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Alternative Control of Plant Pathogen Fungi Through Ethanolic Extracts of Avocado Seeds (Persea Americana Mill.)

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ABSTRACT

The aim of this study was to evaluate the antifungal action of ethanolic extracts of avocado seeds, by conducting two experiments in the laboratory of plant pathology of EPAMIG SUL/Lavras, MG, Brazil, in a completely randomized design with three replicates. The first consisted of the following treatments ('Breda' 3%, 'Breda' 2%, 'Margarida' 3%, 'Margarida' 2%, Control, and Ethanol) testing its inhibitory effect on two fungal species (Colletotrichum gloeosporioides and Monilinia fructicola) and the second consisted of different concentrations of extracts (0%, 0.25%, 0.50%, 0.75%, 1.0%, 1.5%, and 2.0%) on the fungus F fructicola. The evaluations were performed at three times (7, 14, and 21 days of incubation) by measurements of the mycelial diameter using a digital caliper. The data were submitted to analysis of variance and the averages were compared by Scott-Knott test at 5% probability. The results demonstrated the positive potential of the ethanolic extracts of avocado seed on the mycelial development of fungi M. fructicola and C. gloeosporioides during the evaluated days.

Keywords: antifungal activity, Colletotrichum gloeosporioides, Monilinia fructicola, postharvest



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INTRODUCTION

Fruit-growing is one of the most important segments of Brazilian agriculture, being Brazil the third largest producer of fresh fruit, surpassed only by China and India. Nevertheless, Brazil's share of world fruit exports is still incipient, accounting for less than 3% of its production¹. One of the main limiting factors to the increase in the export of fresh fruits is postharvest losses caused by pests, nematodes, physiological disturbances, mechanical damage and diseases caused by plant pathogen microorganisms, including fungi.

As an example of these fungi, *Colletotrichum gloeosporioides* and *Monilinia fructicola* are plant pathogen fungi involved in a large number of postharvest diseases, being developed primarily in the fruit storage stage^{2, 3}.

C. gloeosporioides is responsible for anthracnose, a disease that attacks temperate, subtropical and tropical fruit trees. Moreover, the genus *Colletotrichum* is responsible for other diseases in a wide variety of crops, including cereals, legumes and vegetables⁴. According to Phoulivong et al.⁵, the importance of this pathogen goes beyond postharvest losses, since it allows a market's behavioral variation, leading to socioeconomic changes.

The pathogen *M. fructicola* is the causative agent of brown rot, the main postharvest disease of seed rosacea (peach, plum and nectarine)⁶.

Traditionally, the control of postharvest diseases of fruits is based on the use of synthetic chemicals, especially fungicides. However, due to the several problems caused by them, such as harmful effects on human health and the environment, besides problems with plant pathogen resistance, has driven the search for more efficient controls and less aggressive to society and the environment^{7, 8}.

Reports on the use of biofungicides extracted from plants are common over the centuries, these are rich in compounds as essential oils, terpenoids and alkaloids, besides lectins, polypeptides and phenolic substances and polyphenols, which are subdivided into single phenols, phenolic acids, quinines⁹.

In the case of the avocado *Persea americana* Mill., the species contains specialized idioblast cells that are almost completely filled with alkaloid-containing oil, sesquiterpene hydroperoxides and possibly other terpenes¹⁰, being the most active constituent known as "persina" ¹¹. Moreover, this substance has many biological activities described by Rodriguez-Saona et al.¹² and belongs to the group of acetogenins, which are biologically active substances, restricted to the families of *Annonaceae* and *Lauraceae*¹³.

Several studies using extracts and essential oils of different plant species have been carried out in several countries¹⁴⁻¹⁹, showing its potential in control of plant pathogens through its direct fungitoxic action, acting as inhibitor of mycelial growth, formation of appressorium and the fungal spore germination, besides the effect on plant by the ability to induce the accumulation of phytoalexins in resistance mechanisms²⁰.

Based on the above, this study aimed to evaluate the antifungal activity of ethanol extracts of avocado seeds (*Persea americana* Mill.) in the control of mycelial growth *in vitro* of fungi *Colletotrichum gloesporioides* and *Monilinia fructicola*.

MATERIAL AND METHODS

The study was developed in the Laboratory of Plant Pathology of EPAMIG SUL/Lavras - MG, Brazil, from March to August 2015 and was constituted by *in vitro* tests.

Obtaining the isolates

For tests involving ethanol extract, isolates of *Colletotrichum gloeosporioides* and *Monilinia fructicola* from apple (*Malus domestica* L.) and peach (*Prunus persica* (L.) Batsch) were used, respectively.

Apple fruits with symptoms of anthracnose and peach with symptoms of brown rot were washed in running water and subsequently disinfested in 1% sodium hypochlorite solution for 1 min, then transferred to a vessel containing distilled water. The excess moisture was withdrawn with sterile filter paper. After drying the fruits, a wet chamber was made at 25°C for the development of fungi present in the fruit for later identification (seven days). The fragments containing the fungal structures of interest were plated in PDA (Potato-Dextrose-Agar) culture medium under aseptic conditions and incubated in a growth chamber (BOD) at $25 \pm 2^{\circ}$ C for seven days. The identification of the fungi was done through taxonomy based on morphological aspects. After identification of fungi were stored in mycology collection of the Laboratory of Plant Pathology EPAMIG.

At the moment of the assay assembly, the fungi were repixed in PDA medium and conditioned in BOD for seven days to obtain an active culture, which were used for mycelial growth inhibition tests.

Obtaining the ethanolic extract

The extracts were obtained from two commercial cultivars of avocado (*Persea americana* Mill.) 'Breda' and 'Margarida'.

The seeds of fruits from the cultivars constituted the plant material used to accomplishment of the extracts. Ten seeds of each cultivar were collected from an avocado producing farm in São Sebastião do Paraíso, Minas Gerais. After the collection, they were placed for drying in natural environment until reaching 10 to 15% of humidity.

After drying, the seeds were crushed to the mill powder. A total of 20 g of the dried material each cultivar was weighed, which was immersed in 100 ml of 70% ethanol (v/v), separately. The solutions were stored in glass bottles, closed and wrapped in aluminum foil so that there was no interference of light. This condition was maintained for 30 days and the solutions were stirred three times a day during this time. Finally, these solutions were filtered using a funnel with a layer of hydrophilized gauze. From the filtration, 100 ml of the extracts were obtained which were kept in a light-free environment until the test was carried out. The filtrate was added to the potato-dextrose-agar (PDA) culture medium in order to obtain the different concentrations of the extracts to be evaluated. The dilutions used in the two tests started from the initial extract and were made according to the pharmaceutical method of maceration²¹.

Verification of the antifungal activity of ethanolic extracts

The experiment consisted of two tests: First test was conducted with high concentrations as a starting point to define the fungitoxic activity of the extracts on fungi growth. In this first trial the tests were composed of the following treatments: (Breda 3%, Breda 2%, Margarida 3%, Margarida 2%, Control and Alcohol), and two species of fungi (*C. gloeosporioides* and *M. fructicola*). The evaluations were performed at three times (7, 14 and 21 days of incubation) by measurements of the mycelial diameter, using a digital caliper. All tests were performed *in vitro*, in a laminar flow hood with all sterilized equipment, to maintain a complete asepsis condition.

After the results of the first test, *M. fructicola* fungus that had high percent inhibition with the 2% and 3% extracts composed the second assay. For this, smaller concentrations were used: alcohol extract (0%, 0.25%, 0.50%, 1.0%, 1.5%, and 2.0%) were used for the two avocado cultivars. The evaluations were performed at three times (7, 14, and 21 days of incubation) by measurements of the mycelial diameter using a digital caliper, being calculated the inhibition percentage by the following formula adapted from Edington et al.²²: I = [(GFC - GFT) / GFC] x 100, Where: I = inhibition percentage; GFC = growth of fungus in the control; GFT = growth of the fungus in the treatment. All the experiments were conducted in a randomized complete block design with three replicates. Each experimental plot was represented by a 9 cm Petri dish containing 15 ml PDA and 0.3 cm diameter discs containing fragments of the pathogens. The data were submitted to analysis of variance, and the averages were compared by the Skott-Knott test at 5% error probability, using the SISVAR software²³.

RESULTS AND DISCUSSION

The results of the effect of ethanol extracts on the growth of *in vitro* fungi *Colletotrichum gloeosporioides* and *Monilinia fructicola* are presented in Table 1. The analysis of variance indicated differences in the antifungal activity of plant extracts on the pathogens used in the study.

Ethanolic extracts	Days		
	7	14	21
	Monilinia fructíicola		
Breda 2%	77.43 a	89.09 a	91.82 a
Breda 3%	79.52 a	90.86 a	93.56 a
Margarida 2%	79.52 a	90.86 a	89.80 a
Margarida 3%	79.52 a	90.86 a	93.56 a
Alcohol	37.92 b	8.45 b	1.92 b
Control	0.00 c	0.00 b	0.00 b
CV (%)	40.49	31.88	11.89
	Colletotrichum gloesporioides		
Breda 2%	46.64 b	10.87 a	8.96 a
Breda 3%	58.41 a	17.75 a	2.54 a
Margarida 2%	51.23 b	15.94 a	4.71 a
Margarida 3%	59.37 a	21.01 a	10.14 a
Alcohol	19.80 c	3.62 b	3.98 a
Control	0.00 d	0.00 b	0.00 a
CV (%)	6.48	6.55	4.69

Table 1. Percent inhibition (%) of *Monilinia fructicola* and *Colletotrichum gloesporioides* under the influence of different ethanol extracts at different concentrations.

Averages followed by the same letter on the columns do not differ among themselves by Scott-Knott test at 5 % probability.

For *M. fructícola*, it was observed that regardless of the used extract, all showed antifungal activity on colony growth with increased efficiency throughout the evaluation period, with the highest inhibition percentage at 21 days, with an average of 93.56% for the extracts of 3% for both evaluated cultivars. These results show that ethanol has efficiency to extract the antifungal substances for this pathogen.

However, for *C. gloeosporioides*, the efficiency was only observed up to the 14th day of evaluation, with the higher average percentages of inhibition in the first seven days (58.41% and 59.37%) for the extracts from the cultivar

Breda and Margarida, respectively, both at concentrations of 3%. Similar results were found by Pansera *et al.*²⁴, where the essential oil of *Eucalyptus globulus* inhibited mycelial growth on the third day at concentrations of 0.15% and 0.20%. However, from the seventh day, there was pathogen growth at all tested concentrations. These results differed from the results found by Silva *et al.*²⁵, which obtained 100% inhibition percentage testing clove extract (*Syzygium aromaticum*), whereas the Nim extract (*Azadirachta indica* L.) inhibited 20.22% in a concentration of 3%.

From the 14th day of evaluation, there was a reduction in the inhibition capacity of *C. gloeosporioides*, regardless of the cultivar used to obtain the extract, which can be attributed to the evaporation of chemical constituents with inhibitory activity of extracts as well as to the instability of the same in the presence of abiotic factors such as light, heat, air and water inside the Petri dishes.

However, despite the results, both extracts proved to be much more efficient in the control of the pathogen causing brown rot on peach, being necessary to identify an ideal concentration for the control. Therefore, significant interactions were observed among the concentrations and the ethanol extracts in the mycelial growth of the pathogen at seven days (Fig. 1).

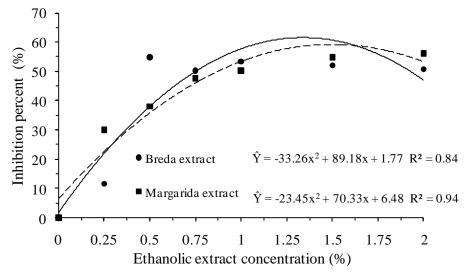


Figure 1. Percentage inhibition of mycelial growth (%) of *Monilinia fructicola* under influence of different ethanol extracts at different concentrations at 7 days of incubation.

Both extracts showed a quadratic model that showed the best fit to describe the inhibition percentage as a function of the used concentrations. The 'Breda' extract inhibited 61.55% of the mycelial growth when adjusted to the concentration of 1.35%, whereas 'Margarida' extract reached 59.25% of inhibition at concentration of 1.5%.

Similar results were found by Sellamuthu *et al.*²⁶, evaluating the antifungal effect of different concentrations of thyme (*Thymus vulgaris* L.), peppermint (*Mentha piperita* L.) and citronella (*Cymbopogen nardus*) oils in the vapor phase at seven days of incubation. However, the results differed from those found by Flores²⁷, in which the inhibition percentage was 78%, using 10% of canola extract (*Brassica napus*) in the control of *M. fructicola* at 8 days of incubation.

The present study showed a difference among avocado cultivars in the pathogen control in the first seven days. A possible explanation for this difference may be related to the contents of antifungal substances present in each cultivar, especially persina. Little is known of the mode of action of

persin. Most of the speculation on its potential mode of action has focused on the close similarity between persin and the monoglyceride of linoleic acid. Persin may mimic the monoglyceride of linoleic acid in glyceride synthesis ²⁸. Thus, persin appears to affect tissue cells by interfering with normal lipid biosynthesis²⁹.

At the 14th day of evaluation, there was no difference among the used cultivars, only among doses. The quadratic model presented the best fit in relation to the different concentrations (Fig. 2).

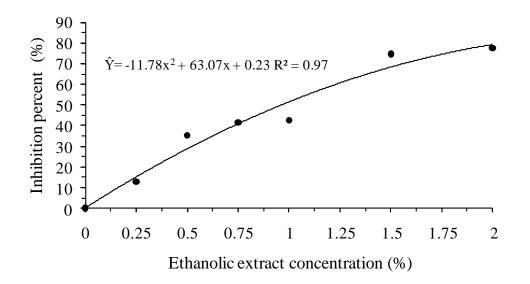


Figure 2. Percentage inhibition of mycelial growth (%) of *Monilinia fructicola* under influence of different ethanol extracts at different concentrations at 14 days of incubation.

It is noteworthy the inhibition of mycelial growth of *M. fructicola*, due to the lower concentration of the extract in relation to its absence, with a higher rate observed in the concentration of 2.68%, which showed values of 84.65%. These results corroborate those found by Elshafie et al.³⁰ in studies on antifungal activity of the essential oil of constituents *Origanum vulgare* L. and differ from those found by Pansera et al.²⁴ in studies performed with the use of different essential oils, in which essential oils of *Cymbopogum citratus* and *Cinnamonum camphora* completely inhibited the mycelial growth of the pathogen after three days at concentrations of 0.05% and 0.20% respectively, whereas *E. globulus* was not able to inhibit mycelial growth at any tested concentration.

There was a trend for linear increase in the inhibition percentage of mycelial growth at 21 days inasmuch as the extract concentration increased, with the highest value (76.4%) occurring when the concentration of 2.0% was used (Fig. 3).

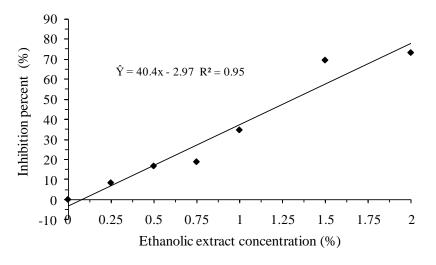


Figure 3. Percentage inhibition of mycelial growth (%) of *Monilinia fructicola* under influence of different ethanol extracts at different concentrations at 21 days of incubation.

These results are corroborate with Gomes et al.³¹, which also observed the trend to increase the percentage of mycelial inhibition with increasing concentration of cinnamon extracts on the pathogen Quambalaria eucalypti. Bernardo et al.³² also found that inasmuch as the concentration of extracts increased, there was an increase in the inhibition percentage, and the carqueja (Baccharis trimera) extract showed the best results on Alternaria alternata, Colletotrichum graminicola, Phytophthora sp., Rhizoctonia solani, and Sclerotinia rolfsii. However, they differed from Nascimento et al.³³ and Vasconcelos *et al.*³⁴, where the first author evaluated the antifungal activity of several extracts on the pathogen Cercospora calendulae and found that the confrei (Symphythum officinale L.) extract decreased the inhibition percentage inasmuch as the concentration increased, with rates of 15.03% in the concentration of 500 mg L⁻¹, reducing to 6.62% when used 10000 mg L⁻¹; the second author found that crude ethanol extract of cotton (Gossypium arboretum L.) leaves did not show antifungal activity in vitro against the fungus Lasiodiplodia theobromae in the tested concentrations, being possible to detect a growth increase of the fungus at the concentration of 10 mg mL⁻¹.

The results of this study show the efficiency of avocado seed extracts in the inhibition of *in vitro* mycelial growth of pathogens *C. gloeosporioides* and *M. fructicola* at low concentrations. This fact can be justified by the presence of acetogenins, since this compound is distributed throughout the plant, although its highest concentration is found in the seeds. This compound comprises a variety of structures, among them the persina, and their mode of action varies accordingly. Generally, they act as potent inhibitors of the respiratory chain, affecting the mitochondrial complex I, causing respiratory chain blocking through the inhibition of the NADH ubiquinone oxidoreductase, an essential enzyme in complex I. This prevents oxidative phosphorylation, directly affecting the electron transport in the cell mitochondria and causing a decrease in ATP levels, leading to apoptosis^{35, 36}.

CONCLUSIONS

Ethanol extracts from avocado seeds from cultivars Breda and Margarida were efficient in the *in vitro* control of *Colletotrichum gloeosporioides* and *Monilinia fructicola*, but *C. gloeosporioides* was susceptible only until the 14th day.

The highest control of *M. fructicola* was detected at 14th day with a dose of 2.68% and an inhibition rate of 84.65% when tested for different extract concentrations from both cultivars.

This study demonstrates that subsequent studies should be performed *in vivo* to confirm the inhibitory effect of these extracts on *C. gloeosporioides* and *M. fructicola*, by observing the disease symptom in the fruits.

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