CLINICAL ASPECTS OF THE POISONING BY THE PESTICIDE ENDOSULFAN

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Endosulfan is an organochlorine insecticide, widely used in insect control. Unfortunately, it is also an acute neurotoxic compound to both insects and mammals, including humans, and has been responsible for many severe poisonings and several fatal cases. Endosulfan also imitates or enhances the effect of the female hormone estrogen, having the capability of causing reproductive and developmental damage in both, animals and humans, and its exposure has been linked to liver tissue injury. This persistent lipophilic compound is one of the most abundant organochlorine pesticides in the environment, capable of undergoing long range transport to remote locations such as the Arctic. It is practically water-insoluble, but readily adheres to clay particles and persists in soil and water for several years. Its indiscriminate and injudicious use in the control of insects on a wide range of agricultural products and in the extermination of household pests, has considerably increased the hazard risk for human health. Also, this compound has a high fatality rate in humans when ingested accidentally, from food or water contaminated, or in suicidal cases. The aim of this article is to review and summarize chemical, biochemical, environmental, and toxicological data of endosulfan and draw attention to its toxicological potential risk to human health.

Keywords: endosulfan; organochlorine insecticides; hazardous substance; hormonal disruptor; health hazard.

INTRODUCTION

Endosulfan is an organochlorine manufactured insecticide which was first introduced in the 1950s, and is commonly known by its trade-name Thiodan[®]. It is widely used to control a number of insects on a wide range of agricultural products, such as grains, tea, fruits, vegetables, tobacco and cotton, among others. It is also used as a wood preservative.¹ The toxicity of endosulfan is well known on non-target organisms and has been responsible for many severe poisonings, including several fatal cases.²

Endosulfan is a highly toxic insecticide that produces tonic-clonic convulsions, headache, dizziness and ataxia. It can also cause life threatening metabolic disturbances.³ Treatment is only limited to symptomatic and supportive measures. Endosulfan is considered a persistent organic pollutant (POP) and belongs to the organochlorine insecticides group that includes molecules notorious for being POPs. High environmental persistence coupled with their potential for bioconcentration and biomagnification may lead to toxic levels in plants and animals. Nowadays, there is also a call for a worldwide ban on POPs because of their possible links to cancer and effects on hormones, the immune system, and reproduction. However, when compared with other POPs which disperse across the world, endosulfan tends to remain close to its region of use, although it has been found in high concentrations in many areas around the world because it is widely used.⁴

Endosulfan is associated with a high fatality rate in humans when ingested accidentally, from food or water contaminated, or in suicide cases.⁵⁻¹³ It is also a substance that imitates or enhances the effect of the female hormone estrogen, which means it can cause reproductive and developmental damage in both animals and humans. Researchers studying children from an isolated village in Kerala, India, have linked endosulfan exposure to delays in sexual maturity among boys.^{14,15}

As a result of its poisonous and persistent nature, endosulfan has been banned from many developed countries, and is gradually being phased out in others although extensive use continues in less-developed parts of the world. It is already banned in Australia, New Zealand, Singapore, Bangladesh, Brazil, Indonesia, Korea, Thailand, but is still in use in India, Taiwan and the US, as well as in some EU countries such as Denmark and Great Britain.

CHEMISTRY

Endosulfan (6,7,8,9,10,10-hexachloro-1,5,5a,6,9,9a-hexahydro-6,9-methano-2,4,3-benzodioxathiepin 3-oxide, CAS Registry Number 6994-04-3) is a polychlorinated compound (Figure 1). It is a cream to brown colored solid that may appear crystalline or in flakes. It has a distinct odor similar to turpentine and does not burn. Its physical properties are summarized in Table 1.

Endosulfan is produced by the Diels-Alder reaction of hexachlorocyclopentadiene with cis-butene-1,4-diol. The product is then reacted with thionyl chloride, liberating HCl. ¹⁶ This process produces a preparation containing approximately 30% of β -endosulfan and 70% of α -endosulfan (Figure 2).

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Table 1. Physical properties of endosulfan*

Physical Property	Value	Units	Temperature (° C)
Melting Point	106	°C	
log P (octanol-water)	3.83	(none)	
Water Solubility	0.325	mg/L	22
Vapor Pressure	1.73E-07	mm Hg	25
Henry's Law Constant	6.50E-05	atm.m³/mole	20
Atmospheric OH Rate Constant	1.00E-11	cm³/molecule.sec	25

^{*} Physical property data available at TOXNET.17

Figure 1. Chemical structure of endosulfan; 6,7,8,9,10,10-hexachloro-1,5,5a,6,9,9a-hexahydro-6,9-methano-2,4,3-benzodioxathiepin 3-oxide; CAS Registry Number 6994-04-3

alpha-Endosulfan

beta-Endosulfan

Figure 2. Endosulfan is a mixture of stereoisomers, designated α and β . β -endosulfan is a symmetrical compound, whereas α -endosulfan is asymmetric and exists as two twist chair forms. Technical product contains two stereoisomeric endosulfans, α - and β -endosulfan, in the proportion variously reported as from 4:1 to 7:3. The technical material is a 90-95 per cent pure mixture of the two isomers

STEREOCHEMISTRY

Endosulfan is actually a mixture of stereoisomers, designated α and β . α -Endosulfan is more thermodynamically stable, while β -endosulfan slowly converts irreversibly to the α -form at room temperature. Commercial endosulfan has traditionally been described as a mixture of two diastereoisomers in a ratio of 70% of α -endosulfan and 30% of β -endosulfan. Early reports on the conformation of the endosulfan isomers were based on incorrect NMR assignments. More recently, Schmidt *et al.* ¹⁹ used NMR spectroscopy and X-ray crystallography to determine the structure of the isomers. They reported that β -endosulfan is a symmetrical compound, whereas α -endosulfan

exists as two asymmetric isomers (Figure 2). In mammals, the α -isomer is significantly more toxic than the β -isomer, with a LD₅₀ of the α -isomer in rats of 76 mg/kg in comparison to 340 mg/kg for the β -isomer.²⁰

ENVIRONMENTAL TOXICOLOGY

Endosulfan is an important pollutant of high priority for international environmental agencies.²¹ It enters air, water and soil when manufactured or used as a pesticide, commonly applied to crops by using sprayers. Some endosulfan in the air may travel long distances before it lands on crops, soil, or water. Endosulfan on crops usually breaks down within a few weeks; however, it may stay in soil for several years before it all breaks down. Endosulfan does not dissolve easily in water; therefore, in surface water it is usually attached to floating soil particles or can be found attached to soil at the bottom. The small amounts of endosulfan that dissolve in water breaks down over time. Animals that live in endosulfan-contaminated soil and waters (invertebrates, insects, fish, shellfish, etc.) can build up endosulfan into their bodies, and achieve internal amounts of this pesticide that may be several times greater than in the surrounding water. Endosulfan is extremely toxic to fish and aquatic invertebrates, ^{22,23} however, the atlantic salmon (Salmo salar) tolerates dietary technical endosulfan levels up to 500 ug/kg.²⁴ The photochemical degradation route of endosulfan in water involves the formation of the endosulfan diol, its transformation to endosulfan ether and finally the ether's complete degradation.²⁵

This chemical also has been gradually implicated in mammalian gonadal toxicity, $^{26-29}$ genotoxicity, 30 and neurotoxicity. 31 In the environment, endosulfan can be converted by attack at the sulfite group via either oxidation or hydrolysis, to form the toxic endosulfate (endosulfan sulfate) and the nontoxic endodiol (endosulfan diol), respectively (Figure 3). $^{32-33}$ The degradation of endosulfan is stereoselective, 34 as β -endosulfan is hydrolyzed faster than the α -isomer to endosulfan diol, which is then rapidly degraded to endosulfan ether, endosulfan α -hydroxyether (major product), and endosulfan lactone. 35 The formation of endosulfate is thought to occur only through biological transformation, whereas hydrolysis to the diol occurs readily at alkaline pH. 36 It was observed that isolated strains of *Aspergillus niger* can degrade endosulfan to endodiol. 37 As endosulfan degrades to several different compounds their half-life in nature are unknown and highly dependent on a number of factors.

Martens³⁸ studied the degradation of endosulfan by different bacterial and fungal cultures, and found that endodiol and endosulfate, respectively, were the major metabolites accumulated. Besides, small amounts of endohydroxy ether and endolactone were also formed.³⁹ He has proposed a pathway, wherein, the endosulfan is converted to endosulfate followed by endodiol, endohydroxy ether and endolactone. Formation of these metabolites has also been confirmed by other investigators.⁴⁰⁻⁴² Unlike the isomers of endosulfan, the lipophilic metabolite endosulfate can accumulate in animal fat.^{1,43,44} As a result,

pasture contamination can result in unacceptably high endosulfate residues in locally grown production animals.

Endosulfan present at trace levels in air and total atmospheric precipitations of Paris was found by Trajkovska *et al.*⁴⁵ About its distribution predicate fact, endosulfan was also found in the eggs of broad-snouted caimans (*Caiman latirostris*), thus caiman eggs could be useful to biomonitor local contamination by endosulfan. The origin of endosulfan in caiman eggs is not known but appears to come from food contaminated by air during spraying.⁴⁶

Trace amounts of endosulfan (from non-detectable to 27 pg m⁻³) were recorded also in Australia's atmosphere in 2012, together with many other persistent organic pollutants.⁴⁷ Endosulfan sulfate was one of the most frequently pesticide detected in the Arctic, with peak deposition fluxes of 1.0 and 0.4 pg.cm⁻² per year. While endosulfan sulfate was more abundant than its parent compounds in most years, endosulfan (sum of α and β isomers) was predominant in 2003 and 2006 which, together with air mass backward trajectories, suggests a possible origin from ongoing use in Eurasia.⁴⁸

Potter *et al.*⁴⁹ measured endosulfan wet deposition in precipitation over a 4-year period within an area of high agricultural use in Southern Florida (USA) and in nearby Biscayne and Everglades National Parks. Endosulfan's two isomers and endosulfan sulfate were detected at high frequency with the order of detection and concentration being: β -endosulfan > α -endosulfan > endosulfan sulfate. Within the agricultural area, detection frequency (55 to 98%), mean concentrations (5 to 87 ng L⁻¹) and total daily deposition (200 ng m⁻² day⁻¹) exceeded values at other sites by 5 to 30-fold.

Endosulfan in soil can be degraded by some soil bacteria, as demonstrated by the findings of recently discovered endosulfan-degrading bacterial strain *Alcaligenes faecalis* JBW4 which was isolated from activated sludge. This strain is able to use endosulfan as a carbon and energy source.⁵⁰

The estrogenic activity of environmentally relevant doses of endosulfan was investigated by injecting ovariectomized adult rats once a day for 3 days with sesame oil (control), 0.02 mg kg⁻¹ day⁻¹ with 17 β-estradiol (an uterotrophic dose; UD), 0.0002 mg kg⁻¹ day⁻¹ with 17 β-estradiol (a non-uterotrophic dose; NUD), and 0.006, 0.06, 0.6 or 6 mg kg⁻¹ day⁻¹ of endosulfan. After 24 hours of treatment, the uteri were weighed (uterotrophic assay) and the luminal epithelial cell height (LECH), progesterone receptor (PR) and estrogen receptor alpha (ERalpha) protein levels were measured. PR, ERalpha, and complement factor-3 (C3) mRNAs were evaluated using real-time PCR. Uterine weight and LECH were only increased in UD-treated rats. PR, ERalpha and C3 expression levels were modified in most of the endosulfan-treated groups, showing an identical pattern of expression to the NUD-group.⁵¹ These results show that endosulfan mimics non-uterotrophic actions, strengthening the hypothesis that endosulfan is a widespread xenoestrogen.

BIOCHEMICAL DATA

Endosulfan administration to rats significantly increased the activity of choline kinase and phosphocholine cytidylyltransferase (both cytosolic and microsomal) of lung and liver. The induction of

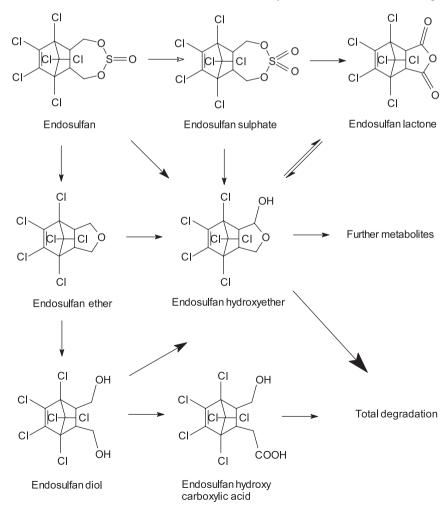


Figure 3. The proposed metabolic pathway for endosulfan in human, as published by Lee et al. 33

phosphatidylcholine (PC) biosynthesis in lung and liver of rats has been shown following the intratracheal administration of endosulfan (1 mg 100 g⁻¹ body weight) for 3 days.⁵² As pointed by Narayan *et al.*,⁵³ biochemical effects of endosulfan are different in many cases from other organochlorine insecticide. For example, DDT and endosulfan have similar effects on microsomal lipid metabolism, but produce different biochemical manifestations on the secretion of surfactant phospholipids.⁵⁴

Endosulfan significantly inhibited testicular androgen biosynthesis in adult rats (p.o. administered 7.5 and 10 mg kg⁻¹ body weight), consecutively for 15 and 30 days. No appreciable alterations were apparent in body weights, testicular wet weights, and cytosolic and microsomal protein contents of testis in treated rats. Profound decrease in the levels of plasma gonadotrophins (FSH and LH) along with plasma testosterone and testicular testosterone were observed at both administration treatments, particularly after the longer exposure of 30 days. Activities of 3 β - and 17 β -hydroxysteroid dehydrogenases were considerably lowered on longer exposure treatments. 26

Rousseau *et al.*⁵⁵ observed that endosulfan is able to induce a substantial increase of prolactin expression while not increasing cell growth. These results suggest that endosulfan could modulate an estrogen-inducible gene such as prolactin, possibly acting via second messenger-mediated cellular mechanisms instead of solely competing with estrogens for the nuclear estrogen receptor sites.

Endosulfan is reported to suppress humoral as well as cellular immune responses. It did not have any influence on nitrite production, but suppressed the LPS-induced TNF-α generation. ⁵⁶ Endosulfan exposure (8 and 16 mg kg⁻¹) to rats significantly decreased the activities of superoxide dismutase and catalase, reduced glutathione levels and increased lipid peroxidation. ⁵⁷ Endosulfan is also able to deplete glutathione (GSH) and induce apoptosis in human peripheral blood mononuclear cells (PBMC) *in vitro*. ⁵⁸

Oral rat administration of 10 mg kg⁻¹ body weight/day for two and four weeks showed toxic interference with liver and kidney's biochemistry and histology. Biochemical parameters, like aspartate amino transferase, alanine amino transferase, acid phosphatase, alkaline phosphatase, bilirubin, urea and creatinine, were increased, which clearly showed the hepatotoxic and nephrotoxic effects of endosulfan. Histopathologically findings included increased liver size, sinusoidal dilation, pyknotic nuclei, cytoplasmic degranulation and various nuclear aberrations. Similarly, pathological alterations were also observed in the kidney, especially chronic glomerulonephritis, glomerulosclerosis, adenoma and glomerulus deposits.⁵⁹

Endosulfan is an important hepatotoxic agent that generates free oxygen radicals in the liver. Results of the enzyme and histological analyses show that exposure of rats to endosulfan cause liver tissue damage, independent of the route of exposure.⁶⁰

Endosulfan can cause toxic effects on rabbit pancreas. Microscopy examination indicated degenerative changes and immunohistochemistry analyses showed marked decreases in proinsulin-, insulin-, and amylin-secreting cells and slight decreases in glucagon-secreting cells, whereas cells expressing caspase 3 increased.⁶¹

Endosulfan induced alterations in serum biochemical markers of oxidative stress and antioxidant capacity in rabbits, but vitamin C has an ameliorative effect. It was suggested that vitamin C supplementation might be helpful in preventing the detrimental effects of increased oxidative stress caused by endosulfan exposure.⁶²

Finally, endosulfan was able to inhibit cholinesterase activity *in vitro* and *in vivo* in Wistar rats and to cross the placental barrier and/or to be eliminated through milk, affecting the enzyme activity in male rat pups. ⁶³ *In vitro*, the enzyme activity was found to be inhibited in a concentration dependent manner.

HUMAN EXPOSURE

Humans are exposed to various environmental chemicals such as organochlorine pesticide residues, heavy metals, polychlorinate-dbiphenyls (PCBs), among many others. The most likely way for human endosulfan exposure is through consumption of contaminated food in countries where this pesticide was not banned yet. In fact, endosulfan has been found in food products such as oils, fats, and fruit and vegetable products.^{64,65} Exposure can also occur by smoking cigarettes made from tobacco that has endosulfan residues on it and by skin contact or inhalation, especially if safe handling and application procedures are not followed. Accidental spills and releases to the environment at hazardous waste disposal sites are other possible sources of endosulfan exposure.

Exposure to endosulfan has been reported in people living near hazardous waste sites through contact with contaminated soils. ⁶⁶ Pathak *et al.* ⁶⁷ first reported endosulfan levels in North Indian population. This study was designed to analyze the levels of organochlorine pesticide residues in maternal and cord blood samples of normal healthy women with full term pregnancy to gain insight into the current status of pesticide burden in newborns. Hexachlorocyclohexane was the main organochlorine present, followed by endosulfan. Published data indicates a transfer rate of 60-70% of these pesticides from mothers to newborns, which is of great concern as it may adversely affect the growth and development of newborn. No studies of fatal cases after inhalation exposure to endosulfan have been found.

Symptoms of endosulfan poisoning

Occupational exposure during manufacture and application has resulted in human poisoning cases. Symptoms of endosulfan poisoning have also been seen in people who intentionally or accidentally ingested large amounts of endosulfan. Most of these people experienced nausea, vomiting, headache, dizziness, convulsions or other nervous system effects, begining $2.7 \pm 0.5 \, h$ after ingestion. 68

Endosulfan poisoning in humans causes blood pressure and eletrocardiogram alterations.⁸ Over half of the patients developed complications, such as rhabdomyolysis, hepatic toxicity and hypotension.⁶⁹ These complications resolved without sequelae in the survival group. Refractory status epilepticus⁷⁰ was the most common cause of death in this series (75%). An ingestion of more than 35 g of endosulfan is considered as an independent variable to predict patient prognosis. Patients who have ingested more than 35 g (referred to a human weighing 70 kg) must then be immediately treated. Treatment is symptomatic, and includes gastric lavage, avoiding aspiration into the lungs, followed by intragastric administration of activated charcoal and 30 g of magnesium or sodium sulfate in 30% solutions.⁷¹ Convulsions require *i.v.* administration of benzodiazepines. In cases of skin contamination, decontamination with soap and water should also be included in the treatment.

Neurotoxicity

Endosulfan is an acute neurotoxic compound in humans. The US Environmental Protection Agency classifies it as Category I (Highly Acutely Toxic) while the World Health Organization classifies it as Class II (Moderately Hazardous). Symptoms of acute poisoning include hyperactivity, tremors, convulsions, lack of coordination, staggering, difficulty breathing, nausea, vomiting, and diarrhea. Doses as low as 500 mg kg¹ have been documented to cause death in humans and, in many cases, led to permanent brain damage. Farm workers with chronic endosulfan exposure have shown rashes and skin irritation. To Convulsions were reported in nine individuals exposed

to the endosulfan-containing insecticide Thiodan® during bagging.⁷³ Other effects noted in at least one of the subjects prior to the onset of convulsions, included malaise, nausea, vomiting, dizziness, confusion and weakness. Endosulfan can induce convulsions that could lead to brain damage at low doses.⁷⁴ A case of long-term, possibly permanent brain damage in an industrial worker was attributed by Aleksandrowicz⁷⁵ to endosulfan exposure.

As for its neurotoxicological mechanism of action, endosulfan interferes with the function of γ -aminobutyric acid (GABA), the main inhibitory neurotransmitter. It inhibits the binding of [35 S]-t-butylbicyclophosphorothionate (TBPS) to the picrotoxinin-binding site of the GABA receptor in rat brain synaptic membranes, interfering with chloride ion flux through the GABA-gated chloride channels (GABA_A), responsible for decreasing neuronal excitability.

Cardiotoxicity

Cardiovascular effects were part of the clinical syndrome displayed by a man who attempted suicide by ingesting 200 mL of a 30% endosulfan formulation. Although the man's stomach contents were aspirated and he was given activated charcoal to limit absorption during the first 16 hours following ingestion, episodes of tachycardia and hypertension occurred, followed by cardiogenic shock. Similar observations have been described in other fatal cases of acute intoxication. 67,77,78

Hepatotoxicity

Elevated serum alanine aminotransferase (ALT) and aspartate aminotransferase (AST) activities were reported in a woman 2 days after being admitted to hospital because of ingestion of endosulfan-contaminated food. The patient died 8 days after admission, following acute renal failure, disseminated intravascular coagulation, thrombi in the pulmonary arteries and aorta, and cardiogenic shock. *Postmortem* examination revealed dilation and congestion of hepatic sinusoids. Centrilobular congestion and slight prominence of the bile canaliculi were among *postmortem* observations in an additional fatal case of acute poisoning with endosulfan. The autopsy of a man who ingested approximately 260 mg kg⁻¹ showed liver congestion 4 days after exposure.

Nefrotoxicity

Hemorrhage of the medullary layer of the kidneys was reported in three persons who died following ingestion of endosulfan. ¹² Acute renal failure was a major contributor to the deaths of two individuals who ingested unknown amounts of endosulfan. ^{68,78} In both cases, *postmortem* examination revealed extensive tubular necrosis. In contrast, no kidney lesions were found in a man who died 4 days after ingesting approximately 260 mg kg⁻¹ of endosulfan. ⁷⁷

Immunotoxicity

The immune system is adversely affected by endosulfan due to white blood cell count decreases. ⁷⁹ Endosulfan also inhibits leucocytes and macrophage migration, causing adverse effects on humoral and cell-mediated immune system. ⁸⁰

Genotoxicity

The genotoxic effect of endosulfan is questionable. DNA damage in mononuclear leukocytes, as measured with the alkaline comet assay, was significantly increased in two of four French agricultural workers on the day following the application of pesticide mixtures (including endosulfan), when compared to levels of DNA damage prior to application.⁸¹ However, the contribution of endosulfan to the observed effect is uncertain because of co-exposure to fungicides, herbicides, and other insecticides. Evaluations for micronuclei in human peripheral blood lymphocytes provided mixed results, depending on the analytical method used.⁸² The results of all genotoxicity studies in humans should be treated with caution because the multiple-chemical exposures confound the interpretation, and exposure levels of endosulfan were not reported.⁸³⁻⁸⁵

Carcinogenicity

According to Antherieu *et al.*, ⁸⁶ endosulfan has not been classified by the International Agency for Research on Cancer (IARC) as a carcinogen, and was described by the International Programme on Chemical Safety (IPCS)⁸⁷ as not carcinogenic. However, despite no accurate data related to the carcinogenicity of endosulfan in humans is available, there are concerns about possible carcinogenic properties in chronic exposures.⁸⁸ Some studies have also shown that it induces proliferation of human breast estrogen sensitive MCF7 cells *in vitro*, which may lead to greater breast cancer risk.⁸⁷⁻⁹⁰ Studies also indicate the contribution of endosulfan in the combined effect of environmental estrogens in inducing breast cancer.⁸⁸⁻⁹³

Teratogenicity

Physical malformations observed in fetus which fathers were exposed to endosulfan 8 hours *per* day, in spring and winter, during 1 – 20 years of work, include cleft palates, harelips, club feet, limb malformations, eye deformities and extra fingers and toes. 4 Low concentrations of endosulfan around 0.1 nM have also shown to strongly inhibit the ability of human sperm to fertilise ova *in vitro*. 5 According to Lemaire *et al.* 6 endosulfan disrupts the retinoid signalling pathway in cells, and this is thought to explain the teratogenic effect of long-term exposure to low levels of the chemical, as has been experienced in India, 4 as retinoids play an essential role in the proliferation, development and differentiation of cells.

ANIMAL TOXICITY

Similar to the data from humans, respiratory effects have been observed in animals tested almost exclusively in acute, lethal-dose exposure. Male rats given single gavage doses of 200 mg kg⁻¹ of endosulfan exhibited dyspnea and cyanosis prior to death. ¹² Necropsy revealed hemorrhages in the interalveolar partitions of the lung and acute emphysema of the lungs. ⁹⁷ Also studies in which a limited number of dogs were given single oral doses of endosulfan as low as 10 mg kg⁻¹ or 50 mg kg⁻¹, demonstrated respiratory paralysis and death. Autopsy of the dogs revealed congestion of the lungs. Local inflammation of the lungs and dilated alveoli were observed in rats administered 10 mg kg⁻¹ day⁻¹ of endosulfan in peanut oil by gavage for 15 days. ⁹⁸

Acute toxicity of endosulfan for different animals and administration routes is summarized in Table 2.

Neurotoxicity

Sheep was reported to become blind one week after exposure to pasture previously sprayed with endolsulfan, although recovery was observed after one month.¹¹⁰ Similar exposure in lambs and pigs showed ataxia and inability to stand.¹¹¹

Table 2. Toxicity parameters of endosulfan acute toxicity

Organism	Test Type *	Route**	Reported Dose	References
Mouse	LD ₅₀	i.p.	7.0 mg.kg ⁻¹	99
Mouse	LD_{50}	p.o.	7.36 mg.kg ⁻¹	100
Rat	LC_{50}	inhalation	80 mg.m ⁻³ , 4 hr	73
Rat	LC_{50}	inhalation	12.6 mg.m ⁻³ , 4 hr (female) 34.5 mg.m ⁻³ , 4 hr (female)	101
Rat	LD_{50}	i.p.	8 mg.kg ⁻¹	1
Rat	LD_{50}	skin	34 mg.kg ⁻¹	102
Rat	LD_{50}	unreported	40 mg.kg ⁻¹	103
Rat	LD_{50}	p.o.	18 mg.kg ⁻¹	104
Rabbit	LD_{50}	s.c.	360 mg.kg ⁻¹	1
Rabbit	LD_{50}	p.c.	90 mg.kg ⁻¹	73
Rabbit	LD_{50}	i.p.	80 mg.kg ⁻¹	105
Rabbit	LD_{50}	p.o.	118 mg.kg ⁻¹	106
Cat	LC_{50}	inhalation	90 mg.m ⁻³ , 4 hr	102
Cat	LD_{50}	p.o.	2 mg.kg ⁻¹	107
Bird - wild	LD_{50}	p.o.	35 mg.kg ⁻¹	108
Man	TDLo	p.o.	260 mg.kg ⁻¹	77
Man	TDLo	p.o.	86 mg.kg ⁻¹	76
Women	TDLo	i.v.	6 mg.kg ⁻¹	109

^{*} Toxicity parameters: LD_{50} – Lethal Dose, 50%, LC_{50} – Lethal Concentration, 50%, TDLo – Toxic Dose Low, Lowest published toxic dose. ** Abbreviations used:i.p. – intraperitoneal, i.v. – intravenously, s.c. – subcutaneously, p.c. – percutaneously, p.o. – perorally.

Genotoxicity

Lajmanovich *et al.*¹¹² used the micronucleus test in erythrocytes of *Hyla pulchella* tadpoles as an experimental model for detecting genotoxic effects of endosulfan. The frequency of micronuclei was examined in blood smears obtained from tadpoles exposed *in vivo* to three different concentrations (2.5, 5, and 10 µg L⁻¹) and fixed at two sampling times (48 and 96 h). As a positive control larvae were exposed to 40 mg L⁻¹ of cyclophosphamide. Results obtained in this study demonstrated the genotoxic effects of endosulfan.

Carcinogenicity

As evidences on the carcinogenicity of endosulfan to animals are regarded as inconclusive, it has been reported that this chemical has no carcinogenic potential for animals. However, increases in several kinds of tumors in rats and mices, have already been reported. 55,114,115 α -Endosulfan was also reported as a tumour promoter causing a significant and dose-related increase in hepatocytes, 55,116 and to rapidly inhibit gap junctional intercellular communication (GJIC) in liver cells. 55,117,118

Teratogenicity

A relationship has been observed between maternal exposure and fetal malformations in the skull, ribs and spine of rats. 100,119 Similar teratogenic effects, as well as accumulation of cerebrospinal fluid in the brain, underdeveloped cerebrum, incomplete ossification of skull bones, and malformations of the liver, kidneys, ribs and renal pelvis were observed. 120

Reproductive toxicity

Detailed studies in adult rats exposed to endosulfan for 5 days *per* week for 10 weeks showed reduced intratesticular spermatid counts, sperm abnormalities, and changes in the marker enzymes of testicular activities, providing further evidence of effects on

spermatogenesis. ¹²¹ A study by Dalsenter *et al.* ¹²² indicates that pre and postnatal exposure to low doses of endosulfan (0.5 and 1.5 mg kg⁻¹) does not induce significant adverse effects in the reproductive system of male off-spring Wistar rats at adulthood.

CONCLUSION

The chemical, biochemical, environmental, and toxicological data of endosulfan reviewed in this paper show its toxicological potential risk to human health. In 2010, the *Persistent Organic Pollutants Review Committee* (POPRC) nominated endosulfan to be added to the Stockholm Convention in April 2011, which should finally result in a global ban. ¹²³ In Brazil the comercialization and use of endosulfan is already banned since July 31st 2013, according to the resolution RDC no 28 of August 9th 2010.

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REFERENCES

- 1. Maier-Bode, H.; Residue Rev. 1968, 22, 1.
- Naqvi, S. M.; Vaishnavi, C.; Comp. Biochem. Physiol., C: Comp. Pharmacol. 1993, 105, 347.
- Sharma, R. K.; Kaul, A.; Gupta, A.; Bhadauria, D.; Prasad, N.; Jain, A.; Gurjar, M.; Rao, B. P.; *Indian J. Pharmacol.* 2011, 43, 469.
- Becker, L.; Scheringer, M.; Schenker, U.; Hungerbühler, K.; Environ. Pollut. (Oxford, U. K.) 2011, 159, 1737.
- Brandt, V. A.; Moon, S.; Ehlers, J.; Metehner, M. M.; Struttmann, T.; Am. J. Ind. Med. 2001, 39, 643.
- 6. Demeter, J.; Heyndrickx, A.; J. Anal. Toxicol. 1978, 2, 68.
- Kucuker, H.; Sahin, O.; Yavuz, Y.; Yürümez, Y.; Basic Clin. Pharmacol. Toxicol. 2009, 104, 49.
- 8. Moon, J. M.; Chun, B. J.; Hum. Exp. Toxicol. 2009, 28, 309.

993

- 9. Moses, V.; Peter, J. V.; Clin. Toxicol. (Philadelphia, Pa.) 2010, 48, 539.
- Parbhu, B.; Rodgers, G.; Sullivan, J. E.; Clin. Toxicol. (Philadelphia, Pa.) 2009, 47, 899.
- 11. Satar, S.; Sebe, A.; Alpay, N. R.; Bratisl. Lek. Listy 2009, 110, 301.
- 12. Terziev, G.; Dimitrova, N.; Rusev, F.; *Folia Med. (Plovdiv)* **1974**, *16*, 325
- Wesseling, C.; Corriols, M.; Bravo, V.; Toxicol. Appl. Pharmacol. 2005, 207(2 Suppl), 697.
- Saiyed, H.; Dewan, A.; Bhatnagar, V.; Shenoy, U.; Shenoy, R.; Rajmohan, H.; Patel, K.; Kashyap, R.; Kulkarni, P.; Rajan, B.; Lakkad, B.; Environ. Health Perspect. 2003, 111, 1958.
- Dewan, A.; Bhatnagar, V. K.; Mathur, M. L.; Chakma, T.; Kashyap, R.; Sadhu, H. G.; Sinha, S. N.; Saiyed, H. N.; *J. Toxicol., Clin. Toxicol.* 2004, 42, 363.
- 16. French, H.; Goebel, H.; US Pat. 2,799,685, 1957.
- http://chem.sis.nlm.nih.gov/chemidplus/rn/115-29-7 Accessed in March 25th 2016
- Schmidt, W. F.; Bilboulian, S.; Rice, C. P.; Fettinger, J. C.; McConnell,
 L. L.; Hapeman, C. J.; J. Agric. Food Chem. 2001, 49, 5372.
- Schmidt, W. F.; Hapeman, C. J.; Fettinger, J. C.; Rice, C. P.; J. Agric. Food Chem. 1997, 45, 1023.
- Sutherland, T. D.; Horne, I.; Weir, K. M.; Russell, R. J.; Oakeshott, J. G.; Rev. Environ. Contam. Toxicol. 2004, 183, 99.
- 21. Keith, L.; Telliard, W.; Environ. Sci. Technol. 1979, 13, 416.
- Verschueren, K.; Handbook of Environmental Data on Organic Chemicals, 2nd ed., Van Nostrand Reinhold Co.: New York, 1983.
- 23. Ernst, W. R.; Jonah, P.; Doe, K.; Julien, G.; Hennigar, P.; *Environ. Toxicol. Chem.* **1991**, *10*, 193.
- 24. Petri, D.; Glover, C. N.; Ylving, S.; Kolås, K.; Fremmersvik, G.; Waagbø, R.; Berntssen, M. H.; *Aquat. Toxicol.* **2006**, *80*, 207.
- Barcelo-Quintal, M. H.; Cebada-Ricalde, M. C.; Trejo-Irigoyen, A. R.; Rendon-Osorio, R. B.; Manzanilla-Cano, J. A.; *J. Environ. Sci. Health, Part B* 2008, 43, 120.
- 26. Singh, S. K.; Pandey, R. S.; Indian J. Exp. Biol. 1990, 28, 953.
- Sinha, N.; Narayan, R.; Saxena, D. K.; Bull. Environ. Contam. Toxicol. 1997, 58, 79.
- Sinha, N.; Adhikari, N.; Saxena, D. K.; *Environ. Toxicol. Pharmacol.* 2001, 10, 29.
- 29. Turner, K. H.; Sharpe, R. M.; Rev. Reprod. 1997, 2, 69.
- 30. Chaudhuri, K.; Selvaraj, S.; Pal, A. K.; Mutat. Res. 1999, 439, 63.
- Paul, V.; Balasubramaniam, E.; Environ. Toxicol. Pharmacol. 1997, 3, 151
- Walse, S. S.; Shimizu, K. D.; Ferry, J. L.; Environ. Sci. Technol. 2002, 36, 4846.
- Lee, H. K.; Moon, J. K.; Chang, C. H.; Choi, H.; Park, H. W.; Park, B. S.; Lee, H. S.; Hwang, E. C.; Lee, Y. D.; Liu, K. H.; Kim, J. H.; *Drug Metab. Dispos.* 2006, 34, 1090.
- 34. Awasthi, N.; Ahuja, R.; Kumar, A.; Soil Biol. Biochem. 2000, 32, 1697.
- 35. Walse, S. S.; Scott, G. I.; Ferry, J. L.; J. Environ. Monit. 2003, 5, 373.
- Sutherland, T. D.; Horne, I.; Lacey, M. J.; Appl. Environ. Microbiol. 2000, 66, 2822.
- Awasthi, N.; Manickam, N. N.; Kumar, A.; *Bull. Environ. Contam. Toxicol.* 1997, 59, 928.
- 38. Martens, R. Appl. Environ. Microbiol. 1976, 31, 853.
- 39. Miles, J. R. E.; Moy, P.; Bull. Environ. Contam. Toxicol. 1979, 23, 13.
- Kshemkalyani, S. B.; Vasudevan, P.; Patki, A. H.; Naik, R. B.; Rahalkar,
 S. B.; Francis, R. P.; Patel, G. S.; *Indian J. Environ. Health* 1987, 29,
- 41. Katayama, A.; Matsumura, F.; Environ. Toxicol. Chem. 1993, 12, 1059.
- 42. Kullman, S. W.; Matsumura, F.; Appl. Environ. Microbiol. 1996, 62, 593.
- Beck, E. W.; Johnson, J. C.; Woodham, D. B.; Leuck, D. B.; Dawsey, L. H.; Robbins, J. E.; Bowman, M. C.; *J. Econ. Entomol.* 1966, 59, 1444.
- 44. Dorough, H. W.; Huhtanen, K.; Marshall, T. C.; Bryant, H. E.; Pestic.

- Biochem. Physiol. 1978, 8, 241.
- Trajkovska, S.; Mbaye, M.; Gaye, S. M. D.; Aaron, J. J.; Chevreuil, M.;
 Blanchoud, H.; Anal. Bioanal. Chem. 2009, 394, 1099.
- Stoker, C.; Repetti, M. R.; García, S. R.; Zayas, M. A.; Galoppo, G. H.; Beldoménico, H. R.; Luque, E. H.; Muñoz-de-Toro, M.; *Chemosphere* 2011, 84, 311.
- 47. Wang, X.; Kennedy, K.; Powell, J.; Keywood, M.; Gillett, R.; Thai, P.; Bridgen, P.; Broomhall, S.; Paxman, C.; Wania, F.; Mueller, J. F.; *Environ. Sci.: Process Impacts* **2015**, *17*, 525.
- 48. Zhang, X.; Meyer, T.; Muir, D. C.; Teixeira, C.; Wang, X.; Wania, F.; Environ. Sci.: Process Impacts 2013, 15, 2304.
- Potter, T. L.; Hapeman, C. J.; McConnell, L. L.; Harman-Fetcho, J. A.; Schmidt, W. F.; Rice, C. P.; Schaffer, B.; Sci. Total Environ. 2014, 468, 505
- Kong, L.; Zhu, S.; Zhu, L.; Xie, H.; Su, K.; Yan, T.; Wang, J.; Wang, F.; Sun, F.; J. Environ. Sci. (China) 2013, 25, 2257.
- Varayoud, J.; Monje, L.; Bernhardt, T.; Muñoz-de-Toro, M.; Luque, E. H.; Ramos, J. G.; Reprod. Toxicol. 2008, 26, 138.
- Narayan, S.; Dani, H. M.; Misra, U. K.; J. Biochem. Toxicol. 1989, 4, 205.
- Narayan, S.; Dani, H. M.; Misra, U. K.; J. Environ. Sci. Health, Part B 1990, 25, 243.
- Narayan, S.; Dani, H. M.; Misra, U. K.; J. Environ. Sci. Health, Part B 1990, 25 259
- Rousseau, J.; Cossette, L.; Grenier, S.; Martinoli, M. G.; Gen. Comp. Endocrinol. 2002, 126, 175.
- 56. Ayub, S.; Verma, J.; Das, N.; Int. Immunopharmacol. 2003, 3, 1819.
- Pal, R.; Ahmed, T.; Kumar, V.; Suke, S. G.; Ray, A.; Banerjee, B. D.;
 Indian J. Exp. Biol. 2009, 47, 723.
- Ahmed, T.; Tripathi, A. K.; Ahmed, R. S.; Das, S.; Suke, S. G.; Pathak, R.; Chakraboti, A.; Banerjee, B. D.; *J. Biochem. Mol. Toxicol.* 2008, 22, 299.
- Choudhary, N.; Sharma, M.; Verma, P.; Joshi, S. C.; J. Environ. Biol. 2003, 24, 305.
- Uboh, F. E.; Asuquo, E. N.; Eteng, M. U.; Toxicol. Ind. Health 2011, 27, 483
- 61. Ozmen, O.; Sahinduran, S.; Mor, F.; Pancreas 2010, 39, 367.
- 62. Ozdem, S.; Nacitarhan, C.; Gulay, M. S.; Hatipoglu, F. S.; Ozdem, S. S.; Toxicol. Ind. Health 2011, 27, 437.
- Silva de Assis, H. C.; Nicaretta, L.; Marques, M. C.; Crestani, S.; Soares, K. C.; Olmedo, D.; Dalsenter, P. R. Bull. Environ. Contam. Toxicol. 2011, 86, 368.
- 64. Conacher, H. B. S.; Mes, J.; Food Addit. Contam. 1993, 10, 5.
- 65. Silva, M. H.; Carr, W. C.; Regul. Toxicol. Pharmacol. 2010, 56, 18.
- Wang, Y.; Guo, S.; Xue, R.; Qi, S.; Xu, Y.; Xue, B.; Yuan, D.; Environ. Monit. Assess. 2011, 180, 489.
- Pathak, R.; Suke, S. G.; Ahmed, R. S.; Tripathi, A. K.; Guleria, K.;
 Sharma, C. S.; Makhijani, S. D.; Mishra, M.; Banerjee, B. D.; Bull.
 Environ. Contam. Toxicol. 2008, 81, 216.
- Blanco-Coronado, J. L.; Repetto, M.; Ginestal, R. J.; Vicente, J. R.;
 Yelamos, F.; Lardelli, A.; Clin. Toxicol. 1992, 30, 575.
- Karatas, A. D.; Aygun, D.; Baydin, A.; Singapore Med. J. 2006, 47, 1030.
- 70. Sood, A. K.; Yadav, S. P.; Sood, S.; Indian J. Med. Sci. 1994, 48, 68.
- Spencer, P. S.; Schaumburg, H. H.; Chlorinated cyclodienes. In: Spencer,
 P. S., Scheumburg, H. H., Ludolph, A. C., eds.; Oxford University Press: Oxford, 2000.
- 72. Dalvie, M. A.; Africa, A.; Solomons, A.; London, L.; Brouwer, D.; Kromhout, H.; *J. Environ. Sci. Health, Part B* **2009**, *44*, 271.
- Ely, T. D.; MacFarlane, J. W.; Galen, W. P.; Hine, C. H.; *J. Occup. Med.* 1967, 9, 35.
- Scremin, O. U.; Chialvo, D. R.; Lavarello, S.; Berra, H. H.; Lucero, M. A.; Neurotoxicology 2011, 32, 31.

- 75. Aleksandrowicz, D. R.; Arch. Toxicol. 1979, 43, 65.
- Shemesh, Y.; Bourvine, A.; Gold, D.; Bracha, P.; J. Toxicol., Clin. Toxicol. 1988, 26, 265.
- 77. Boereboom, F. T.; van Dijk, A.; van Zoonen, P.; Meulenbelt, J.; *J. Toxicol.*, Clin. Toxicol. 1998, 36, 345.
- 78. Lo, R. S. K.; Chan, J. C. N.; Cockram, C. S.; Lai, F. M. M.; Clin. Toxicol. 1995, 33, 67.
- Galatone, V.; Annex E of the Stockholm Convention pursuant to Article 8 of the Convention, Stockolm, Sweden, 2009.
- 80. Sang S, Petrovic S. Endosulfan A Review of its toxicity and its effects on the endocrine system, WWF: Canada, 1999.
- Lebailly, P.; Vigreux, C.; Lechevrel, C.; Ledemeney, D.; Godard, T.;
 Sichel, F.; LeTalaër, J. Y.; Henry-Amar, M.; Gauduchon, P.; Cancer Epidemiol., Biomarkers Prev. 1998, 7, 929.
- 82. Venegas, W.; Zapata, I.; Carbonell, E.; Marcos, R.; *Teratog.*, *Carcinog.*, *Mutagen.* **1998**, *18*, 123.
- Falck, G. C. M.; Hirvonen, A.; Scarpato, R.; Saarikoski, S. T.; Migliore, L.; Norppa, H.; Mutat. Res., Fundam. Mol. Mech. Mutagen. 1999, 441, 225
- Scarpato, R; Landini, E.; Migliore, L.; Mutat. Res., Fundam. Mol. Mech. Mutagen. 1996, 372, 195.
- Scarpato, R.; Migliore, L.; Angotzi, G.; Fedi, A.; Miligi, L.; Loprieno,
 N.; Mutat. Res., Genet. Toxicol. Environ. Mutagen. 1996, 367, 73.
- Antherieu, S.; Ledirac, N.; Luzy, A. P.; Lenormand, P.; Caron, J. C.; Rahmani, R.; J. Cell. Physiol. 2007, 213, 177.
- IPCS. 2000. Poisons Information Monograph 576 Endosulfan. International Programme On Chemical Safety, World Health Organisation, Geneva. http://www.inchem.org/documents/pims/chemical/pim576.htm Accessed in Feb 14th 2016.
- Soto, A. M.; Chung, K. L.; Sonnen, S. C.; Environ. Health Perspect. 1994, 102, 380.
- 89. Preziosi, P.; Pure Appl. Chem. 1998, 70, 1617.
- 90. Zhu, Z.; Edwards, R. J.; Boobis, A. R.; Toxicol. Lett. 2008, 181, 93.
- 91. Li, X. R.; Zhang, S.; Safe, S.; J. Steroid Biochem. Mol. Biol. 2006, 98,
- Bonefeld-Jorgensen, E. C.; Grunfeld, H. T.; Gjermandsen, I. M.; Mol. Cell. Endocr. 2005, 244, 20.
- Ibarluzea, J. J.; Fernandez, M. F.; Santa Marina, L.; Olea, S. M. F.; Rivas, A. M.; Aurrekoetxea, J. J.; Enposito, J.; Lorenzo, M.; Torne, P.; Villalobos, M.; Pedraza, V.; Sasco, A. J.; Olea, N.; Cancer, Causes Control, Pap. Symp. 2004, 15, 591.
- 94. Rupa, D.; Reddy, P.; Reddi, O.; Environ. Res. 1991, 55, 123.
- ATSDR. 2000; Toxicological Profile for Endosulfan, Agency of Toxic Substances and Disease Registry, Atlanta, USA. http://www.atsdr.cdc. gov/toxprofiles/tp41.html, Acessed in February 2016.
- Lemaire, G.; Balaguer, P.; Michel, S.; Rahmani, R.; Toxicol. Appld. Pharmacol. 2005, 202, 38.

- Gupta, P. K.; Chandra, S. V.; Bull. Environ. Contam. Toxicol. 1975, 14, 513
- Gupta, P. K.; Chandra, S. V.; Bull. Environ. Contam. Toxicol. 1977, 18, 378.
- 99. Gupta, P. K.; Bull. Environ. Contam. Toxicol. 1976, 15, 708.
- 100. Gupta, P. K.; Murthy, R. C.; Chandra, S. V.; Toxicol. Lett. 1981, 7, 221.
- 101. Miller, L. L.; Gefell, D.; Avallone, A.; Llados, F.; Toxicological profile for endosulfan. U.S. Department of Health and Human Services, Agency for Toxic Substances and Disease Registry: Atlanta, 2000.
- 102. Izmerov, N. F.; Sanotsky, I. V.; Sidorov, K. K.; GKNT Moscow 1982, 72.
- Melnikov, N. N.; Chemistry of Pesticides, 1st ed., Springer-Verlag: New York, 1971.
- 104. Agricultural Research Service, USDA Information Memorandum, 1966,
- 105. Dzwonkowska, A.; Hübner, H.; Arch. Toxicol. 1986, 58, 152.
- Truhaut, R.; Gak, J. C.; Graillot, C.; Eur. J. Toxicol. Environ. Hyg. 1974,
 159.
- 107. Perkow, W.; Wirksubstanzen der Pflanzenschutz und Schadlingsbekampfungsmittel, Verlag Paul Parey: Berlin, 1971-1976.
- 108. Schafer, E. W.; Toxicol. Appl. Pharmacol. 1972, 21, 315.
- 109. Grimmett, W. G.; Dzendolet, I.; Whyte, I.; J. Toxicol., Clin. Toxicol. 1996, 34, 447.
- 110. Doman, I.; Magy. Allatorv. Lapja 1971, 26, 342.
- 111. Utklev, H. E.; Westbye, C.; Nor. Veterinaertidsskr. 1971, 83, 31.
- 112. Lajmanovich, R. C.; Cabagna, M.; Peltzer, P. M.; Stringhini, G. A.; Attademo, A. M.; Mutat. Res. 2005, 587, 67.
- 113. Hack, R.; Ebert, E.; Leist, K. H.; Food Chem. Toxicol. 1995, 33, 941.
- 114. Reuber, M. D.; Sci. Total Environ. 1981, 20, 23.
- 115. Ferdousi, Z.; Islam, M. S.; Khan, M. Z. H. J. Korean Soc. Appl. Biol. Chem. 2008, 51, 294.
- 116. Fransson-Steen, R.; Flodstrom, S.; Warngard, L.; Carcinogenesis 1992, 13, 2299.
- 117. Dubois, M.; Pfohl-Leszkowicz, A.; De Waziers, I.; Kremers, P.; Environ. Toxicol. Pharmacol. 1996, 1, 249.
- 118. Warngard, I.; Bager, Y.; Kato, Y.;, Kenne, K.; Ahlborg, U. G.; Arch. Toxicol. Suppl. 1996, 18, 149.
- 119. Singh, N.; Sharma, A.; Dwivedi, P.; Patil, R.; Kumar, M.; J. Appl. Toxicol. 2007, 27, 143.
- 120. Singh, N. D.; Sharma, A. K.; Dwivedi, P.; Patil, R. D.; Kumar, M.; J. Appl. Toxicol. 2006, 27, 589.
- 121. Khan, P. K.; Sinha, S. P.; Mutagenesis 1996, 11, 33.
- 122. Dalsenter, P. R.; de Araújo, S. L.; de Assis, H. C.; Andrade, A. J.; Dallegrave, E.; *Hum. Exp. Toxicol.* **2003**, *22*, 171.
- 123. Lubick, N.; Science 2010, 328, 1416.