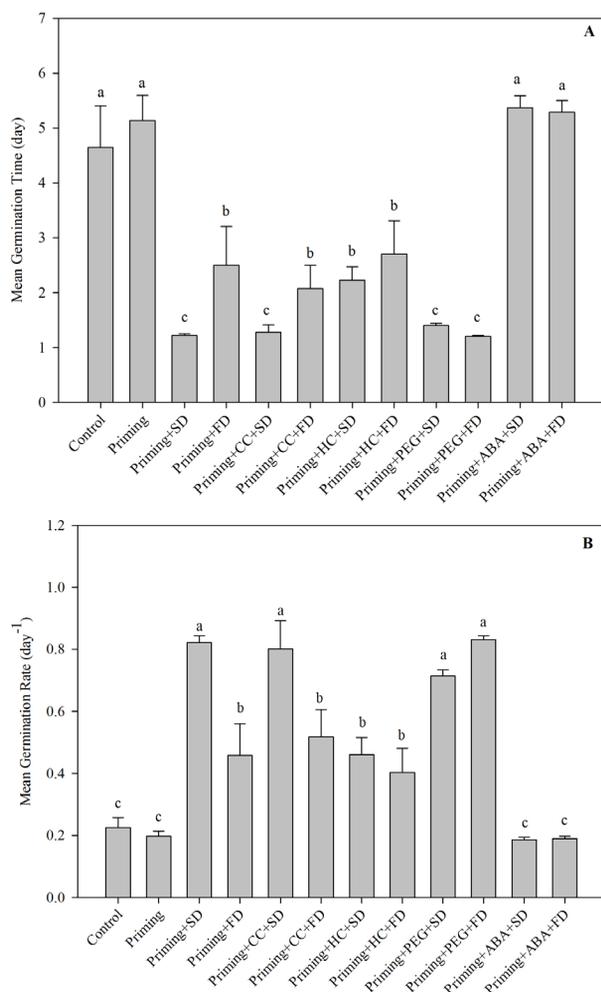


Figure 4: Mean values and standard errors for (A) mean germination time (day) and (B) mean germination rate in melon seeds observed under different conditions. Means were compared by the Scott-Knott test.

CONCLUSIONS

The results suggest that stress submission subsequent to priming could work promote cross tolerance in melon seeds, maintaining the priming benefits even after drying. Also, slow drying seems more promising than fast drying after priming.

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