



AURÉLIO DE JESUS RODRIGUES PAIS

**ANTHROPOGENIC DISTURBANCES AND ECOLOGICAL
MODIFICATION ON ATLANTIC FOREST TREE
COMMUNITY**

**LAVRAS – MG
2020**

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Tese apresentada à Universidade Federal de Lavras, como parte das exigências do Programa de Pós-Graduação em Engenharia Florestal, área de concentração em Ecologia Florestal, para obtenção do título de Doutor.

Prof. Dr. Rubens Manoel Dos Santos
Orientador

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COMUNIDADES ARBÓREAS DO DOMÍNIO ATLÂNTICO**

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**LAVRAS – MG
2020**

*À minha Esposa Maria Luís Bracho
Aos meus filhos: Wesley Pais e Welmer Pais
Dedico*

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RESUMO GERAL

Os distúrbios em florestas tropicais constituem um mecanismo fundamental para a dinâmica das comunidades arbóreas e manutenção de processos ecológicos no ecossistema. Porém quando estes são de natureza antropogênica, podem afetar negativamente a biodiversidade na sua estrutura, composição e funcionalidade. O presente estudo procurou avaliar os efeitos do histórico de distúrbios antropogênicos, suas relações com fatores abióticos na estruturação de comunidade arbóreas do Domínio Atlântico. Para tal foram testadas duas hipóteses, nomeadamente: (i) os distúrbios antropogênicos passados alteram a funcionalidade das comunidades, sendo esperado que o aumento da sua intensidade diminuiria a estocagem de carbono, a capacidade de dispersão de sementes e a dinâmica do processo sucessional; e (ii) o histórico de distúrbios antropogênicos afetam a composição florística no tempo, sendo esperado que áreas mais impactadas tenham alteração na composição num gradiente temporal. O levantamento da comunidade arbórea consistiu numa amostragem em 12 fragmentos florestais do sul do estado de Minas Gerais, em 544 parcelas, onde foram medidos o diâmetro a altura do peito (DAP) com inclusão a partir de 5.0 cm para a vegetação, variáveis texturais e químicas para o solo e 19 variáveis bioclimáticas do banco de dados *Worldclim* 1.4 para a caracterização climática da mesorregião. Foi também obtida a caracterização dos impactos antropogênicos pretéritos feito na região por Pereira (2003) e usando a mesma metodologia foi também feita outra caracterização por este estudo para o ano de 2019. Posteriormente, foram criados modelos lineares generalizados mistos (GLMMs) de modo a avaliar se os padrões ecológicos da comunidade respondem aos efeitos do distúrbio, com isto, o fragmento foi utilizado como fator aleatório para lidar com a pseudorreplicação. Os resultados encontraram um total de 36638 indivíduos de 550 espécies representando 76 famílias nos 12 fragmentos amostrados. Os resultados demonstraram que os distúrbios influenciaram significativamente as síndromes de dispersão de zoocoria e autocoria nos modelos, sendo que quanto maior a sua intensidade, maior a abundância de espécies autocóricas e menor das espécies zoocóricas. A produtividade respondeu de forma inversa ao distúrbio antropogênico, sendo que o aumento do distúrbio antropogênico implicou na redução da produtividade. A média ponderada da comunidade para a densidade da madeira (CWMWD) respondeu de forma significativa, sendo que o aumento do distúrbio antropogênico, também implicou no seu aumento. Por outro lado, o distúrbio antropogênico e as variáveis edáficas não tiveram efeito significativo sobre a biomassa aérea. O solo foi um preditor importante com efeito na diversidade beta temporal, na CWMWD, e na autocoria diferentemente do clima que não mostrou algum efeito nos padrões ecológicos. Portanto, nossos resultados demonstram que, de fato, os distúrbios antropogênicos podem ser um preditor importante na modificação de padrões ecológicos das comunidades arbóreas, podendo afetar serviços ecossistêmicos de que a humanidade depende.

Palavras-chave: Distúrbios antropogênicos. Fatores abióticos. Modificações ecológicas. Comunidades arbóreas.

ABSTRACT

The disturbances in tropical forests are a fundamental mechanism for tree community dynamics and the maintenance of ecosystem ecological processes. However, when these are from anthropogenic origin, they can negatively affect biodiversity on its structure, composition and functionality. The present study sought to evaluate the history effects of anthropogenic disturbances, their relationship with abiotic factors in the structuring Atlantic domain tree communities. For this purpose, two hypotheses were tested, namely: (i) the history of anthropogenic disturbances affect the floristic composition over time, and it is expected that most impacted areas will change in the temporal gradient composition, and (ii) past anthropogenic disturbances modify the communities' functionality, and increasing its intensity would decrease carbon uptake, seed dispersal capacity and successional dynamics. Tree community survey consisted of 12 forest fragments sampling in the south of Minas Gerais state, making up 544 plots, where the diameter at breast height (DBH) was measured from 5.0 cm for vegetation inclusion; textural and chemical variables for the soil and 19 bioclimatic variables for mesoregion climatic characterization from the Worldclim 1.4 database. The past anthropogenic impacts characterization in the region made by Pereira (2003) was also obtained, and using the same methodology, another characterization was also made by this study for the year 2019. Subsequently, mixed generalized linear models (GLMMs) were created in order to evaluate if the communities' ecological variables respond to the disturbances effects, with this, the fragment was used as a random factor to deal with pseudoreplication. A total of 36638 individuals from 550 species representing 76 families were found in the 12 fragments sampled. The results showed that anthropogenic disturbances significantly influenced the zoochory and autochory dispersion syndromes in the models, where its greater intensity results in greater autochorous species abundance and lower zoochoric species abundance. Productivity responded inversely to anthropogenic disturbances, thus its increase implied a productivity decrease. The community-weighted mean of wood density (CWMWD) responded significantly, and the anthropogenic disturbances increasing impact implied its increase. On the other hand, the anthropogenic disturbances and soil variables had no significant effect on aboveground biomass. Soil was an important driver of beta temporal diversity, CWMWD, and autochory, unlike the climate that did not show any effect on ecological variables. Therefore, our results clearly demonstrate that, in fact, anthropogenic disturbances can be an important driver in modifying the tree communities ecological parameters, and can affect ecosystem services upon which humanity depends.

Keywords: Anthropogenic disturbances. Abiotic factors. Ecological modifications. Tree communities.

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PRIMEIRA PARTE

1 INTRODUÇÃO

Desde a sua existência, o homem sempre usou os recursos naturais, em particular a biodiversidade, para sua sobrevivência. Porém, com o crescimento populacional e consequente aumento da demanda por recursos naturais, surgiu a industrialização como forma de aumento da produção e com isto a obtenção de maiores ganhos econômicos, o que levou a uma elevada sobre-exploração destes recursos. Esta pressão antropogênica tem levado à destruição, à degradação e à fragmentação dos habitats com reflexo nas mudanças da distribuição e abundância dos organismos. É fato que a biodiversidade está sendo perdida a uma taxa sem precedentes a nível global, como consequência do uso da terra, pondo em risco a integridade dos ecossistemas e seus serviços de que os seres humanos dependem para sua sobrevivência. Atualmente estima-se que a população humana seja em torno de sete bilhões de habitantes e com isto, promovendo uma perda de biodiversidade sem precedentes históricos. Tanto que os distúrbios antropogênicos são considerados como uma das principais causa de perda da biodiversidade global.

É importante que os seres humanos percebam como suas ações afetam a biodiversidade e a importância da sua conservação, pois sem ela não haveria existência humana. Portanto, compreender como as espécies e suas interações são afetadas por ações antropogênicas é crucial para a conservação das funções e serviços do ecossistema. Por outro lado, se não houver ações concretas em relação ao uso racional da biodiversidade, poderá haver uma degradação dos recursos dela provenientes, comprometendo a vida humana no planeta. A Mata Atlântica, um dos principais *hotspots* mundiais de biodiversidade não está fora desta situação. Atualmente tem a sua área reduzida entre 12 a 16% da sua extensão original e a maioria em pequenos fragmentos. O uso de terra humano, em particular a expansão agrícola, a urbanização e a sobre-exploração da biodiversidade figuram entre as principais causas da redução da extensão do Domínio Atlântico, o que resulta em elevada perda de espécies, degradação e perda de habitats.

O Laboratório de Fitogeografia e Ecologia Evolutiva, pertencente ao departamento de Ciências Florestais da Universidade Federal de Lavras liderado pelo Prof. Dr. Rubens Manoel dos Santos, vem desenvolvendo pesquisas em ecologia e dinâmica de comunidades e sobretudo como os fatores ambientais e antropogênicos influenciam a distribuição das espécies de árvores e a funcionalidade da comunidade. Com isto, surgiu o interesse em se responder à seguinte pergunta: como os fatores ambientais (solo e clima) e distúrbios antropogênicos influenciam a composição e a funcionalidade de comunidades arbóreas? Este

trabalho é fruto de um banco de dados composto por 12 fragmentos de Mata Atlântica, com vários anos de inventários de dinâmica de comunidades arbóreas (desde o ano de 2003 até 2019). Adicionalmente, recorreu-se a um trabalho feito por Pereira (2003). Em uma matriz para as mesmas áreas, este autor classificou o impacto dos distúrbios humanos sobre variáveis estruturais. Com isto, fizemos modelos da dinâmica de biomassa (carbono) para estes fragmentos, associando-a aos fatores ambientais e à matriz de impactos antropogênicos elaborada por Pereira (2003). Esta associação nos permitiu inferir sobre a variação da funcionalidade da comunidade, em particular nos atributos de produtividade, síndromes de dispersão e densidade da madeira.

A presente tese foi estruturada em duas partes, na qual a primeira parte apresenta uma abordagem de revisão bibliográfica contendo conceitos, contribuições relevantes e lacunas existentes sobre o tema. A segunda parte corresponde à apresentação de um artigo científico formatado e redigido de acordo com as normas do periódico *Conservation Biology*.

2 REFERENCIAL TEÓRICO

2.1 Fatores abióticos e estruturação de comunidades arbóreas

Os fatores abióticos são fortes propulsores dos processos demográficos nas comunidades arbóreas, pois determinam a disponibilidade de recursos para o crescimento e a sobrevivência das plantas (CONDIT et al., 2013; FELDPAUSCH et al., 2011). Nos Neotrópicos, a composição das características reflete as diferenças de clima e fertilidade do solo (VAN DER SANDE et al., 2017). Em uma escala mais local, nas florestas africanas, os solos arenosos em comparação com os solos argilosos, apresentam maior abundância de espécies com alta densidade da madeira e mais tolerantes à seca. Não só, como também se adaptam melhor a ambientes com recursos bastante limitados (FAYOLLE et al., 2012; VAN DER SANDE et al., 2017). Os recursos naturais tais como a disponibilidade de água, nutrientes e luz são vitais para o crescimento das plantas (TOLEDO et al., 2011). Nas florestas tropicais, esses recursos variam em escalas espaciais e temporais e, como resultado, o crescimento das árvores varia com a disponibilidade de recursos (FELDPAUSCH et al., 2011; QUESADA et al., 2012). Geralmente, o crescimento de plantas aumenta com a precipitação e diminui com o aumento da sazonalidade (BERNARDI et al., 2016). Portanto, nestas formações o clima é tido como um dos principais preditores de produtividade das plantas (TOLEDO et al., 2011; WAGNER et al., 2012).

Independentemente da precipitação, a fertilidade do solo também pode afetar a taxa de crescimento das árvores, com melhor crescimento em solos mais ricos em nutrientes (PRADO-JUNIOR et al., 2016; VAN DER SANDE et al., 2017; VAN DER SANDE et al., 2018). Os ecossistemas tropicais são amplamente considerados como ricos em nitrogênio e relativamente pobres em nutrientes fortemente intemperizados, como é o caso do fósforo (PRADO-JUNIOR et al. 2016; QUESADA et al., 2012). Nas florestas amazônicas, o teor de fósforo no solo é considerado o melhor preditor das taxas de crescimento destas florestas (QUESADA et al., 2012). Outros macronutrientes do solo tais como cálcio, potássio e magnésio também demonstraram ser importantes na determinação das taxas de crescimento e produtividade das comunidades arbóreas de florestas tropicais (BARIBAULT et al., 2012; WRIGHT et al., 2011). Portanto, as florestas em solos mais férteis tendem a ter maiores taxas de crescimento e maior produtividade lenhosa, se comparadas às de solos pouco férteis (HEDIN et al., 2009, TOLEDO et al., 2011).

2.2 Fragmentação florestal

A fragmentação e perda de habitats são os principais processos de ameaça à biodiversidade tropical, levando à extinção de espécies em várias escalas e afetando negativamente o funcionamento do ecossistema (HADDAD et al., 2015; LIU et al., 2018). É um dos principais processos que contribuem para o declínio da população, perda de biodiversidade e alteração da estrutura da comunidade e do funcionamento do ecossistema (IBANEZ et al., 2017; TAPIA-ARMIJOS et al., 2015). É frequentemente definida como um processo durante o qual uma grande extensão de habitat é transformada em um número maior de manchas menores, isolados uns dos outros por uma matriz diferente da original (FAHRIG, 2017). A definição de fragmentação de habitat implica quatro efeitos deste processo no padrão de habitat, nomeadamente: (i) redução na quantidade de habitats, (ii) aumento no número de manchas, (iii) diminuição nos tamanhos de manchas e (iv) aumento no isolamento de manchas. Estes quatro efeitos formam a base da maioria das medidas quantitativas de fragmentação do habitat (FAHRIG, 2017; MARTIN; FAHRIG, 2012; THORNTON; BRANCH; SUNQUIST, 2011).

A fragmentação elimina árvores grandes de elevada densidade de madeira em processos sucessionais tardios (SANTO-SILVA et al., 2016). Estas árvores contribuem significativamente para a biomassa acima do solo e são vitais para o estoque de carbono, microclima, refúgio de animais entre outras funções (TABARELLI et al. 2010). Caso estas estejam nas bordas, sofrem maior efeito da temperatura que pode levar à dessecação da copa e promover elevada abundância de espécies pioneiras invasoras (FAHRIG, 2017; SANTO-SILVA et al., 2016). Quanto menor for o fragmento, mais intenso será este efeito, sendo que as espécies pioneiras e de rápido crescimento tendem a invadir o fragmento, alterando a estrutura da comunidade e o funcionamento do ecossistema (LIU et al., 2018).

Por outro lado, vários usos de terra modificados pelo homem tais como agricultura, plantações florestais, cafezais e pasto, circundam os fragmentos florestais (a matriz) nas florestas tropicais afetando sua estrutura, composição e dinâmica (RUFFELL; CLOUT; DIDHAM, 2017). A matriz pode fornecer recursos complementares que podem compensar a disponibilidade limitada de recursos nas manchas de habitat (HERNÁNDEZ-RUEDAS et al., 2018; RIES et al., 2004). Um aumento na qualidade de matriz foi associado com um aumento na riqueza e abundância de espécies nos fragmentos (OCKINGER et al., 2012; RUFFELL; CLOUT; DIDHAM, 2017). Várias evidências sugerem que a qualidade da matriz é crucialmente importante na determinação da abundância e composição de espécies dentro de

fragmentos de habitat (COOK et al., 2002; HERNÁNDEZ-RUEDAS et al., 2018). Ela pode afetar a dispersão e o movimento de indivíduos entre manchas (DAVIES et al., 2001; OCKINGER et al., 2012), sendo determinantes da dinâmica de extinção e colonização dos fragmentos (BROTONS et al., 2003; KUPFER et al., 2006). Por exemplo, as florestas plantadas são usos de terra muito comuns na matriz dos biomas florestais, e elas podem ajudar a reduzir os impactos do desmatamento, aumentando a conectividade entre os fragmentos, fornecendo habitat e reduzindo o efeito de borda (PAWSON et al. 2010; RUFFELL; CLOUT; DIDHAM, 2017). Contudo, podem também trazer alguns impactos negativos principalmente para comunidades de vertebrados (ESTADES; TEMPLE, 1999). Por estas razões, o estudo da estrutura da matriz é frequentemente visto como uma das vias mais importante para se entender sobre a dinâmica das manchas na paisagem, pois a modificação da qualidade da matriz poderá afetar a dispersão, o tamanho da população e a probabilidade de persistência da população nos fragmentos florestais (DONALD; EVANS, 2006; SMITH; FAHRIG; FRANCIS, 2011).

2.3 Dinâmica de florestas tropicais

As florestas tropicais são repositórios de grande parte da biodiversidade do mundo e desempenham um papel crucial na regulação do clima global (FUJII et al., 2018; LEWIS et al., 2015). Estas formações florestais estão sob enorme pressão do desenvolvimento de atividades humanas, tais como urbanização, agricultura, mineração, entre outros (BROOKS et al., 2002; HADDAD et al. 2015). Identificar e entender as mudanças na composição e estrutura da floresta é fundamental para entender como as florestas estão respondendo à variabilidade e mudanças ambientais (NUNES et al., 2003; ROITMAN et al., 2016). Portanto, entender a dinâmica da composição da comunidade de plantas é fundamental para entender numerosos processos ecológicos em ecossistemas complexos e ricos em espécies, como as florestas tropicais (URIARTE, 2004). Pesquisadores compararam a dinâmica de comunidades de florestas tropicais de diferentes partes do mundo e encontraram taxas altamente variáveis de recrutamento, crescimento e mortalidade, destacando diferentes mecanismos para diferentes sistemas tropicais (FELFILI, 1995; SURESH; DATTARAJA; SUKUMAR, 2010; WOODS, 2000).

A mortalidade, o crescimento e o recrutamento de espécies arbóreas são fatores-chave que influenciam a estrutura, composição e sucessão das comunidades florestais (HURST, 2012; ROITMAN et al., 2016). A mortalidade das árvores é reconhecida como um dos

processos mais importantes na dinâmica florestal, e é influenciada por muitos fatores (BIN et al., 2012; VINCENT et al. 2018). Pode facilitar a rotatividade na composição de espécies, afetar a estrutura da comunidade e alterar as taxas de ciclagem de nutrientes ou acumulação de biomassa (SHEN et al., 2014). Também pode determinar a dinâmica ou sucessão florestal e contribuir para a coexistência de espécies arbóreas (URIARTE, 2004). A probabilidade de mortalidade de uma árvore individual é avaliada em função de seu tamanho, crescimento e o padrão espacial das árvores circundantes, também conhecido como vizinhança competitiva (RUNKLE, 2000; VINCENT et al. 2018).

O crescimento e o recrutamento de comunidades também são processos importantes que moldam a estrutura e a dinâmica da floresta. A transição da germinação para o crescimento da regeneração é um estado crítico no estabelecimento de árvores (CONDIT et al., 1999; ROITMAN et al., 2016). O padrão espacial de recrutamento de plântulas influencia os padrões de distribuição a longo prazo das espécies e pode ter efeitos significativos na composição e abundância de espécies em comunidades vegetais. Portanto, fatores que influenciam o recrutamento de plântulas são de grande importância para ecólogos florestais e pesquisadores (MA et al., 2016; MONTES-HERNANDEZ et al., 2013; SANTOS et al., 2018). Estudos experimentais e observacionais descobriram padrões de recrutamento e mortalidade de espécies arbóreas dependentes de distância e densidade, coerentes com a hipótese de Janzen-Connell em comunidades temperadas e tropicais (HURST, 2012). Estes estudos apontam para a dependência de densidade como uma importante força estabilizadora que promove a coexistência de espécies em sistemas florestais (SHEN, 2014). A limitação de dispersão é outro mecanismo que impulsiona a diversidade de espécies, refere-se ao fenômeno do declínio da densidade de sementes com o aumento da distância em relação à árvore materna (N'DRI et al., 2014; WOODS, 2000). Sementes de plantas mais competitivas podem não chegar a micro-locais adequados e espécies menos competitivas terão mais chances de tomar seus lugares, diminuindo a exclusão competitiva e promovendo a coexistência de espécies (MONTES-HERNANDEZ et al., 2013).

MA et al. (2016) constataram que a competição é o principal fator nas mudanças de longo prazo na mortalidade, no crescimento e no recrutamento de árvores. O clima regional tem um efeito mais fraco, ainda que significativo, sobre a mortalidade, o crescimento e o recrutamento de árvores (HE; DUNCAN, 2000). Isso indica que os processos internos da comunidade, mais do que os fatores climáticos externos, impulsionam a dinâmica das florestas (MA et al., 2016; SANTOS et al., 2018). A dinâmica florestal é de ação lenta e, portanto, exige que os dados de longo prazo sejam caracterizados com precisão. Dados

demográficos, como recrutamento, crescimento e mortalidade, oferecem chaves para entender as mudanças direcionais na composição das comunidades florestais (MONTES-HERNANDEZ et al., 2013; SANTOS et al., 2018).

2.4 Distúrbios antropogênicos nas florestas tropicais

Pickett & White (2013) definem distúrbios como qualquer evento no tempo que perturbe a estrutura do ecossistema, da comunidade ou da população e altera os recursos disponíveis ou o ambiente físico. Nas florestas, a perturbação (distúrbios) varia em intensidade, escala e frequência, a partir de lacunas formadas regularmente pela morte de árvores em agregado ou individuais (HILTNER et al. 2018; POWERS; MARÍN-SPIOTTA, 2017). A perturbação pode dividir-se em duas categorias: origem natural ou origem antropogênica. Distúrbios naturais, como por exemplo os deslizamentos de terra, tempestades e raios, causam a morte de organismos em seus ecossistemas (CRAUSBAY; MARTIN, 2016; MITCHELL, 2013). A morte de um indivíduo cria espaço livre no ecossistema, o que dá oportunidade a novos indivíduos para colonizarem a área (COLE; BHAGWAT; WILLIS, 2014; FOLEY et al. 2005).

As plantas se adaptaram para se recuperar de distúrbios naturais, a exemplo da densidade da madeira, uma característica funcional essencial na comunidade, que se correlaciona fortemente com as estratégias de história de vida das espécies arbóreas (FALSTER, 2006). É um indicador da resistência que demonstra a capacidade de uma árvore de resistir a danos durante distúrbios (CURRAN et al., 2008). No entanto, há uma correlação negativa entre a capacidade de resistência a distúrbios e a capacidade de recuperar a biomassa após um distúrbio (CURRAN et al., 2008; FALSTER, 2006). Tanto que o investimento em biomassa em suporte estrutural resulta em árvores com alta densidade da madeira e baixas taxas de crescimento; enquanto o baixo investimento em suporte estrutural está associado a baixa densidade da madeira e a rápidas taxas de crescimento (KING et al., 2006). No entanto, distúrbios antropogênicos podem resultar em um impacto profundo e prolongado na comunidade (BARLOW et al., 2016; TURNER et al., 2003). Os distúrbios variam em ampla escala espacial e temporal, podendo deixar legados persistentes e interações entre eles, tornando-os difíceis de avaliar (BROWN; BOUTIN, 2009; TURNER et al., 2003).

O uso da terra é uma forma de perturbação humana que cria um tipo diferente de pressão para o ecossistema em relação aos processos naturais de perturbação (DECAENS et al. 2018; MEYFROIDT et al., 2014). Segundo a FAO (2002), a agricultura e a silvicultura

causam a maior pressão humana nos ecossistemas terrestres. A perturbação altera os processos do ecossistema, como estruturas tróficas, fluxo de energia, ciclagem química e outros processos (BARLOW et al., 2016; CONNELL, 1978).

De acordo com Laurance (2008), a conversão de habitats por humanos é um processo não aleatório. Acessibilidade e proximidade de estradas ou assentamentos humanos a uma área natural são questões de alta importância (LAURANCE, 2008, TOIVONEN et al. 2011). Os remanescentes florestais frequentemente persistem em áreas íngremes, em solos mais pobres, em altitudes mais elevadas e em terras parcialmente inundadas (LAURANCE, 2008). A perturbação humana altera o tamanho dos fragmentos e diminui a biodiversidade (WHITTAKER; TRIANTIS; LADLE, 2008).

As florestas tropicais de montanha remanescentes estão sob ameaça direta por causa da conversão para terras agrícolas, extração de madeira e atendimento às necessidades energéticas (ATTA-KRAH; YA, 2000; TOLEDO-ACEVES et al., 2011). Cerca de 12% da população humana global habita áreas montanhosas, a maioria em países em desenvolvimento (HUDDLESTON et al., 2003; MOWO et al., 2007; PRICE; MESSERLI, 2002). Segundo Huddleston et al. (2003), a maioria dos habitantes rurais em zonas montanhosas está ligada à atividade agrícola para sua subsistência. O pastoreio e a silvicultura são usos predominantes das terras de montanha em todas as regiões do mundo. Na América Central e do Sul, as práticas de uso misto da terra (cultivo, pastagem e exploração de recursos florestais) são típicas das comunidades em zonas montanhosas (MOWO et al., 2007).

A criação de gado, especialmente em altitudes mais elevadas nos países em desenvolvimento, é a principal forma de subsistência (BLACKHALL et al., 2008; HUDDLESTON et al., 2003). Esta atividade pode afetar a ecologia de determinada área de várias maneiras, nomeadamente: (i) alterando a riqueza e composição de espécies das comunidades, (ii) perturbando a função do ecossistema e (iii) alterando a estrutura do ecossistema (CHATURVEDI; RAGHUBANSHI; SINGH, 2012). Não só, como também vários autores demonstraram que solos florestais severamente compactados pela criação de gado afetam a regeneração florestal, inibindo a germinação e crescimento de plântulas e aumentando a sua mortalidade (RATOVONAMANA et al. 2013; SCHULZ et al. 2016).

A exploração madeireira contribui significativamente para a degradação florestal, afetando a produtividade, estrutura, biomassa e composição de espécies (FOLEY et al., 2005; TOIVONEN et al., 2011). Esta atividade geralmente recorre ao abate de árvores emergentes para fins econômicos (OLIVEIRA; SANTOS; TABARELLI, 2008). Segundo De Paula (2011), as espécies de árvores emergentes geralmente representam apenas cerca de 10% da

riqueza total de espécies de árvores tropicais, mas têm uma elevada influência sobre a estrutura da floresta e no funcionamento do ecossistema. As árvores emergentes armazenam grande parte da biomassa acima do solo, contribuindo decisivamente para serviços ecológicos, como ciclagem de nutrientes, captação de água, controle de erosão do solo, fornecimento de produtos florestais, entre outros (DE PAULA, 2011; OLIVEIRA; SANTOS; TABARELLI, 2008).

2.5 Síndromes de dispersão e interações ecológicas

As interações planta-animal têm um papel fundamental na manutenção da biodiversidade (VALIENTE-BANUET et al., 2014). Em particular, frugivoria e dispersão de sementes são interações importantes, pois estão envolvidas no estabelecimento de sementes e no recrutamento de plantas (GOMES et al., 2008). Pelo menos 78% das espécies lenhosas em florestas tropicais e temperadas dependem de vetores animais para dispersão primária ou secundária de sementes (AU; CORLETT; HAU, 2006; MULLER-LANDAU et al., 2008). No entanto, a perturbação do habitat afeta as populações de dispersores e as interações de dispersão de sementes, levando à redução do recrutamento de plantas e do tamanho das populações, e aumento do risco de extinção local (FONTÚRBEL et al., 2015; MARKL et al., 2012).

Em paisagens fragmentadas, a dispersão de sementes tem uma grande influência sobre a persistência das espécies de plantas, assim como na recuperação da vegetação quando a perturbação é reduzida (BOVO et al., 2018; CHAVES; BICCA-MARQUES; CHAPMAN, 2018). Poucos tipos de vegetação natural podem persistir em seu estado atual sem a dispersão de sementes mediada por animais (VALIENTE-BANUET et al., 2014). Vários autores propõem três hipóteses principais para explicar por que a dispersão de sementes em geral e em particular por animais constitui um processo crucial no ciclo de vida das plantas (CONNELL, 1971; FONTÚRBEL, 2015; HOWE; SMALLWOOD, 1982; VALIENTE-BANUET et al., 2014). Todas as três hipóteses postulam que a dispersão de sementes mediada por animais melhora o recrutamento de plantas e, mantém a diversidade genética e viabilidade populacional das plantas, a destacar: (i) hipótese de fuga, assume que fungos patogênicos e herbívoros ocorrem em alta densidade próximos às árvores progenitoras, o que aumenta a mortalidade de sementes e plântulas próxima à árvore progenitora (CONNELL, 1971; HOWE; SMALLWOOD, 1982); (ii) hipótese da colonização, propõe que a dispersão de sementes aumenta a probabilidade de as sementes atingirem novos locais onde possam

germinar e se estabelecer (HOWE; SMALLWOOD, 1982; MARKL et al., 2012; VALIENTE-BANUET et al., 2014); e por último (iii) hipótese de dispersão dirigida, sugere que os agentes de dispersão de sementes depositam sementes em locais particularmente adequados para germinação e recrutamento (FONTÚRBEL, 2015; HOWE; SMALLWOOD, 1982; KUHLMANN; RIBEIRO, 2016), portanto as interações planta-frugívoro têm múltiplas implicações evolutivas e na história de vida das plantas (KUHLMANN; RIBEIRO, 2016). Estas interações envolvem benefícios recíprocos em que as plantas aumentam a sua chance de sobrevivência e persistência na paisagem, e os animais fazem o uso de frutos ou sementes carnudas como recursos alimentares (AU; CORLETT; HAU, 2006). Ademais, a dispersão a longa distância por megafauna frugívora também permite a colonização de vários fragmentos florestais isolados e a manutenção da conectividade em paisagens fragmentadas (BOVO et al., 2018; MULLER-LANDAU et al., 2008).

2.6 Síndromes de dispersão e distúrbios antropogênicos

A dispersão de sementes é um mecanismo essencial para a distribuição espacial e expansão da variabilidade genética de populações e comunidades e sua persistência nos ecossistemas (FARWIG; BERENS, 2012; HERRERA; MORALES; GARCÍA, 2011). É o único meio pelo qual as comunidades colonizam novos habitats mais favoráveis ou escapam da competição, patógenos e herbívoros associados às mães (CONNELL, 1971; HOWE; SMALLWOOD, 1982). Sendo esta por meio abiótico (vento, água, entre outros) ou por meio biótico (LEVINE; MURRELL, 2003; PEREDO et al., 2013).

A perturbação humana pode afetar a dispersão de sementes mediada por animais e a regeneração natural de populações de plantas (MARKL et al., 2012). Na matriz, os dispersores diferem em sua capacidade de se mover e na tolerância à degradação do habitat, determinando o impacto final da dispersão das sementes na paisagem (HOLBROOK; LOISELLE, 2009; SEKERCIOGLU; DAILY; EHRLICH; 2004). Portanto, a natureza da matriz circundante dos fragmentos influencia o movimento e a abundância de sementes e frugívoros (MCCONKEY et al., 2012), sendo que a abundância e diversidade de plantas e frugívoros nos fragmentos diminui ao longo de um gradiente de floresta secundária, agrossilvicultura, plantações exóticas e pastagens (FONTURBEL et al., 2015; MCCONKEY et al., 2012). A matriz circundante também influencia os padrões de circulação do vento, afetando sementes anemocóricas em habitats fragmentados (DAMSCHEN et al., 2008; MCCONKEY et al., 2012). Por exemplo, os corredores de habitat podem atuar como quebra-

vento em matrizes abertas e levar à acumulação de sementes dispersas pelo vento (DAMSCHEN et al., 2008). Por outro lado, as matrizes de pastagem podem levar ao isolamento das manchas, uma vez que pastagens abertas não fornecem habitat adequado para o estabelecimento de sementes (PREVEDELLO; VIEIRA, 2010).

A persistência de espécies vegetais em paisagens fragmentadas também é influenciada pelas características dos frutos e sementes e pelos atributos do dispersor (MCCONKEY et al., 2012). Espécies de plantas com sementes maiores, dispersas por animais, geralmente experimentam dispersão reduzida nos fragmentos de habitat em comparação com espécies de frutos menores em paisagens tropicais e temperadas (CRAMER; MESQUITA; WILLIAMSON, 2007). Distúrbios antropogênicos como a caça e a exploração seletiva de madeira, contribuem em grande escala para redução e extinção de dispersores (FONTURBEL et al., 2015), sendo que a caça elimina diretamente o agente mamífero dispersor enquanto a exploração seletiva diminui a disponibilidade de alimento ao dispersor, o que pode levar à sua fuga (PERES; PALACIOS, 2007).

Por outro lado, a perda de frugívoros por ações antropogênicas pode limitar a capacidade das plantas de responderem aos efeitos das mudanças climáticas (CORLETT, 2009), porque a maioria das populações de plantas na Terra precisará percorrer longas distâncias nos próximos 50 a 100 anos para acompanhar as mudanças no clima (CHEN et al., 2011). Estas espécies precisarão se mover no espaço à procura de recursos e condições climáticas adequadas (CHEN et al., 2011; CORLETT, 2009). Contudo, a megafauna (herbívoros de grande porte), capaz de dispersar sementes grandes e a longas distâncias, necessárias para que as plantas mantenham o ritmo das alterações climáticas, é frequentemente ameaçada pela caça ou perda de habitat, podendo isto levar a uma extinção da biodiversidade em macroescala (FONTURBEL et al., 2015; MARKL et al., 2012).

2.7 Serviços ecossistêmicos e sequestro de carbono

As florestas cobrem cerca de um terço da superfície terrestre (4 bilhões de hectares) e desempenham um papel importante no ciclo global do carbono, representando também uma proporção substancial da biodiversidade terrestre do mundo (FAO, 2010; LINDSELL; KLOP, 2013; PARROTTA et al., 2012). Florestas armazenam enormes quantidades de carbono, o que as torna componente vital para mitigar os efeitos das mudanças climáticas globais (PARROTTA et al., 2012). Distúrbios antropogênicos, tais como exploração madeireira, incêndios, fragmentação, caça excessiva, entre outros, contribuem negativamente para a

capacidade das florestas de fornecerem bens e serviços ecossistêmicos (BARLOW et al., 2016; BERENGUER, et al., 2014; LINDSELL; KLOP, 2013). Estes bens e serviços incluem a ciclagem de nutrientes, a formação de solo e produtividade primária; alimentos, água, madeira e medicamentos; serviços de regulação, como controle de erosão, regulação climática, mitigação de inundações, purificação de água e ar, polinização e serviços culturais como recreação, ecoturismo, valores educacionais, entre outros (BERENGUER, et al., 2014; MEA, 2005; PARROTTA et al., 2012). Um dos principais serviços prestados pelas florestas é a remoção de carbono da atmosfera (sequestro) e o armazenamento a longo prazo desse carbono em biomassa, matéria orgânica morta e no solo (PAN et al., 2011). LEWIS et al. (2009) estimaram em medições de longo prazo em parcelas não perturbadas que as florestas tropicais podem sequestrar anualmente cerca de 1,3 Pg de carbono.

Por outro lado, as florestas podem emitir volumes significativos de dióxido de carbono dependendo de como são manejadas, contribuindo para o agravamento dos efeitos das mudanças climáticas globais (ARAGÃO; SHIMABUKURO, 2014). Atualmente, as regiões florestadas contribuem substancialmente para emissão do dióxido de carbono atmosférico, com emissões anuais globais provenientes de desmatamento e degradação florestal estimadas em 1,2 Pg entre 1997 e 2006, equivalentes a 12% das emissões globais provocadas pelo homem (LINDSELL; KLOP, 2013; PAN et al., 2011; VAN DER WERF et al., 2009). A Convenção-Quadro das Nações Unidas sobre Mudanças Climáticas (UNFCCC) em 2007, em sua 13ª Conferência das Partes, reconheceu a degradação florestal como um dos principais fatores para as emissões globais de carbono, sendo este incorporado ao mecanismo de Redução de Emissões por Desmatamento e Degradação Florestal (REDD+) (PARROTTA et al., 2012). Nas florestas tropicais, a maioria das pesquisas ecológicas sobre estoques de carbono tem se concentrado no monitoramento de mudanças em florestas primárias relativamente não perturbadas (BERENGUER, et al., 2014; MALHI et al., 2006; PHILLIPS et al., 2008). Portanto, entender os efeitos das perturbações humanas nos estoques de carbono da floresta é crucial para avaliação de melhores práticas de manejo florestal e medidas de conservação (BERENGUER, et al., 2014).

3 CONSIDERAÇÕES FINAIS

A estrutura e composição atual das comunidades arbóreas são resultados das interações de vários processos de dinâmica florestal com os fatores ambientais, distúrbios naturais e antropogênicos passados. Muitas vezes se usa os distúrbios presentes no estudo de seus efeitos sobre a comunidade, sendo que os históricos de distúrbios passados acabam negligenciados. Neste trabalho, usamos os distúrbios passados para inferir sobre os dados atuais da comunidade, pois, acreditamos que a forma como se apresenta a comunidade arbórea hoje é reflexo dos efeitos dos distúrbios passados e suas interações. Esperamos com este trabalho contribuir com conhecimentos e abrir novos horizontes de pesquisas nesta área, de modo que estas possam ser usadas em atividades de restauração florestal e em políticas de conservação de ecossistemas.

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SEGUNDA PARTE – ARTIGO 1
(a ser submetido no periódico *Conservation Biology*)

1 **Do abiotic factors drive ecological modifications in tropical forest fragments with a**
2 **history of anthropogenic disturbances?**

3
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14
15 **Keywords:** Anthropogenic Disturbances, Tree Community, Abiotic Factors.

16 **Article Impact Statement:** Anthropogenic disturbances can have greater effects than abiotic
17 factors in the tree community structure and ecosystem function modification.

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26 Abstract

27 Abiotic factors are important drivers of tree community structure in tropical forests, but may
28 be minimized or hidden by the effects of anthropogenic disturbances. Here, we aimed to
29 understand how anthropogenic disturbances affect ecological modifications in the tropical tree
30 communities around 12 fragments of the Brazilian Atlantic Domain. We hypothesized that a
31 history of anthropogenic disturbances would alter community temporal beta diversity and
32 function. We found that disturbance intensity significantly influenced the abundance of
33 zoochorous (negative effect) and autochorous (positive effect) species. Disturbance history
34 negatively affected wood productivity in the time interval considered, but positively affected
35 community-weighted means (CWM) of wood density. Conversely, anthropogenic
36 disturbances and soil variables did not significantly affect aboveground woody biomass.
37 Temporal beta diversity, wood density CWM and autochorous dispersal syndrome were
38 significantly related to soil, whereas climate did not significantly influence any of the
39 ecological attributes considered. Our results show that anthropogenic disturbances can be
40 important drivers of ecological modifications in the tree communities, ultimately affecting the
41 provision of essential ecosystem services.

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52 **Introduction**

53 Tropical forests are highly biodiverse terrestrial systems that store large amounts of
54 carbon and have an important role on climate change mitigation strategies (Poorter et al.
55 2015; Van der Sande et a. 2017; Sullivan et al. 2017; Fujii et al. 2018). Climate and soil are
56 important drivers of tree community structure in these forests (Feldpausch et al. 2011; Condit
57 et al. 2013; Van der Sande et al. 2016). For instance, climate factors such as precipitation
58 directly influence tree growth and, consequently, plant productivity in forest communities
59 (Toledo et al. 2011; Wagner et al. 2012). In addition, soil properties (e.g., fertility) can
60 directly affect tree productivity and growth, selecting species and functional traits (Quesada et
61 al. 2012; Prado-Junior et al. 2016; Van der sande et al. 2017, 2018). For example, although
62 scarce in tropical formations, phosphorus is considered the main predictor of tree growth rates
63 (Hedin et al. 2009, Quesada et al. 2012). Calcium, potassium and magnesium also seem to be
64 important determinants of growth and productivity rates (Wright et al. 2011; Baribault et al.
65 2012). Another example is found in African tropical forest communities, where soil texture
66 affects the functional characteristics displayed by the species: sandy soils harbor a larger
67 number of dry-tolerant and high wood density species in comparison with clayey soils
68 (Fayolle et al. 2012). Therefore, changes in climate patterns and spatial variation in the
69 edaphic environment may entail shifts in growth rates, floristic composition and community
70 function (Toledo et al. 2011; Wagner et al. 2012; Quesada et al. 2012). However, although the
71 effects of past disturbances (from both natural and anthropogenic origins) may have long-
72 lasting effects on tree community ecological processes (Burgi et al. 2017; Sfair et al. 2018),
73 the aforementioned studies have not accounted for their influence.

74 Natural disturbances play an important role in community dynamics, contributing to
75 landscape heterogeneity and determining the evolutionary history of plants (Crausbay &
76 Martin 2016; Lewis et al., 2015; Seidl et al. 2011). Sudden disturbance events alter

77 community structure, composition, demographic parameters (mortality and recruitment) and
78 ecosystem function (Rodrigues et al. 2017). But human-driven disturbances, prompted by
79 land use change and overexploitation of natural resources, currently represent the main threat
80 to global biodiversity (Laurance et al. 2011; Simberloff et al. 2013; Giam 2017).
81 Anthropogenic disturbances also alter the atmospheric composition and climate, posing
82 further threats to the provision of ecosystem services (Laurance et al. 2011).

83 The Brazilian Atlantic Forest Domain, one of the world's biodiversity hotspots and
84 provider of ecosystem services in the most industrialized region of Brazil (Ribeiro et al.
85 2009), has suffered from severe fragmentation and habitat loss. Today, only 12 to 16% of its
86 original extent remains, mostly confined to small fragments (Ribeiro et al. 2009; SOS Mata
87 Atlântica, 2020). The future of the Atlantic Forest and the ecosystem services it provides
88 depend upon our ability to understand the changes and effects promoted by past
89 anthropogenic disturbances on present-day ecosystem function (Metzger et al. 2009; Santo-
90 Silva et al. 2016).

91 The dynamics of human-disturbed landscapes is a central concern of tropical forest
92 conservation biology. Although evidence has been gathered that anthropogenic disturbances
93 affect different ecological processes (Johnson & Miyanishi, 2010; Keane et al., 2015; Peres et
94 al. 2016), the extent to which populations and communities are restructured by environmental
95 change remains uncertain (Santo-Silva et al. 2016). For example, logging affects the light
96 availability in the interior of forest fragments, promoting shifts in tree community structure,
97 diversity, dynamics and species composition (Morris, 2010; Decaens et al. 2018; Santos et al.
98 2020). However, little is known about how these changes will alter long-term community
99 dynamics, since the disturbance effects may be substantially obscured by tree longevity
100 (Norden et al. 2015; Martínez-Ramos et al. 2016).

101 Data about past disturbances and their interactions with community dynamics are
102 lacking for most tropical forests (McMichael et al. 2017). Knowledge about the history of
103 human disturbances is an essential component of ecology, which focuses on understanding the
104 importance and long-term effects of past events on ecosystems (Burgi et al., 2017; Szabó,
105 2015). Disturbance history effects can persist for a long time and influence ecosystem
106 function and biodiversity (Szabó 2015; Perring et al., 2016). Seemingly subtle present-day
107 human impacts may produce long-term effects on soil and vegetation, sometimes being
108 detected only centuries later (Burgi et al., 2017).

109 In this study, we start from the premise that, other than the main abiotic drivers of tree
110 community structure (i.e., climate and soil), the history of anthropogenic disturbances on the
111 Brazilian Atlantic Domain play a key role on the functionality and temporal composition
112 displayed by their tree communities. We expect present-day vegetation to reflect past
113 anthropogenic disturbances undergone by the target communities. We therefore tested two
114 hypotheses: (i) past anthropogenic disturbances alter community functional parameters, with
115 increasing intensity predicting a decrease in carbon uptake, seed dispersal capacity and
116 successional dynamics; and (ii) a history of anthropogenic disturbances affect floristic
117 composition over time, in such a way that the most impacted areas tend to be more
118 homogenized in a temporal gradient. We monitored the temporal behavior of 12 fragments of
119 seasonal semideciduous forest fragments of the Atlantic Domain between years 2003 and
120 2019, accounting for the influence of past disturbances.

121

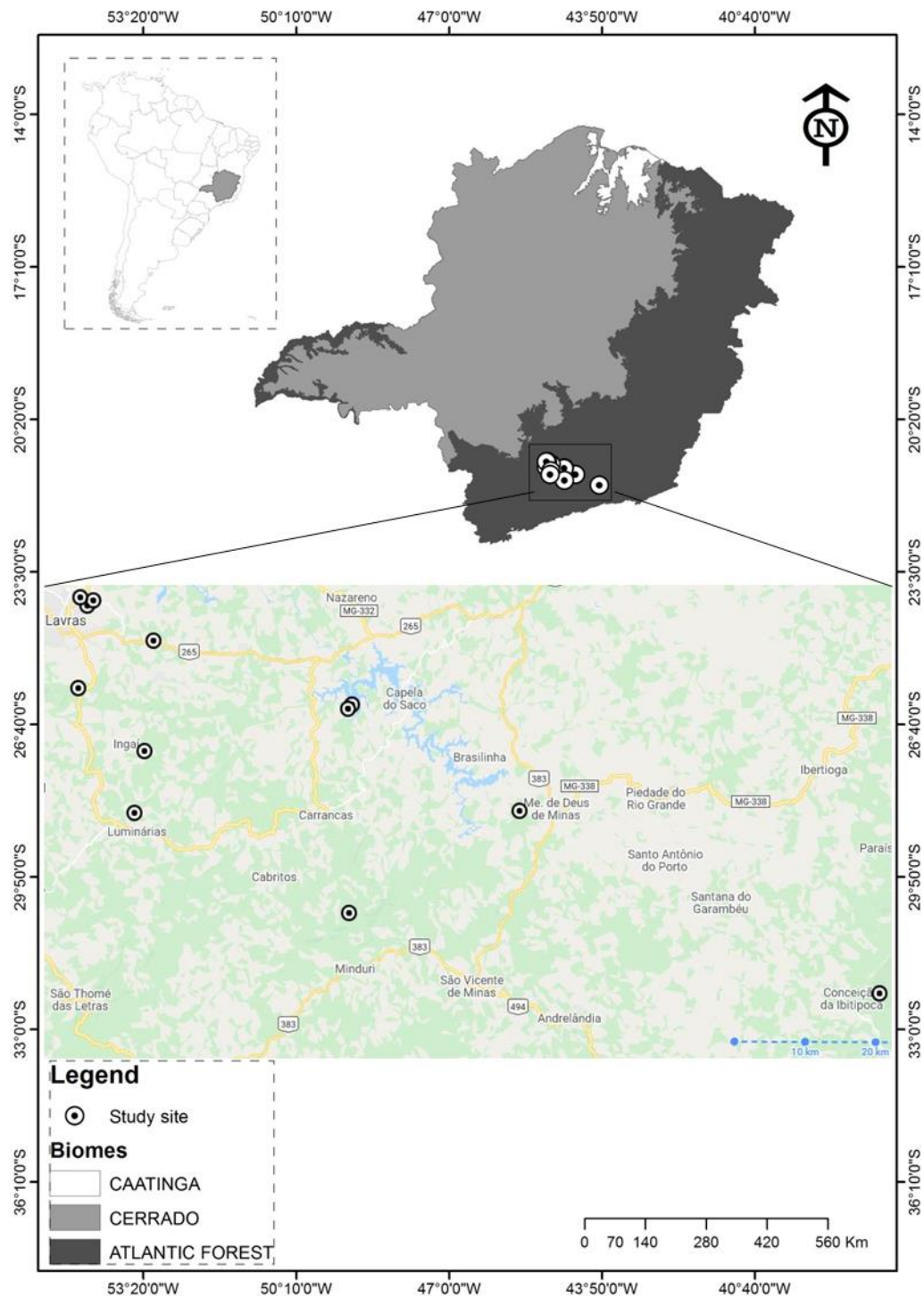
122 **Material and Methods**

123 ***Study site description***

124 We gathered data about 12 seasonal semideciduous forest fragments of the Atlantic
125 Domain in the southeast region of Brazil, in the southern portion of Minas Gerais state
126 (Figure 1). Climate in this region falls into the Aw and Cwa categories of Köppen's

127 classification (wet summers and dry winters) (Reboita et al. 2015). Mean annual temperature
 128 and mean annual precipitation are, respectively, 19.2°C e 1545,9 mm (Hijmans et al., 2005)
 129 (refer to Table 1 for further details).

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132 **Figure 1.** Location of the 12 Atlantic Domain fragments sampled in the southern portion of
 133 Minas Gerais state, Brazil

Table 1. Environmental description of the 12 Atlantic Domain fragments sampled in the southern portion of Minas Gerais state, Brazil

Id.	Fragment	Site area (ha)	Latitude (S)	Longitude (W)	MAP (mm)	MAT (°C)	Altitude (m)	Inventories (years)
1	Mata da UFLA	5.72	-21.2287905°	-044.9711195°	1529.7	19.4	918-937	2006, 2010, and 2017
2	Poço Bonito	88.56	-21.3312690°	-044.9837098°	1529.7	18.6	925-1210	2006, 2011 and 2016
3	Madre de Deus	20.66	-21.4839350°	-044.3790290°	1588.5	19.4	915- 980	2006, 2009 and 2018
4	Itutinga	3.77	-21.3524520°	-044.6088975°	1514.5	19.4	913-945	2004, 2010 and 2019
5	Camargos	10.36	-21.3565432°	-044.6146468°	1514.5	19.4	913-960	2003, 2009 and 2019
6	Ibitipoca	95.05	-21.7102307°	-043.8867144°	1544.2	14.8	1150- 1510	2005, 2013 and 2019
7	Capivari	9.78	-21.2729492°	-044.8810300°	1529.7	19.4	825- 875	2003, 2008 and 2017
8	Subestação	8.73	-21.2229634°	-044.9629445°	1529.7	19.4	910- 940	2005, 2010 and 2016
9	Mata da Lagoa	3.97	-21.2195251°	-044.9802359°	1529.7	19.4	855-902	2005, 2011 and 2017
10	Carrancas	35.98	-21.6105958°	-044.6131448°	1483.0	14.8	1440- 1513	2005, 2010 and 2018
11	Ingaí	16.14	-21.4094428°	-044.8924867°	1529.7	19.4	860-890	2004, 2009, and 2017
12	Luminárias	77.91	-21.4857763°	-044.9061968°	1529.7	19.4	880- 1000	2005, 2010 and 2018

Note: MAP – mean annual precipitation; MAT – mean annual temperature.

132 *Data collection*

133 Systematic sampling was employed to characterize the tree communities in the 12
134 study sites, totaling 544 plots of 200 m², 225 m², 300 m² e 400 m² (18.28 ha of total area
135 sampled). All living individuals meeting the inclusion criterion of 5.0 cm of diameter at breast
136 height (DBH, 1.30 m) were recorded. The DBH of multiple stemmed individuals was
137 represented by the square-root of the sum of squared values of each stem's DBH (MacDicken
138 et al. 1991). Non-arboreal life forms were not included in the dataset, and the same criteria
139 were followed in each inventory carried out throughout the timeframe of data collection (see
140 Table 1). We assembled data about dispersal syndromes for the species occurring in our
141 dataset based on the literature. The dataset used herein belongs to the Laboratory of
142 Phytogeography and Evolutionary Ecology, located in the Department of Forest Sciences at
143 the Federal University of Lavras (UFLA), Brazil. All data are hosted in the forestplots.net
144 system (Lopez-Gonzalez et al. 2009; Lopez-Gonzalez et al. 2011), available upon request.

145 To characterize the edaphic environment of each fragment, composite soil samples of
146 0.5 kg were collected from three points within each plot (20 cm of depth). The following
147 chemical variables were analysed: Ca, Mg, Na, K, P, pH, cation exchange capacity, base
148 saturation, aluminum saturation and soil organic matter. Percentages of sand, silt and clay
149 characterized soil texture. All soil analyses were carried out in the Laboratory of Soil
150 Analyses at Federal University of Lavras (UFLA). To characterize the climate of the study
151 region, we extracted the 19 bioclimatic variables from WorldClim 1.4 (11 related to
152 temperature, and 8 related to precipitation) (Hijmans et al. 2005).

153 To assess the influence of past anthropogenic impacts on present-day vegetation
154 patterns, we used the data assembled by Pereira (2003), which relied upon the Leopold matrix
155 for environmental impact assessment (Leopold et al. 1971) (Appendix S8 in the Supporting
156 Information). The rows of this matrix list the 12 sampled fragments, whereas the columns

157 contain environmental and disturbance-related variables, namely: (1) type of anthropogenic
158 disturbance and (ii) surrounding matrix. Pereira (2003) attributed weights (ranging from 1 to
159 6) to all disturbance variables present in the columns, in order to characterize the impact's
160 damaging potential. Each intersection between rows and columns was divided into four
161 quadrants, each of which aimed to characterize one of the following disturbance attributes: (i)
162 severity (quadrant 1), (ii) spatial extent (quadrant 2), (iii) duration (quadrant 3), and (iv)
163 magnitude of the impact (quadrant 4), representing the sum of the three former quadrants. The
164 scores of each of these disturbance attributes varied between 0 and 4 (except for magnitude,
165 which ranged between 0 and 12), meaning: 0 – absent disturbance, 1 – low disturbance
166 effects, 2 – moderate disturbance effects, 3 – high disturbance effects, and 4 – very high
167 disturbance effects. Therefore, for each fragment, the intersection between rows and columns
168 resulted in a given score regarding each of the anthropogenic disturbance impacts assessed,
169 which were the following: fire, logging (which received the highest impact weight), trails,
170 cattle, internal and external roads, fence and littering (which received the lowest impact
171 weight). We categorized the land use of the surrounding matrix as agriculture, silviculture and
172 pasture. Finally, we also employed the same methodology as Pereira (2003) (i.e., Leopold
173 matrix) to characterize the anthropogenic disturbance impacts underwent by the 12 fragments
174 in the year 2019 (Appendix S7 in the Supporting Information).

175

176 *Data analysis*

177 To test for the disturbance effects on vegetation composition and function, we
178 obtained the mean values of each vegetation variable from each inventory (structural
179 variables) and from the intervals in-between (dynamics variables). We also extracted the
180 following information from the Leopold matrix (i) final score, from the weighted average
181 between the fragment's score (impact magnitude) and impact weight, and (ii) fragment size

182 and fragment shape index (the ratio between the fragment's perimeter and the square-root of
183 its area).

184 We analyzed the following response variables: (i) average aboveground biomass
185 recorded in the timeframe assessed (average between the multiple inventories); (ii) average
186 productivity, (iii) mean temporal beta diversity, (iv) wood density community-weighted
187 means (CWMWD) and (v) dispersal syndrome (percentage of individuals belonging to a
188 given category, e.g., anemochory, zoochory, autochory). We calculated biomass values for
189 each of the multiple inventories based on the pantropical equation by Chave et al. (2014),
190 using package *biomass* (Rejou-Mechain et al. 2017). We averaged the plot-level values to
191 characterize whole-fragment structure for each of the sites. The same logic was applied to
192 calculate wood density community-weighted means (obtained with the *FD* package by
193 Laliberté et al., 2014), and to calculate the proportion of individuals belonging to the dispersal
194 syndromes. For each of these cases, we calculated a final fragment-level value based on the
195 observed values across the multiple inventories. Productivity was obtained by the difference
196 between biomass values recorded in different inventories divided by the number of years in
197 between them, and later corrected following Talbot et al. (2014), to account for the different
198 time intervals (in years) between the inventories. We calculated the trend of temporal biomass
199 variation for each fragment by averaging the productivity recorded in each interval. We
200 employed the same approach to obtain an average measure of the temporal trend in species
201 composition variation, by calculating the mean temporal beta diversity recorded in each
202 interval, using package *betapart* (Baselga and Orme 2012).

203 Due to the collinearity verified between the environmental control variables (i.e.,
204 climate and soil), we applied principal component analysis (PCA) to obtain a better depiction
205 of their variation. We performed a PCA for climate and another for soil, separately. Adhering
206 to the Kaiser-Guttman rule, whereby axes with eigenvalues above 1 are maintained, we used

207 three PCA axes from each (climate and soil) analysis. We used these six axes as control
208 variables in the models. We pre-selected the highly correlated variables (Pearson's correlation
209 $\geq |0.95|$) by choosing to maintain the higher order variables, e.g., mean annual temperature
210 instead of mean temperature of wettest quarter (Dormann et al., 2012).

211 We used generalized linear mixed models to assess the effects of the environmental
212 impacts on the vegetation variables. Fragment was added as a random factor to deal with
213 pseudoreplication arising from the nested sampling of our data (plots within fragments). We
214 included in the models the six PCA axes to control for the expected climate and soil effects.
215 For each response variable, we adopted the model selection approach based on Akaike
216 Information Criterion of second order (AICc). From the global model containing all
217 explanatory variables, we selected the best models ($\Delta\text{AICc} < 4$) (Burnham et al. 2011) that
218 only contained variables with Pearson's correlation $< |0.6|$ (Dormann et al. 2012; Burnham et
219 al. 2011). We then applied a multimodel inference approach, obtaining the coefficients'
220 conditional averages and confidence intervals based on the selected models. When necessary,
221 we ensured normality of residuals through log or cube-root transformation. We also checked
222 for spatial independence in each model's residuals (both for models of the ecological
223 variables and the anthropogenic disturbance matrices).

224 We used packages lme4 and lmerTest for model fitting (Bates et al. 2015, Kuznetsova
225 et al. 2016), MuMIn for model selection and multimodel inference (Bartón 2009), and *ncf* to
226 test for spatial correlation (Bjørnstad 2016). All analyses were performed in the R
227 environment (R Core Team 2018), adopting a significance level of 5%.

228

229

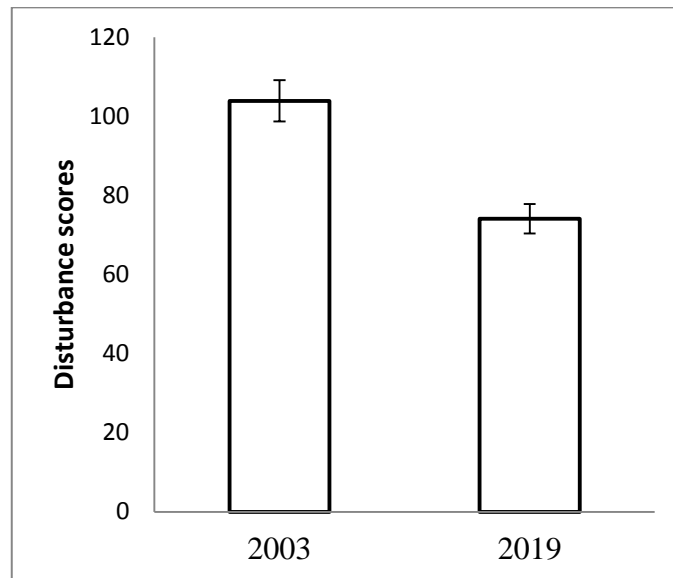
230

231

232 **Results**

233 We recorded a total of 36,638 individuals belonging to 544 species in 76 families in
 234 the 12 fragments sampled. The intensity of anthropogenic disturbances on the forest
 235 communities decreased significantly in the studied timeframe ($p < 0.05$): Pereira (2003)
 236 recorded a final score of 103.9 points of human impacts, whereas we recorded 74.1 points, all
 237 12 fragments considered (Figure 1). In this decrease, the highlight goes to absence of clear-
 238 cutting activity in the current matrix and the selective logging that had an almost non-existent
 239 impact (**Appendices S7 and S8** in the Supporting Information).

240



241

242 Figure 1. Temporal mean's classification of anthropogenic disturbances applied by Pereira
 243 (2003) and in 2019 (that was recently scored by this study). The bars represent the standard
 244 deviation between sample units within each category, according to the *lsmeans* contrasts
 245 applied on the model obtained in GLMM, at a significance level of 5%

246 Three dispersal syndromes were represented among all recorded species in the 12
 247 fragments: anemochory (wind-dispersed species), autochory (self-dispersed species) and
 248 zoochory (animal-dispersed species) (refer to Table 2 for details). The abundance of self-
 249 dispersed species was positively and significantly affected by anthropogenic disturbances ($p <$

250 0.01), but not by soil variation. The coefficient of determination (R^2) for the self-dispersed
 251 species abundance model was 0.432, meaning that 43.2% of the variation in these species'
 252 abundances were explained by the explanatory variables. Animal-dispersed species
 253 abundances were also significantly, albeit negatively, affected by the recorded disturbances (p
 254 < 0.001 , $R^2 = 0.408$). This syndrome also responds to soil variations, however, the soil showed
 255 a lesser effect (-1.894) than anthropogenic disturbances (-5.650). Conversely, the abundances
 256 of wind-dispersed species were not significantly affected by anthropogenic disturbances, but
 257 were significantly influenced by soil variation (Table 3).

258

259 **Table 2.** Generalized linear mixed model results for self-dispersal (autochory) and
 260 animal-dispersal (zoochory) syndromes as function of impact magnitude, soil, climate,
 261 fragment size and shape index. Note: (*) 5% significance level; (**) 1% significance level;
 262 (***) 0.1% significance level. ADs: Anthropogenic disturbances, FS: fragment size, FSI:
 263 fragment shape index, Est: estimate, R^2 : coefficient of determination.

	Autocory			Zoocory		
	Est.	<i>p</i> -value	R^2	Est.	<i>p</i> -value	R^2
Intercept	1.134	<2e-16 ***	0.432	79.308	<2e-16 ***	0.408
ADs	0.523	7.3e-06 ***		-5.650	0.00644 **	
FS	-0.189	0.114		3.873	0.083	
FSI	0.035	0.293		-0.922	0.073	
Soil: 1st axis	0.081	0.120		-1.894	0.013 *	
Soil: 2nd axis	-0.078	0.069		-0.597	0.360	
Soil: 3rd axis	-0.015	0.790		1.532	0.075	
Climate: 1st axis	0.146	0.204		0.089	0.967	
Climate: 2nd axis	-0.121	0.231		2.963	0.128	
Climate: 3rd axis	-0.165	0.127		0.264	0.904	

264

265 **Table 3.** Generalized linear mixed model results for wind-dispersal syndrome
 266 (anemochory) as function of impact magnitude, soil, climate, fragment size and shape index.
 267 Note: (*) 5% significance level; (**) 1% significance level; (***) 0.1% significance level.
 268 ADs: Anthropogenic disturbances, FS: fragment size, FSI: fragment shape index, Est:
 269 estimate, R^2 : coefficient of determination.

Anemochory			
	Est.	<i>p</i> -value	R^2
Intercept	2.235	<2e-16 ***	0.306
ADs	0.172	0.152	
FS	-0.117	0.395	
FSI	0.023	0.452	
Soil: 1st axis	-0.093	0.044 *	
Soil: 2nd axis	0.06	0.100	
Soil: 3rd axis	-0.089	0.800	
Climate: 1st axis	0.083	0.529	
Climate: 2nd axis	-0.017	0.884	
Climate: 3rd axis	0.094	0.449	

270

271 Wood productivity was significantly ($p < 0.05$) and negatively affected by
 272 anthropogenic disturbances, but not by the soil variables ($R^2 = 0.554$) (Table 3). Wood
 273 density community-weighted means (CWMWD), however, responded significantly and
 274 negatively to anthropogenic disturbances ($p < 0.05$, $R^2=0.128$) (Table 4). Unlike productivity,
 275 soil significantly (and negatively) influenced wood density CWM values ($p < 0.05$), with
 276 equal effect sizes to the disturbance effects.

277

278

279

280 **Table 4.** Generalized linear mixed model results for productivity and wood density
 281 community-weighted means (CWMWD) as function of the magnitude of impacts, soil,
 282 climate, fragment size and fragment shape index. Note: (*) 5% significance level; (**) 1%
 283 significance level; (***) 0.1% significance level. ADs: Anthropogenic disturbances, FS:
 284 fragment size, FSI: fragment shape index, Est: estimate, R²: coefficient of determination.

	Productivity			CWMWD		
	Est.	<i>p</i> -value	R ²	Est.	<i>p</i> -value	R ²
Intercept	9.982	<2e-16***	0.554	0.589	<2e-16***	0.128
ADs	-1.057	0.0177*		0.008	0.018*	
FS	-0.113	0.818		0.0004	0.766	
FSI	0.113	0.685		0.0002	0.947	
Soil: 1st axis	0.046	0.893		-0.008	0.0002***	
Soil: 2nd axis	-0.296	0.346		-0.002	0.399	
Soil: 3rd axis	-0.166	0.679		-0.002	0.488	
Climate: 1st axis	0.079	0.75		0.005	0.175	
Climate: 2nd axis	0.590	0.152		0.001	0.495	
Climate: 3rd axis	0.364	0.415		-0.002	0.440	

285
 286 According to the models, aboveground biomass (AGB) and temporal beta diversity
 287 were not significantly affected by anthropogenic disturbances ($p > 0.05$), although the latter
 288 had a positive, significant relationship with soil (axis 1 and axis 3). Other explanatory
 289 variables also associated with disturbance effects (i.e., fragment size and shape index) did not
 290 significantly influence any of the other response variables.

291

292 **Table 5.** Temporal beta diversity as function of impact magnitude, soil, climate,
 293 fragment size and shape index. Note: (*) 5% significance level; (**) 1% significance level;
 294 (***) 0.1% significance level. ADs: Anthropogenic disturbances FS: fragment size, FSI:
 295 fragment shape index, Est: estimate, R²: coefficient of determination.

Temporal beta diversity			
	Est.	<i>p</i> -value	R ²
Intercept	0.525	<2e-16 ***	0.152
ADs	-0.006	0.456	
FS	-0.0008	0.919	
FSI	0.0008	0.837	
Soil: 1st axis	0.021	0.0002***	
Soil: 2nd axis	-0.03	0.502	
Soil: 3rd axis	0.028	3.5e-06 ***	
Climate: 1st axis	-0.001	0.894	
Climate: 2nd axis	-0.006	0.404	
Climate: 3rd axis	0.003	0.672	

296

297 **Discussion**

298 Our results supported our first hypothesis that anthropogenic disturbances significantly
299 affect dispersal syndromes in our study areas. For instance, the animal-dispersal syndromes
300 (zoochory) were more affected by anthropogenic disturbances than soil, in spite of soil's role
301 as a driver of community structure. The higher the impacts, the least abundant were the
302 animal-dispersed species in a given area. Soil also had a negative effect on the abundance of
303 animal-dispersed species, meaning that the most fertile sites had a smaller species abundance
304 with this syndrome. This reduction in animal-dispersed species abundances may be explained
305 by the loss of dispersers caused by the anthropogenic disturbances. These disturbances affect
306 dispersal in three fundamental aspects, namely: (i) by reducing the frequency of disperser
307 visits and abundance, through local extinctions or forced migration (Markl et al. 2012;
308 Fonturbel et al. 2015); (ii) by exposing the seed banks and increasing the likelihood of seed
309 predation and fungi attack; and (iii) by reducing the dispersal distance by changing local

310 biophysical characteristics coupled with the decrease in seed disperser abundance (Caves et
311 al. 2013; Schleuning et al, 2011). These consequences are also reflected in forest regeneration,
312 plant recruitment and other ecological processes in the community (Farwig & Berens 2012;
313 Markl et al. 2012). Several authors highlight that the loss of dispersers alters a community's
314 ecological functions, entailing the homogenization of floristic composition and structure, and
315 loss of ecosystem services (García & Martínez 2012; Schleuning et al. 2015; Caughlin et al.
316 2015).

317 The reduction in seed dispersers driven by anthropogenic disturbances can lead to
318 local extinctions and great biodiversity loss, especially for plant species dispersed by single
319 animal species (Rodríguez-Cabal et al. 2007; Beckman & Rogers 2013; Farwig & Berens,
320 2012). Zoochory is the main dispersal syndrome in tropical forests, where more than 80% of
321 tree species depend on animal dispersers (Muller-Landau et al. 2008; Au et al 2006). The
322 breakdown in plant-animal interactions, specifically seed dispersal, enhances the chances of
323 predator and pathogen attacks on the undispersed seeds, having profound consequences on
324 plant recruitment, population persistence and community dynamics (Markl et al. 2012; Farwig
325 & Berens 2012). Culot et al. (2017) found that plant recruitment decreased by 70.7 and 94.9%
326 due to the loss of frugivore dispersers. Additionally, species that produce larger seeds are
327 under greater threat for being usually dispersed by large sized vertebrates, given the positive
328 relationship between seed size and frugivore disperser size (Jordano 2017; Fuzessy et al.
329 2018). In the case of our study sites, most are small fragments without minimum conditions or
330 resources to support the occurrence of adequate dispersers, which can lead to a lack of
331 recruitment and persistence of tree species dependent on them (Bueno et al. 2013; Corlett
332 2017).

333 Disturbances tend to favor autochorous (or self-dispersal) syndromes, manifested by a
334 higher abundance of self-dispersed species in tree communities under higher disturbance

335 impacts. This may be linked to the higher frequency of gaps in human-disturbed fragments.
336 Several authors have found significant increases of autochorous species abundances in
337 human-disturbed environments, especially in forest gaps resulting from selective logging,
338 where these plants encounter suitable space and resources for colonization (Drezner et al.
339 2001; Kuhlmann & Ribeiro 2016). These self-dispersed species may therefore not require
340 dispersal agents, since they usually produce dehiscent fruits that facilitate seed dispersal
341 through gravity (Jara-Guerrero et al. 2011; Parolin et al. 2013). Conversely, although the
342 abundance of anemochorous, or wind-dispersed, species did not respond significantly to
343 anthropogenic disturbances, it was negatively affected by soil fertility, i.e., the lowest fertility
344 sites had higher abundance of wind-dispersed species. This is congruent with evidence that
345 infertile soils may promote wind-dispersal efficiency (Spasojevic et al. 2014).

346 The temporal beta diversity recorded in this study showed that human disturbances did
347 not significantly affect species turnover of the studied fragments within the timeframe
348 analysed. Based on this, we reject our second hypothesis. However, this variable was
349 significantly and positively affected by the soil, meaning that the most fertile soils had higher
350 species substitution in the timeframe of study. This result provides evidence of the structuring
351 effect of soil on the tree communities of our study fragments. Other studies that analysed the
352 effect of soils on tropical forest community structure have found similar results to us, whereby
353 variation in soil attributes significantly influenced tree species composition (Kumar et al.
354 2010; Long et al. 2012 ; Baldeck et al. 2013; Prada et al. 2017; Van der Sande et a. 2018).
355 For example, Long et al. (2012) found a positive effect of soil fertility on tree species
356 richness.

357 Wood productivity was negatively affected by anthropogenic disturbances, meaning
358 that sites more exposed to impacts had lower productivity. This finding reinforces our first
359 hypothesis that human disturbances influence tree community productivity. This inverse

360 relationship between productivity and anthropogenic disturbance has been explained by other
361 authors, which state that disturbance intensity negatively influences ecosystem productivity,
362 ecological interactions and resilience to the disturbance itself (Amiro et al. 2010; Flower et al.
363 2014; Flower & Gonzalez-Meler 2015; Peters et al., 2013). Gautam & Mandal (2016) found
364 similar results when analyzing the effects of human disturbances on carbon dynamics and
365 productivity. They found that human disturbances decreased productivity by 44%, and
366 negatively affected ecosystem structure and functioning. Other studies have found that, the
367 higher intensity of logging (which we also evaluated), the lower are ecosystem productivity
368 which ultimately lead to the loss of community function (Martin et al. 2015; Mori et al. 2017;
369 Roopsind et al. 2018). Primary productivity is a good proxy of ecosystem carbon uptake
370 (Girardin et al. 2010). Small changes in productivity at global, regional or even local scales
371 can therefore have substantial effects on ecosystem function and net rates of carbon
372 sequestration (Malhi 2011; Malhi et al. 2014). This may ultimately affect the provision of
373 ecosystem goods and services, including nutrient cycling, soil formation, food, water,
374 medicines, erosion control, purification of water and air, polinization and carbon sequestration
375 (Parrotta et al. 2012; Berenguer et al. 2014; Lewis et al. 2015; Shimamoto et al. 2017). The
376 latter makes wood productivity an essential component of climate change mitigation (Parrotta
377 et al. 2012; Kirby et al. 2017).

378 Concerning the community-weighted means (CWM) for wood density, the
379 anthropogenic disturbances had a positive effect, meaning that the higher the impact, the
380 higher the mean values of wood density in the tree community. However, during the
381 timeframe of our study (2003-2019), the incidence of disturbances on the studied
382 communities decreased (Figure 1), which might reflect the adoption of forest conservation
383 policies (Pereira 2003). For instance, logging activities were almost entirely absent from all
384 fragments in the last disturbance impact assessment, in year 2019 (Appendices S7 and S8 in

385 the Supporting Information). The fact that human disturbances were associated with the
386 occurrence of high wood density species may be related to their higher commercial value,
387 which may have stimulated intense human activity in forests with high mean values of wood
388 density in the past (Pereira et al. 2015). The presence of high wood density species today may
389 indicate that these forests are at advanced successional stages, given that high wood density,
390 shade-tolerant and slow-growth species are the last to emerge in the successional process
391 (Nascimento et al. 2005; Powers & Marín-Spiotta, 2017; Hiltner et al. 2018). Moreover,
392 several studies have shown that increases in wood density are associated with increases in the
393 abundance of late successional species, at the expense of decreasing abundance and
394 dominance of pioneer (fast-growing, low wood density) populations (Slik et al. 2008;
395 Chazdon et al. 2014; Lin et al. 2015). Conversely, soil had a negative effect on wood density
396 CWM, meaning that higher fertility predicted lower mean values of wood density. The fact
397 that areas with low wood density species are less impacted by human disturbances may explain
398 their higher fertility. For instance, logging intensity directly impacts soil quality, leading to
399 decreased fertility, soil compaction and consequently the loss of ecosystem function (Singh et
400 al. 2010; Bowd et al. 2019). In addition, the effects of anthropogenic disturbances and soil
401 characteristics had equal effect sizes on this variable, indicating that both may be important
402 determinants of wood density CWM in the studied communities.

403 The results of our study showed that, although human disturbances exert strong
404 negative impacts on productivity and on the dispersal of zoochorous species, they were not
405 enough to alter the advanced successional stage of the studied fragments. This may be
406 explained by a decline in the disturbance impacts over time, recorded in the Leopold matrix of
407 anthropogenic disturbance characterization (Figure 1). Furthermore, unlike other studies that
408 analyzed disturbance effects on a smaller number of (or even single) forest fragments
409 (Kleinschroth & Healey 2017; Sfair et al. 2018; Ding et al. 2019; Gaui et al. 2019), our study

410 provides novel insight into the how the effects of anthropogenic disturbance histories,
411 interacting with natural drivers of tree community structure (i.e., climate and soil), are
412 manifested in tropical forest ecological patterns. Importantly, our study was performed in a
413 highly endangered biodiversity hotspot, the Brazilian Atlantic Forest, adding further relevance
414 to our findings, especially if taken into account that most present-day remnants are small
415 fragments that have been exposed to human disturbances. At last, we highlight that the
416 climate, in spite of being an important driver tree community structure on larger scales, did
417 not produce any significant effects on the ecological variables studied, which might reflect the
418 short climatic variation covered by the studied fragments (Schuur 2003; Adler & Levine
419 2007; Bai et al. 2008; Reboita et al. 2015). At last, our study shows that with exhaustive
420 sampling, anthropogenic disturbances can be important drivers of ecological modifications in
421 the tropical forest tree communities. These disturbances may compromise the provision of
422 essential ecosystem goods and services, relied upon by humanity.

423

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432

433

434

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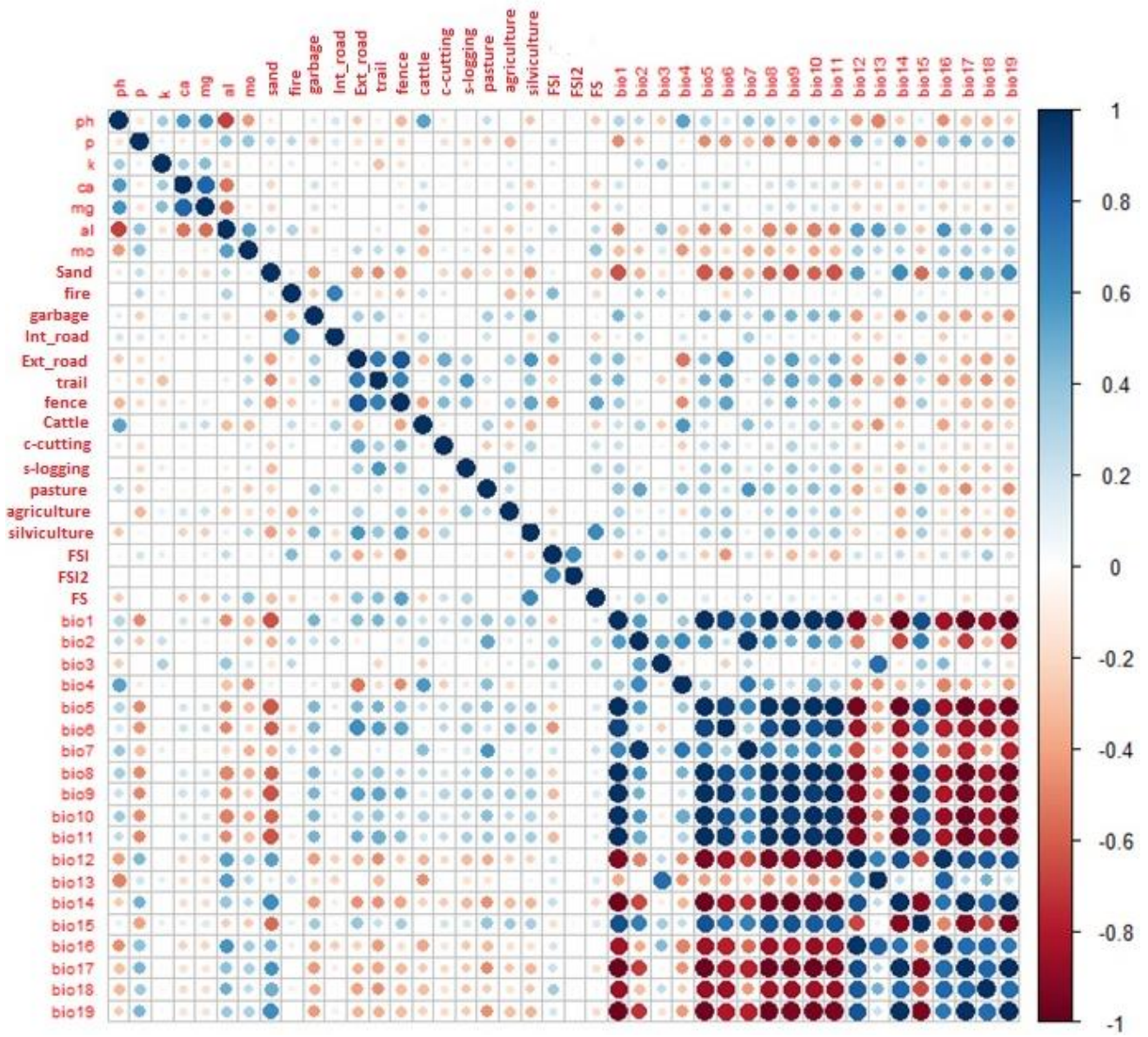
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Supporting information for the paper

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Appendix S1: Correlations among the 19 bioclimatic variables, soil and impact variables in the 12 forest fragments. Note: Note: P = phosphorus, K = potassium, Ca = calcium, Mg = magnesium, Al = aluminium, MO= organic matter; BIO1 = Annual Mean Temperature, BIO2 = Mean Diurnal Range, BIO3 = Isothermality, BIO4 = Temperature Seasonality, BIO5 = Max Temperature of Warmest Month, BIO6 = Min Temperature of Coldest Month, BIO7 = Temperature Annual Range, BIO8 = Mean Temperature of Wettest Quarter, BIO9 = Mean Temperature of Driest Quarter, BIO10 = Mean Temperature of Warmest Quarter, BIO11 = Mean Temperature of Coldest Quarter, BIO12 = Annual Precipitation, BIO13 = Precipitation of Wettest Month, BIO14 = Precipitation of Driest Month, BIO15= Precipitation Seasonality, BIO16 = Precipitation of Wettest Quarter, BIO17 = Precipitation of Driest Quarter, BIO18 = Precipitation of Warmest Quarter, BIO19 = Precipitation of Coldest Quarter; c-cutting= clear cutting; Int_road= internal road; Ext_ road= external road; s-logging= selective logging; FSI-fragment shaping index(weighted); FSI2= Fragment shaping index (simple); FS= fragment size



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Appendix S2: Correlations between the soil PCA axes and all soil variables (SV) included in analysis in 12 forest fragments. Note: Note: Note: P = phosphorus, K = potassium, Ca = calcium, Mg = magnesium, Al = aluminium, MO= organic matter.

SV	PC1	PC2	PC3	PC4	PC5	PC6	PC7	PC8
ph	20.6112	0.0033	5.9837	0.4048	11.3731	47.9130	13.2174	0.4934
p	4.0862	25.7951	13.7490	0.1887	47.5410	7.5278	1.0609	0.0513
k	5.7443	20.5127	2.0348	58.5461	10.4868	0.3478	0.7199	1.6076
ca	18.9309	7.4781	0.2155	18.1001	1.3952	7.5680	10.0133	36.298
mg	20.5787	7.2909	0.0231	7.9115	3.7403	4.3384	0.5023	55.614
al	21.4196	6.1033	0.0724	0.1688	4.0528	0.0933	63.1695	4.9203
mo	5.7456	32.6658	11.8213	7.1329	2.1782	30.2721	9.4991	0.6851
sand	2.8833	0.1507	66.1001	7.5472	19.2326	1.9397	1.8176	0.3286

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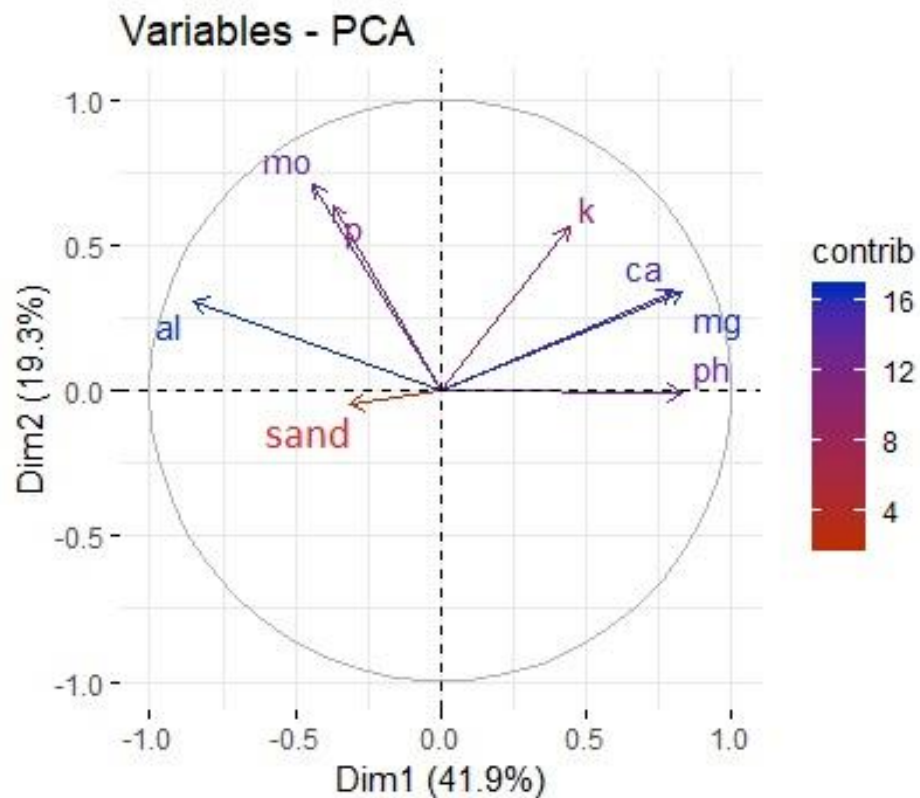
Appendix S3: Correlations between the bioclimatic PCA axes and all bioclimatic variables (BCV) included in analysis, in 12 forest fragments. Note: BIO1 = Annual Mean Temperature, BIO3 = Isothermality, BIO4 = Temperature Seasonality, BIO6 = Min Temperature of Coldest Month, BIO7 = Temperature Annual Range, BIO12 = Annual Precipitation, BIO13 = Precipitation of Wettest Month, BIO15= Precipitation Seasonality, BIO17 = Precipitation of Driest Quarter, BIO18 = Precipitation of Warmest Quarter.

BCV	PC1	PC2	PC3	PC4	PC5	PC6	PC7	PC8
bio1	16.003 5	0.3893	1.3212	0.0862	2.33E+0 0	2.7375	6.7865	8.07E+0 0
bio3	0.2413	43.004 7	0.5372	12.598 1	1.36E+0 1	28.4676	0.0029	1.16E+0 0
bio4	3.9797	0.7076	49.157 6	24.549 6	9.95E-01	18.7089	0.0025	6.04E-01
bio6	12.403 3	0.3789	16.623 4	0.0094	5.68E+0 0	7.6201	3.8856	3.26E+0 0
bio7	8.7338	5.2155	23.028 9	8.0761	1.21E+0 0	34.2186	1.3861	4.41E+0 0
bio12	15.362 3	1.8302	0.0068	14.319 9	1.31E-05	0.6318	2.9696	1.45E+0 1
bio13	3.9323	32.001 5	2.6803	20.140 6	1.64E+0 0	3.8616	7.6545	1.74E+0 1
bio15	11.116 8	13.132 3	1.5635	4.0707	7.38E+0 0	1.4225	57.8105	2.52E+0 0
bio17	15.591 9	2.2658	0.0037	0.0689	3.05E+0 0	1.4249	19.4510	4.81E+0 1
bio18	12.635 1	1.0741	5.0775	16.080 5	6.42E+0 1	0.9059	0.0498	1.25E-06

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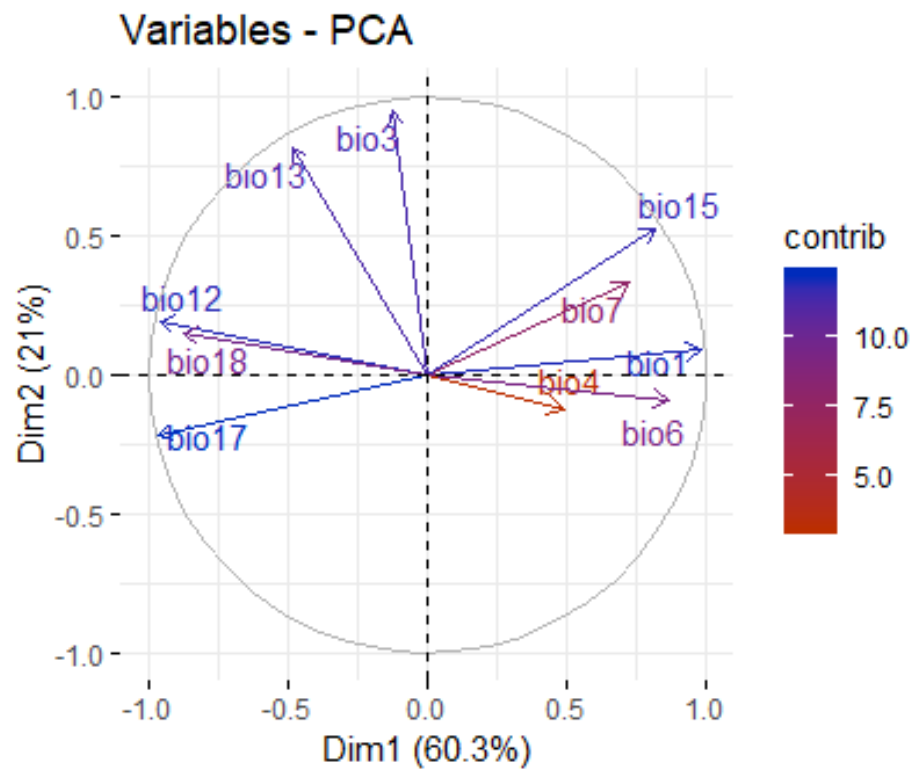
Appendix S4: Principal Component Analysis (PCA) of soil fertility, sampled in 12 forest fragments. Note: P = phosphorus, K = potassium, Ca = calcium, Mg = magnesium, Al = aluminium, MO= organic matter.



Supporting information for the paper

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Appendix S5: Principal Component Analysis (PCA) of bioclimatic variables, sampled in 12 forest fragments. Note: BIO1 = Annual Mean Temperature, BIO3 = Isothermality, BIO4 = Temperature Seasonality, BIO6 = Min Temperature of Coldest Month, BIO7 = Temperature Annual Range, BIO12 = Annual Precipitation, BIO13 = Precipitation of Wettest Month, BIO15 = Precipitation Seasonality, BIO17 = Precipitation of Driest Quarter, BIO18 = Precipitation of Warmest Quarter.



Supporting information for the paper

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Appendix S6: Temporal mean's classification of anthropogenic disturbances (ADs) applied by Pereira (2003) (Z) and in 2019 (that was recently scored by this study) (A), according to the obtained GLMM, at a significance level of 5%, using. Significant codes: 0.001 '***' 0.01 '**' '0.05 *'

Random effects:

Groups Name	Variance	Std.Dev.
fragmento (Intercept)	3165.6	56.26
Residual	441.6	21.01

Fixed effects:

	Estimate	Std. Error	df	t value	Pr(> t)
(Intercept)	74.083	15.712	12.761	4.715	0.000425
Z	29.833	8.579	11.000	3.477	0.00517 **

Lsmeans of impact:

ADs	lsmean	SE	df	lower.CL	upper.CL
A	74.1	15.7	12.8	40.1	108
Z	103.9	15.7	12.8	69.9	138

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Appendix S7: Current anthropogenic disturbances characterization matrix (year 2019) on the 12 Southern Minas Gerais forest fragments, Brazil.

Note: LV= Mata da UFLA, PB= Poço Bonito, MD= Madre de Deus, IT= Itutinga, CM= Camargos, IB= Ibitipoca, CP= Capivarí, SE= Subestação, ML= Mata da lagoa, CR= Carrancas, IN= Indaí, LU= Luminárias

Fragments		Anthropogenic disturbances																					Final score			
																				Surrounding matrix						
		Fire		Garbage		Internal roads		External roads		Trails		Fence		Cattle		Clear-cutting		Seletive logging		Pasture	agriculture	silvicultur e				
weights	5	1		4		3		2		1		4		6		5		3		2		1				
1	LV	0	0	1	4	0	0	3	11	4	10	4	12	0	0	0	0	0	0	1	5	0	0	3	10	94
		0	0	1	2	0	0	4	4	3	3	4	4	0	0	0	0	0	0	1	3	0	0	3	4	
2	PB	2	7	1	3	1	7	1	6	1	6	0	0	2	7	0	0	0	0	3	9	0	0	1	3	154
		2	3	1	1	2	4	1	4	1	4	0	0	1	4	0	0	0	0	2	4	0	0	1	1	
3	MD	0	0	1	5	0	0	0	0	1	4	0	0	2	5	0	0	0	0	3	10	0	0	1	3	66
		0	0	2	3	0	0	0	0	1	2	0	0	1	2	0	0	0	0	3	4	0	0	1	1	
4	IT	0	0	2	9	0	0	1	5	0	0	0	0	0	0	0	0	0	0	0	0	1	3	3	10	40
		0	0	3	4	0	0	1	3	0	0	0	0	0	0	0	0	0	0	0	0	1	1	3	4	
5	CM	0	0	1	3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	2	6	3	8	2	9	46
		0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	2	2	2	3	3	4	
6	IB	0	0	0	0	0	0	0	0	1	3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	6
		0	0	0	0	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
7	CP	0	0	3	10	0	0	1	3	3	9	0	0	0	0	0	0	1	3	4	10	0	0	0	0	82
		0	0	3	4	0	0	1	1	3	3	0	0	0	0	0	0	1	1	3	3	0	0	0	0	

Continues...

Fragments		Anthropogenic disturbances																						Final scores		
		Fire	Garbage		Internal roads		External roads		Trails		Fence		Cattle		Clear-cutting		Seletive logging		Surrounding matrix							
																			Pasture		agriculture		silviculture			
weights	5		1		4		3		2		1		4		6		5		3		2		1			
8	SE	0	0	1	3	0	0	2	6	0	0	1	3	0	0	0	0	0	0	3	9	3	9	0	0	69
		0	0	1	1	0	0	2	2	0	0	1	1	0	0	0	0	0	0	3	3	3	3	0	0	
9	ML	0	0	2	7	0	0	2	8	3	9	3	9	4	10	0	0	1	3	3	9	2	8	0	0	156
		0	0	2	3	0	0	3	3	3	3	3	3	3	3	0	0	1	1	3	3	3	3	0	0	
10	CR	0	0	0	0	0	0	0	0	1	3	0	0	0	0	0	0	0	0	2	8	0	0	0	0	30
		0	0	0	0	0	0	0	0	1	1	0	0	0	0	0	0	0	0	3	3	0	0	0	0	
11	IN	0	0	1	3	0	0	0	0	1	3	0	0	2	4	0	0	0	0	1	4	0	0	2	4	41
		0	0	1	1	0	0	0	0	1	1	0	0	1	1	0	0	0	0	1	2	0	0	1	1	
12	LU	0	0	2	6	1	6	0	0	2	5	0	0	1	3	0	0	0	0	3	10	3	10	1	3	105
		0	0	2	2	1	1	0	0	1	2	0	0	1	1	0	0	0	0	4	3	3	4	1	1	

a	d
b	c

Impact quadrants: a = severity, b= spatial extent, c= duration and d= magnitude (d= a+b+c); 0 – absent disturbance, 1 – low disturbance effects, 2 – moderate disturbance effects, 3 – high disturbance effects, and 4 – very high disturbance effects

Supporting information for the paper

Pais et al. Do abiotic factors drive ecological modifications in tropical forest fragments with a history of anthropogenic disturbances?

Conservation Biology

Appendix S8: Past anthropogenic disturbances characterization matrix from Pereira (2003) on the 12 Southern Minas Gerais forest fragments, Brazil

Fragments		Anthropogenic disturbances												Final scores									
		Fire		Garbage		Internal roads		External roads		Trails		Fence			Cattle		Clear-cutting		Seletive logging		Surrounding matrix		
		Pasture	agriculture	silviculture																			
	weights	5	1	4	3	2	1	4	6	5	3	2	1										
1	LV	0 0	1 3	0 0	3 9	4 11	4 12	0 0	0 0	3 10	1 6	1 6	2 6										150
		0 0	1 1	0 0	3 3	3 4	4 4	0 0	0 0	3 4	2 3	2 3	2 2										
2	PB	2 7	0 0	1 7	1 6	1 3	1 4	2 4	1 4	1 3	1 4	0 0	0 0										158
		2 3	0 0	2 4	1 4	1 1	1 2	1 1	1 2	1 1	1 2	0 0	0 0										
3	MD	0 0	0 0	0 0	0 0	2 6	0 0	4 12	0 0	1 3	2 7	0 0	0 0										96
		0 0	0 0	0 0	0 0	2 2	0 0	4 4	0 0	1 1	2 3	0 0	0 0										
4	IT	1 3	0 0	0 0	0 0	0 0	0 0	1 3	0 0	0 0	1 6	0 0	0 0										45
		1 1	0 0	0 0	0 0	0 0	0 0	1 1	0 0	0 0	3 2	0 0	0 0										
5	CM	1 3	0 0	0 0	0 0	0 0	0 0	1 3	0 0	0 0	1 6	0 0	1 3										48
		1 1	0 0	0 0	0 0	0 0	0 0	1 1	0 0	0 0	3 2	0 0	1 1										
6	IB	0 0	0 0	0 0	0 0	0 0	0 0	0 0	0 0	0 0	0 0	0 0	0 0										0
		0 0	0 0	0 0	0 0	0 0	0 0	0 0	0 0	0 0	0 0	0 0	0 0										

Continues...

Fragments		Anthropogenic disturbances																					Final scores			
		Fire		Garbage		Internal roads		External roads		Trails		Fence		Cattle		Clear-cutting		Seletive logging		Surrounding matrix						
																				Pasture		agriculture		silviculture		
Peso		5		1		4		3		2		1		4		6		5		3		2		1		
7	CP	1	3	2	6	0	0	1	4	2	7	0	0	1	3	2	8	3	8	1	3	1	3	0	0	162
		1	1	2	2	0	0	1	2	2	3	0	0	1	1	3	3	2	3	1	1	1	1	0	0	
8	SE	0	0	0	0	0	0	2	7	1	3	3	10	0	0	2	6	1	3	1	3	2	7	0	0	111
		0	0	0	0	0	0	2	3	1	1	3	4	0	0	2	2	1	1	1	1	2	3	0	0	
9	ML	1	3	1	3	2	6	3	7	4	11	0	0	4	12	2	6	2	6	2	6	0	0	0	0	217
		1	1	1	1	2	2	2	2	3	4	0	0	4	4	2	2	2	2	2	2	0	0	0	0	
10	CR	2	5	0	0	0	0	0	0	0	0	0	0	1	3	1	4	1	3	2	6	0	0	0	0	94
		1	2	0	0	0	0	0	0	0	0	0	0	1	1	1	2	1	1	2	2	0	0	0	0	
11	IN	0	0	0	0	0	0	0	0	1	3	0	0	2	7	0	0	1	3	1	3	0	0	0	0	58
		0	0	0	0	0	0	0	0	1	1	0	0	2	3	0	0	1	1	1	1	0	0	0	0	
12	LU	1	3	0	0	1	3	0	0	1	3	1	3	1	3	2	6	1	3	1	3	0	0	0	0	108
		1	1	0	0	1	1	0	0	1	1	1	1	1	1	2	2	1	1	1	1	0	0	0	0	

a	d
b	c

Impact quadrants: a = severity, b= spatial extent, c= duration and d= magnitude (d= a+b+c); 0 – absent disturbance, 1 – low disturbance effects, 2 – moderate disturbance effects, 3 – high disturbance effects, and 4 – very high disturbance effects

Supporting information for the paper

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Conservation Biology

Appendix S9: Species list by dispersal syndrome, occurring in 12 forest fragments. Note: Aut- Autochory, Zoo- Zoochory, Anemo- Anemochory

<i>Species</i>	Syndrome	<i>Species</i>	Syndrome
<i>Actinostemon concolor</i> (Spreng.) Müll.Arg.	aut	<i>Bowdichia virgilioides</i> Kunth	anemo
<i>Actinostemon klotzschii</i> (Didr.) Pax	aut	<i>Byrsonima laxiflora</i> Griseb.	zoo
<i>Actinostemon verticillatus</i> (Klotzsch) Baill.	aut	<i>Byrsonima myricifolia</i> Griseb.	zoo
<i>Aegiphila integrifolia</i> (Jacq.) Moldenke	zoo	<i>Cabrlea canjerana</i> (Vell.) Mart.	zoo
<i>Aegiphila verticillata</i> Vell.	zoo	<i>Callianthe bedfordiana</i> (Hook.) Donnell	anemo
<i>Agonandra excelsa</i> Griseb.	zoo	<i>Callisthene major</i> Mart.	anemo
<i>Aiouea saligna</i> Meisn.	zoo	<i>Calophyllum brasiliense</i> Cambess.	zoo
<i>Albizia polycephala</i> (Benth.) Killip ex Record	anemo	<i>Calycorectes acutatus</i> Cambess.	zoo
<i>Alchornea glandulosa</i> Poepp. & Endl.	zoo	<i>Calyptranthes brasiliensis</i> Spreng.	zoo
<i>Alchornea triplinervia</i> (Spreng.) Müll.Arg.	zoo	<i>Calyptranthes clusiifolia</i> O.Berg	zoo
<i>Allophylus edulis</i> Gracilis Radlk.	zoo	<i>Calyptranthes concinna</i> Spreng.	zoo
<i>Allophylus petiolulatus</i> Radlk.	zoo	<i>Calyptranthes widgreniana</i> O.Berg	zoo
<i>Allophylus racemosus</i> Sw.	zoo	<i>Campomanesia guaviroba</i> (DC.) Kiaersk.	zoo
<i>Allophylus semidentatus</i> (Miq.) Radlk.	zoo	<i>Campomanesia guazumifolia</i> (DC.) Kiaersk.	zoo
<i>Alseis floribunda</i> Schott	anemo	<i>Campomanesia velutina</i> (Cambess.) O.Berg	zoo
<i>Amaioua guianensis</i> Aubl.	zoo	<i>Campomanesia xanthocarpa</i> O.Berg	zoo
<i>Amaioua intermedia</i> Mart. ex Schult.	zoo	<i>Cariniana estrellensis</i> (Raddi) Kuntze	anemo
<i>Anadenanthera colubrina</i> (Vell.) Brenan	anemo	<i>Cariniana legalis</i> (Mart.) Kuntze	anemo
<i>Anadenanthera peregrina</i> (L.) Speg.	anemo	<i>Carpotroche brasiliensis</i> (Raddi) A Gray	zoo
<i>Andira fraxinifolia</i> Benth.	zoo	<i>Casearia arborea</i> (Rich.) Urb.	zoo

<i>Aniba canelilla</i> (Kunth) Mez	zoo	<i>Casearia decandra</i> Jacq.	zoo
<i>Aniba firmula</i> (Nees & Mart.) Mez	zoo	<i>Casearia gossypiosperma</i> Briq.	zoo
<i>Annona cacans</i> Warm.	zoo	<i>Casearia lasiophylla</i> Eichler	zoo
<i>Annona dolabripetala</i> Raddi	zoo	<i>Casearia obliqua</i> Briq.	zoo
<i>Annona emarginata</i> (Schltdl.) H.Rainer	zoo	<i>Casearia sylvestris</i> Jacq.	zoo
<i>Annona neosericea</i> Raddi	zoo	<i>Casearia ulmifolia</i> Vahl ex Vent.	zoo
<i>Annona sylvatica</i> Warm.	zoo	<i>Cassia ferruginea</i> (Schrad.) Schrad. ex DC.	zoo
<i>Aralia warmingiana</i> (Marchal) J.Wen	zoo	<i>Cecropia glaziovii</i> Snethl.	zoo
<i>Aspidosperma australe</i> Müll.Arg.	anemo	<i>Cecropia pachystachya</i> Snethl.	zoo
<i>Aspidosperma cylindrocarpon</i> Müll.Arg.	anemo	<i>Cedrela fissilis</i> Vell.	anemo
<i>Aspidosperma olivaceum</i> Müll.Arg.	anemo	<i>Ceiba speciosa</i> Vell.	anemo
<i>Aspidosperma parvifolium</i> A.DC.	anemo	<i>Celtis brasiliensis</i> (Gardner) Planch.	zoo
<i>Aspidosperma ramiflorum</i> Müll.Arg.	anemo	<i>Cestrum schlechtendalii</i> G.Don	zoo
<i>Aspidosperma spruceanum</i> Benth.	anemo	<i>Cheiloclinium cognatum</i> (Miers) A.C.Sm.	zoo
<i>Astronium fraxinifolium</i> Schott	anemo	<i>Chionanthus crassifolius</i> (Mart.) P.S.Green	zoo
<i>Astronium graveolens</i> Schott	anemo	<i>Chomelia brasiliana</i> A.Rich.	zoo
<i>Aureliana velutina</i> Sendtn.	zoo	<i>Chomelia sericea</i> A.Rich.	zoo
<i>Austrocritonia velutina</i> R.M.King & H.Rob.	anemo	<i>Chrysophyllum gonocarpum</i> Mart. & Eichler	zoo
<i>Balfourodendron molle</i> (Miq.) Pirani	anemo	<i>Chrysophyllum marginatum</i> Mart. & Eichler	zoo
<i>Balfourodendron riedelianum</i> (Miq.) Pirani	anemo	<i>Cinnamodendron dinisii</i> Schwacke	zoo
<i>Banara parviflora</i> (A.Gray) Benth.	zoo	<i>Cinnamomum glaziovii</i> Schwacke	zoo
<i>Bauhinia forficata</i> Link	aut	<i>Cinnamomum triplinerve</i> Schwacke	zoo
<i>Bauhinia longifolia</i> Link	aut	<i>Citronella paniculata</i> (Mart.) R.A.Howard	zoo
<i>Bauhinia rufa</i> (Bong.) Steud.	aut	<i>Clethra scabra</i> Pers.	anemo
<i>Beilschmiedia angustifolia</i> Kosterm.	zoo	<i>Clusia criuva</i> Cambess.	zoo
<i>Beilschmiedia taubertiana</i> Kosterm.	zoo	<i>Clusia organensis</i> Planch. & Triana	zoo
<i>Blepharocalyx salicifolius</i> (Kunth) O.Berg	zoo	<i>Coccoloba alnifolia</i> Casar.	zoo
<i>Boehmeria caudata</i> Sw.	anemo	<i>Coccoloba warmingii</i> Casar.	zoo
<i>Colubrina glandulosa</i> Perkins	zoo	<i>Coffea arabica</i> L.	zoo
<i>Connarus regnellii</i> G.Schellenb.	zoo	<i>Endlicheria paniculata</i> (Spreng.) J.F.Macbr.	zoo

<i>Copaifera langsdorffii</i> Desf.	zoo	<i>Eremanthus erythropappus</i> (DC.) MacLeish	anemo
<i>Cordia ecalyculata</i> Vell.	zoo	<i>Eriotheca candolleana</i> Steud.	anemo
<i>Cordia sellowiana</i> Sw.	zoo	<i>Erythrina falcata</i> Benth.	zoo
<i>Cordia trichoclada</i> L.	zoo	<i>Erythroxyllum citrifolium</i> A.St.-Hil.	zoo
<i>Cordia trichotoma</i> (Vell.) Arráb. ex Steud.	zoo	<i>Erythroxyllum cuneifolium</i> A.St.-Hil.	zoo
<i>Cordiaera concolor</i> Vell.	zoo	<i>Erythroxyllum deciduum</i> Benth.	zoo
<i>Cordiaera elliptica</i> (Cham.) Kuntze	zoo	<i>Erythroxyllum gonocladum</i> Mart.	zoo
<i>Cordiaera myrciifolia</i> (Cham.) Kuntze	zoo	<i>Erythroxyllum pelleterianum</i> A.St.-Hil.	zoo
<i>Cordiaera sessilis</i> (Vell.) Kuntze	zoo	<i>Erythroxyllum suberosum</i> Benth.	zoo
<i>Cordiaera vinosa</i> (Vell.) Kuntze	zoo	<i>Esenbeckia febrifuga</i> A. Juss. ex Mart.	aut
<i>Coussapoa microcarpa</i> (Schott) Rizzini	zoo	<i>Esenbeckia leiocarpa</i> (Cham.) Kuntze	aut
<i>Coutarea hexandra</i> (Jacq.) K.Schum.	anemo	<i>Eugenia acutata</i> Miq.	zoo
<i>Croton echinocarpus</i> Müll. Arg.	aut	<i>Eugenia adenantha</i> O.Berg	zoo
<i>Croton erythroxyloides</i> A.Rich.	aut	<i>Eugenia cerasiflora</i> O.Berg	zoo
<i>Croton floribundus</i> Müll. Arg.	aut	<i>Eugenia dodonaeifolia</i> O.Berg	zoo
<i>Croton organensis</i> Baill.	aut	<i>Eugenia florida</i> Miq.	zoo
<i>Croton urucurana</i> Baill.	aut	<i>Eugenia francavilleana</i> Miq.	zoo
<i>Cryptocarya aschersoniana</i> Mez	zoo	<i>Eugenia handroana</i> Meisn.	zoo
<i>Cryptocarya micranta</i> Meisn.	zoo	<i>Eugenia hiemalis</i> Cambess.	zoo
<i>Cupania emarginata</i> Cambess.	zoo	<i>Eugenia involucrata</i> DC.	zoo
<i>Cupania ludowigii</i> Somner & Ferrucci	zoo	<i>Eugenia longipedunculata</i> Nied.	zoo
<i>Cupania racemosa</i> Cambess.	zoo	<i>Eugenia mansoi</i> DC.	zoo
<i>Cupania vernalis</i> Cambess.	zoo	<i>Eugenia neomyrtifolia</i> Sobral	zoo
<i>Cupania zanthoxyloides</i> Radlk.	zoo	<i>Eugenia prasina</i> O.Berg	zoo
<i>Cybianthus cuneifolius</i> Mart.	zoo	<i>Eugenia puniceifolia</i> (Kunth) DC.	zoo
<i>Cybianthus detergens</i> Müll. Arg.	zoo	<i>Eugenia pyriformis</i> Cambess.	zoo
<i>Cybianthus peruvianus</i> Vell.	zoo	<i>Eugenia sonderiana</i> O.Berg	zoo
<i>Cybistax antisiphilitica</i> (Mart.) Mart.	zoo	<i>Eugenia umbellata</i> (Kunth) DC.	zoo
<i>Dahlstedtia muehlbergiana</i> Hassl. M.J.Silva	aut	<i>Eugenia widgrenii</i> Sond. ex O.Berg	zoo
<i>Dalbergia brasiliensis</i> Vogel	anemo	<i>Eumachia cymuligera</i> (Müll. Arg.) C.M. Taylor	zoo

<i>Dalbergia foliolosa</i> Benth.	anemo	<i>Eupatorium ganophyllum</i> A.St.-Hil.	anemo
<i>Dalbergia frutescens</i> (Vell.) Britton	anemo	<i>Euplassa inaequalis</i> Mart.	zoo
<i>Dalbergia miscolobium</i> Vell.	anemo	<i>Euplassa legalis</i> Benth.	zoo
<i>Dalbergia nigra</i> (Vell.) Allemão ex Benth.	anemo	<i>Euplassa organensis</i> (Gardner) I.M.Johnst.	zoo
<i>Dalbergia villosa</i> Benth.	anemo	<i>Euplassa rufa</i> (Gardner) I.M.Johnst.	zoo
<i>Daphnopsis brasiliensis</i> Mart.	zoo	<i>Faramea hyacinthina</i> Mart.	zoo
<i>Daphnopsis fasciculata</i> Mart.	zoo	<i>Faramea latifolia</i> Mart.	zoo
<i>Daphnopsis martii</i> Mart.	zoo	<i>Faramea nigrescens</i> (Cham. & Schltdl.) DC.	zoo
<i>Daphnopsis utilis</i> Warm.	zoo	<i>Ficus adhatodifolia</i> Schott in Spreng.	zoo
<i>Deguelia hatschbachii</i> A.M.G.Azevedo	anemo	<i>Ficus arpazusa</i> Casar.	zoo
<i>Dendropanax cuneatus</i> (DC.) Decne. & Planch.	zoo	<i>Ficus enormis</i> Casar.	zoo
<i>Dendrophorbium glaziovii</i> (DC.)	anemo	<i>Ficus insipida</i> Willd.	zoo
<i>Diatenopteryx sorbifolia</i> Radlk.	anemo	<i>Ficus mexiae</i> Standl.	zoo
<i>Diospyros hispida</i> A.DC.	zoo	<i>Ficus obtusifolia</i> Kunth	zoo
<i>Diospyros inconstans</i> A.DC.	zoo	<i>Ficus pertusa</i> Standl.	zoo
<i>Duguetia lanceolata</i> A.St.-Hil.	zoo	<i>Galipea jasminiflora</i> (A.St.-Hil.) Engl.	aut
<i>Duguetia salicifolia</i> A.St.-Hil.	zoo	<i>Garcinia brasiliensis</i> Mart.	zoo
<i>Geissanthus ambiguus</i> (Mart.) G.Agostini	zoo	<i>Garcinia gardneriana</i> (Planch. & Triana) Zappi	zoo
<i>Guapira graciliflora</i> Mart.	zoo	<i>Kielmeyera lathrophyton</i> Saddi	anemo
<i>Guapira hirsuta</i> Mart.	zoo	<i>Lacistema hasslerianum</i> Kunth	zoo
<i>Guapira opposita</i> (Vell.) Reitz	zoo	<i>Lafoensia pacari</i> A.St.-Hil.	anemo
<i>Guarea kunthiana</i> Vell.	zoo	<i>Lamanonia ternata</i> Vell.	anemo
<i>Guarea macrophylla</i> Vahl	zoo	<i>Laplacea fruticosa</i> Mart.	anemo
<i>Guatteria australis</i> A.St.-Hil.	zoo	<i>Leandra melastomoides</i> Vell.	zoo
<i>Guatteria pohliana</i> Mart.	zoo	<i>Leptolobium dasycarpum</i> Vogel	anemo
<i>Guatteria sellowiana</i> A.St.-Hil.	zoo	<i>Leucochloron incuriale</i> (Vell.) Barneby	anemo
<i>Guazuma ulmifolia</i> Lam.	zoo	<i>Lithraea molleoides</i> (Vell.) Engl.	zoo
<i>Guettarda uruguensis</i> Cham. & Schltdl.	zoo	<i>Lonchocarpus cultratus</i> Vell.	anemo
<i>Guettarda viburnoides</i> Cham. & Schltdl.	zoo	<i>Luehea candicans</i> Mart. & Zucc.	anemo
<i>Gymnanthes klotzschiana</i> Müll.Arg.	zoo	<i>Luehea divaricata</i> Mart. & Zucc.	anemo

<i>Gymnanthes schottiana</i> Müll.Arg.	zoo	<i>Luehea grandiflora</i> Mart. & Zucc.	anemo
<i>Handroanthus albus</i> (Cham.) Mattos	anemo	<i>Machaerium acutifolium</i> Vogel	anemo
<i>Handroanthus chrysotrichus</i> (Mart. ex DC.)	anemo	<i>Machaerium amplum</i> Benth.	anemo
<i>Handroanthus heptaphyllus</i> (Vell.) Mattos	anemo	<i>Machaerium brasiliense</i> Vogel	anemo
<i>Handroanthus ochraceus</i> (Vell.) Mattos	anemo	<i>Machaerium debile</i> Benth.	anemo
<i>Handroanthus serratifolius</i> (Vahl) S.Grose	anemo	<i>Machaerium hirtum</i> Vogel	anemo
<i>Handroanthus vellosi</i> (Mart. ex DC.)	anemo	<i>Machaerium isadelphum</i> Benth.	anemo
<i>Heisteria silvianii</i> Schwacke	zoo	<i>Machaerium lanceolatum</i> (Vell.) J.F.Macbr.	anemo
<i>Helicteres brevispira</i> A.St.-Hil.	aut	<i>Machaerium nyctitans</i> Vell.	anemo
<i>Helicteres ovata</i> A.St.-Hil.	aut	<i>Machaerium stipitatum</i> Benth.	anemo
<i>Heterocondylus alatus</i> (Vell.) R.M.King	anemo	<i>Machaerium villosum</i> Vogel	anemo
<i>Heteropterys byrsonimifolia</i> A.Juss.	zoo	<i>Machaonia acuminata</i> Bonpl.	aut
<i>Hieronyma ferrugínea</i> A.St.-Hil.	zoo	<i>Maclura tinctoria</i> (L.) D.Don ex Steud.	zoo
<i>Hirtella glandulosa</i> Spreng.	zoo	<i>Macropelplus dentatus</i> (Perkins) I.Santos	zoo
<i>Hirtella hebeclada</i> Spreng.	zoo	<i>Magnolia ovata</i> (A.St.-Hil.) Spreng.	zoo
<i>Hyeronima alchorneoides</i> Allemão	zoo	<i>Manihot grahamii</i> Hook.	zoo
<i>Hymenaea courbaril</i> L.	zoo	<i>Maprounea guianensis</i> Aubl.	aut
<i>Ilex brevicuspis</i> Reissek	zoo	<i>Marlierea gaudichaudiana</i> O.Berg	zoo
<i>Ilex cerasifolia</i> Reissek	zoo	<i>Marlierea obscura</i> O.Berg	zoo
<i>Ilex conocarpa</i> Reissek	zoo	<i>Matayba elaeagnoides</i> Radlk.	zoo
<i>Ilex paraguariensis</i> Reissek	zoo	<i>Matayba guianensis</i> Radlk.	zoo
<i>Ilex sapotifolia</i> Reissek	zoo	<i>Matayba juglandifolia</i> Radlk.	zoo
<i>Ilex theezans</i> Reissek	zoo	<i>Matayba marginata</i> Radlk.	zoo
<i>Inga affinis</i> DC.	zoo	<i>Matayba mollis</i> Radlk.	zoo
<i>Inga cylindrica</i> (Vell.) Mart.	zoo	<i>Maytenus glazioviana</i> Loes.	zoo
<i>Inga ingoides</i> Mart.	zoo	<i>Maytenus gonoclada</i> Mart.	zoo
<i>Inga marginata</i> Vell	zoo	<i>Maytenus salicifolia</i> Mart.	zoo
<i>Inga sessilis</i> (Vell.) Mart.	zoo	<i>Meliosma sellowii</i> Urb.	zoo
<i>Inga striata</i> DC.	zoo	<i>Meriania clausenii</i> (Naudin) Triana	zoo
<i>Inga vera</i> (Vell.) Mart.	zoo	<i>Metrodorea stipularis</i> Mart.	aut

<i>Ixora brevifolia</i> Benth.	zoo	<i>Miconia albicans</i> (Sw.) Triana	zoo
<i>Ixora gardneriana</i> Benth.	zoo	<i>Miconia argyrophylla</i> DC.	zoo
<i>Ixora warmingii</i> Benth.	zoo	<i>Miconia chartacea</i> Triana	zoo
<i>Jacaranda macranta</i> Cham.	anemo	<i>Miconia cinerascens</i> Triana	zoo
<i>Jacaranda puberula</i> Cham.	anemo	<i>Miconia cinnamomifolia</i> DC.	zoo
<i>Jacaranda subalpina</i> Cham.	anemo	<i>Miconia cubatanensis</i> Hoehne	zoo
<i>Jacaratia spinosa</i> (Aubl.) A.DC.	zoo	<i>Miconia eichleri</i> Triana	zoo
<i>Miconia minutiflora</i> (Bonpl.) DC.	zoo	<i>Miconia latecrenata</i> (DC.) Naudin	zoo
<i>Miconia paulensis</i> (Bonpl.) DC.	zoo	<i>Myroxylon balsamum</i> (L.) Harms	anemo
<i>Miconia pepericarpa</i> (Bonpl.) DC.	zoo	<i>Myroxylon peruiferum</i> L.f.	anemo
<i>Miconia prasina</i> (Sw.) DC.	zoo	<i>Myrsine coriácea</i> (Sw.) R.Br. ex Roem.	zoo
<i>Miconia pusilliflora</i> (DC.) Naudin	zoo	<i>Myrsine gardneriana</i> A.DC.	zoo
<i>Miconia schwackei</i> (Sw.) DC.	zoo	<i>Myrsine guianensis</i> (Aubl.) Kuntze	zoo
<i>Miconia sellowiana</i> Naudin	zoo	<i>Myrsine leuconeura</i> Mart.	zoo
<i>Miconia theizans</i> (Sw.) DC.	zoo	<i>Myrsine lineata</i> (Mez) Imkhan.	zoo
<i>Miconia trianae</i> Naudin	zoo	<i>Myrsine umbellata</i> (Mez) Imkhan.	zoo
<i>Miconia tristis</i> Spring	zoo	<i>Myrsine vilosíssima</i> Mart.	zoo
<i>Miconia urophylla</i> DC.	zoo	<i>Naucleopsis oblongifolia</i> (Kuhlm.) Carauta	zoo
<i>Miconia willdenowii</i> Spring	zoo	<i>Nectandra cissiflora</i> Nees	zoo
<i>Mollinedia argyrogyna</i> Perkins	zoo	<i>Nectandra grandiflora</i> Nees	zoo
<i>Mollinedia clavigera</i> Perkins	zoo	<i>Nectandra lanceolata</i> Nees	zoo
<i>Mollinedia lanceolata</i> Perkins	zoo	<i>Nectandra megapotamica</i> Nees	zoo
<i>Mollinedia triflora</i> (Spreng.) Tul.	zoo	<i>Nectandra membranácea</i> (Sw.) Griseb.	zoo
<i>Mollinedia widgrenii</i> (Spreng.) Tul.	zoo	<i>Nectandra nitidula</i> (Sw.) Griseb.	zoo
<i>Monteverdia aquifolia</i> (Mart.) Biral	zoo	<i>Nectandra oppositifolia</i> Nees	zoo
<i>Monteverdia communis</i> (Mart.) Biral	zoo	<i>Neocabreria pennivenia</i> (B.L.Rob.) R.M.King	anemo
<i>Monteverdia evonymoides</i> (Reissek) Biral	zoo	<i>Ocotea aciphylla</i> (Nees & Mart.) Mez	zoo
<i>Monteverdia gonoclada</i> (Mart.) Biral	zoo	<i>Ocotea bicolor</i> (Nees & Mart.) Mez	zoo
<i>Monteverdia ilicifolia</i> (Mart.) Biral	zoo	<i>Ocotea brachybotrya</i> (Meisn.) Mez	zoo
<i>Moquiniastrum paniculatum</i> (Less.) G. Sancho	anemo	<i>Ocotea corymbosa</i> (Meisn.) Mez	zoo

<i>Moquiniastrum polymorphum</i> (Less.) G. Sancho	anemo	<i>Ocotea diospyrifolia</i> (Meisn.) Mez	ZOO
<i>Mouriri glazioviana</i> Cogn.	ZOO	<i>Ocotea dispersa</i> (Nees & Mart.) Mez	ZOO
<i>Muellera campestres</i> (Mart. ex Benth.) M.J.	aut	<i>Ocotea glaziovii</i> Mez	ZOO
<i>Myrceugenia bracteosa</i> (DC.) D.Legrand	ZOO	<i>Ocotea indecora</i> Mez	ZOO
<i>Myrceugenia miersiana</i> (DC.) D.Legrand	ZOO	<i>Ocotea laxa</i> Mez	ZOO
<i>Myrcia aethusa</i> (O.Berg) Mattos	ZOO	<i>Ocotea longifolia</i> Kunth	ZOO
<i>Myrcia amazonica</i> DC.	ZOO	<i>Ocotea odorifera</i> Kunth	ZOO
<i>Myrcia anacardiifolia</i> (O.Berg) Mattos	ZOO	<i>Ocotea puberula</i> (Rich.) Nees	ZOO
<i>Myrcia eriocalyx</i> (O.Berg) Mattos	ZOO	<i>Ocotea pulchella</i> Kunth	ZOO
<i>Myrcia eriopus</i> DC.	ZOO	<i>Ocotea silvestres</i> (Rich.) Nees	ZOO
<i>Myrcia guianensis</i> (Aubl.) DC.	ZOO	<i>Ocotea velloziana</i> (Meisn.) Mez	ZOO
<i>Myrcia hebetepala</i> DC.	ZOO	<i>Ocotea velutina</i> (Meisn.) Mez	ZOO
<i>Myrcia laruotteana</i> DC.	ZOO	<i>Oreopanax capitatus</i> (Jacq.) Decne. & Planch.	ZOO
<i>Myrcia multiflora</i> DC.	ZOO	<i>Ormosia arborea</i> (Vell.) Harms	ZOO
<i>Myrcia mutabilis</i> (O.Berg) Mattos	ZOO	<i>Ormosia fastigiata</i> (Vell.) Harms	ZOO
<i>Myrcia obovata</i> (O.Berg) Nied.	ZOO	<i>Ouratea salicifolia</i> (A.St.-Hil. & Tul.) Engl.	ZOO
<i>Myrcia pulchra</i> (O.Berg) Nied.	ZOO	<i>Ouratea semiserrata</i> Engl.	ZOO
<i>Myrcia racemosa</i> (O.Berg) Kiaersk.	ZOO	<i>Peltogyne angustiflora</i> Ducke	aut
<i>Myrcia retorta</i> Cambess.	ZOO	<i>Peltophorum dubium</i> (Spreng.) Taub.	anemo
<i>Myrcia retusa</i> (O.Berg) Nied.	ZOO	<i>Pera anisotricha</i> Müll. Arg.	ZOO
<i>Myrcia splendens</i> (O.Berg) Nied.	ZOO	<i>Pera glabrata</i> Müll. Arg.	ZOO
<i>Myrcia subcordata</i> Cambess.	ZOO	<i>Persea major</i> (Meisn.) L.E.Kopp	ZOO
<i>Myrcia tomentosa</i> Cambess.	ZOO	<i>Persea willdenovii</i> (Meisn.) L.E.Kopp	ZOO
<i>Myrcia vellozoi</i> Mazine	ZOO	<i>Picramnia ciliata</i> Mart.	ZOO
<i>Myrcia venulosa</i> DC.	ZOO	<i>Picramnia glazioviana</i> Engl.	ZOO
<i>Myrciaria floribunda</i> (H.West ex Willd.)	ZOO	<i>Picramnia parvifolia</i> Engl.	ZOO
<i>Myrciaria tenella</i> (DC.) O.Berg	ZOO	<i>Picrasma crenata</i> (Vell.) Engl.	ZOO
<i>Pimenta pseudocaryophyllus</i> (Gomes) Landrum	ZOO	<i>Pilocarpus spicatus</i> (Vell.) Engl.	aut
<i>Piper aduncum</i> (C.DC.) Yunck.	ZOO	<i>Sacoglottis mattogrossensis</i> Malme	ZOO
<i>Piper arboreum</i> L.	ZOO	<i>Salacia elliptica</i> (Mart.) G. Don	ZOO

<i>Piper cernuum</i> Vell.	zoo	<i>Salix humboldtiana</i> Willd.	anemo
<i>Piper gaudichaudianum</i> Vell.	zoo	<i>Sapium glandulosum</i> (L.) Morong	zoo
<i>Piptadenia adiantoides</i> (Spreng.) J.F.Macbr.	anemo	<i>Schefflera angustissima</i> (Marchal) Frodin	zoo
<i>Piptadenia gonoacantha</i> (Spreng.) J.F.Macbr.	anemo	<i>Schefflera calva</i> Marchal.	zoo
<i>Piptocarpha axillaris</i> (Less.) Baker	anemo	<i>Schinus lentiscifolia</i> Marchand	zoo
<i>Piptocarpha macropoda</i> (Less.) Baker	anemo	<i>Schinus terebinthifolia</i> Marchand	zoo
<i>Pisonia zapallo</i> Griseb.	zoo	<i>Schizolobium parahyba</i> (Vell.) Blake	aut
<i>Platycyamus regnellii</i> Benth.	anemo	<i>Schoepfia brasiliensis</i> A.DC.	zoo
<i>Platypodium elegans</i> Vogel	anemo	<i>Sebastiania brasiliensis</i> Spreng.	aut
<i>Plenckia populnea</i> Reissek	anemo	<i>Sebastiania commersoniana</i> Mull. Arg	aut
<i>Pleroma candolleianum</i> (Mart. ex DC.)	aut	<i>Seguiera langsdorffii</i> Moq.	anemo
<i>Pleroma stenocarpum</i> (Mart. ex DC.)	aut	<i>Senecio glaziovii</i> A.DC.	anemo
<i>Plinia cauliflora</i> (Mart.) Kausel	zoo	<i>Senegalia langsdorffii</i> (Benth.) Seigler & Ebinger	aut
<i>Plinia grandifolia</i> (Mart.) Kausel	zoo	<i>Senegalia polyphylla</i> Benth.	aut
<i>Podocarpus sellowii</i> Mart.	zoo	<i>Senegalia recurva</i> Benth.	aut
<i>Pourouma guianensis</i> Aubl.	zoo	<i>Senna macranthera</i> (DC. ex Collad.) H.S.Irwin	zoo
<i>Pouteria gardneriana</i> (A.DC.) Radlk.	zoo	<i>Senna multijuga</i> (Rich.) H.S.Irwin & Barneby	zoo
<i>Prockia crucis</i> P.Browne ex L.	zoo	<i>Senna velutina</i> (Rich.) H.S.Irwin & Barneby	zoo
<i>Protium heptaphyllum</i> (Aubl.) Marchand	zoo	<i>Sorocea bonplandii</i> (Baill.) W.C.Burger et al.	zoo
<i>Protium spruceanum</i> (Aubl.) Marchand	zoo	<i>Strychnos brasiliensis</i> Mart.	zoo
<i>Protium widgrenii</i> Aubl.	zoo	<i>Strychnos gardneri</i> Mart.	zoo
<i>Prunus myrtifolia</i> (L.) Urb.	zoo	<i>Stryphnodendron polyphyllum</i> Mart.	zoo
<i>Pseudobombax grandiflorum</i> (Cav.) A.Robyns	anemo	<i>Styrax acuminatus</i> (Seub.) Perkins	zoo
<i>Pseudopiptadenia leptostachya</i> Benth.	anemo	<i>Styrax camporum</i> Pohl	zoo
<i>Psidium cattleianum</i> Sabine	zoo	<i>Styrax ferrugineus</i> Pohl	zoo
<i>Psidium cupreum</i> DC.	zoo	<i>Styrax latifolius</i> Pohl	zoo
<i>Psidium guineense</i> Sw.	zoo	<i>Styrax pohlii</i> Pohl	zoo
<i>Psidium rufum</i> Mart. ex DC.	zoo	<i>Symplocos celastrinea</i> Mart.	zoo
<i>Psychotria carthagenensis</i> Jacq.	zoo	<i>Symplocos insignis</i> Mart.	zoo
<i>Psychotria deflexa</i> Jacq.	zoo	<i>Symplocos oblongifolia</i> Casar.	zoo

<i>Psychotria suterella</i> Müll.Arg.	zoo	<i>Symplocos pubescens</i> Klotzsch ex Benth.	zoo
<i>Psychotria vellosiana</i> Müll.Arg.	zoo	<i>Syzygium jambos</i> (L.) Alston	zoo
<i>Qualea cordata</i> Spreng.	zoo	<i>Tabernaemontana hystrix</i> Steud.	zoo
<i>Qualea cryptantha</i> Spreng.	zoo	<i>Tachigali rugosa</i> Müll.Arg.	anemo
<i>Qualea grandiflora</i> Spreng.	zoo	<i>Tapirira guianensis</i> Aubl.	zoo
<i>Qualea multiflora</i> Mart.	zoo	<i>Tapirira obtusa</i> Aubl.	zoo
<i>Randia armata</i> (Sw.) DC.	zoo	<i>Terminalia glabrescens</i> Mart.	anemo
<i>Randia ferox</i> (Cham. & Schltdl.) DC.	zoo	<i>Ternstroemia brasiliensis</i> Cambess.	zoo
<i>Rhamnidium elaeocarpum</i> Reissek	zoo	<i>Tibouchina sellowiana</i> Cogn.	anemo
<i>Rhamnus sphaerosperma</i> Sw.	zoo	<i>Toulicia subsquamulata</i> Radlk.	anemo
<i>Rollinia neolaurifolia</i> Reissek	zoo	<i>Tovomitopsis paniculata</i> (Spreng.) Planch.	zoo
<i>Roupala longepetiolata</i> Pohl	anemo	<i>Tovomitopsis saldanhae</i> Spreng.	zoo
<i>Roupala montana</i> Pohl	anemo	<i>Trattinnickia ferrugínea</i> Kuhlm.	zoo
<i>Rudgea jasminoides</i> (Cham.) Müll.Arg.	zoo	<i>Trema micranta</i> (L.) Blume	zoo
<i>Rudgea triflora</i> Müll.Arg.	zoo	<i>Trembleya parviflora</i> (D.Don) Cogn.	aut
<i>Rudgea viburnoides</i> Müll.Arg.	zoo	<i>Trichilia casaretti</i> C.DC.	zoo
<i>Ruprechtia laxiflora</i> Meisn.	anemo	<i>Trichilia catiguá</i> C.DC.	zoo
<i>Simira pikia</i> (K.Schum.) Steyerl.	anemo	<i>Trichilia pallens</i> Sw.	zoo
<i>Siparuna brasiliensis</i> (Spreng.) A.DC.	zoo	<i>Trichilia pallida</i> Sw.	zoo
<i>Siparuna guianensis</i> (Spreng.) A.DC.	zoo	<i>Urbanodendron verrucosum</i> (Nees) Mez	zoo
<i>Siphoneugena crassifolia</i> Proença & Sobral	zoo	<i>Urera bacífera</i> (L.) Gaudich. ex Wedd.	zoo
<i>Siphoneugena densiflora</i> O.Berg	zoo	<i>Vantanea compacta</i> (Schnizl.) Cuatrec.	zoo
<i>Siphoneugena kiaerskoviana</i> (Burret) Kausel	zoo	<i>Vernonanthura discolor</i> (Spreng.) H.Rob.	anemo
<i>Siphoneugena kuhlmannii</i> Mattos	zoo	<i>Vernonanthura divaricata</i> Spreng.	anemo
<i>Siphoneugena reitzii</i> D.Legrand	zoo	<i>Vernonanthura polyanthes</i> (Sprengel) Vega	anemo
<i>Sloanea guianensis</i> (Aubl.) Benth.	zoo	<i>Viola bicuhyba</i> (Schott ex Spreng.) Warb.	zoo
<i>Sloanea hirsuta</i> Aubl.	zoo	<i>Vismia brasiliensis</i> Choisy	zoo
<i>Solanum cernuum</i> Vell.	zoo	<i>Vismia guianensis</i> (Aubl.) Choisy	zoo
<i>Solanum granuloso-leprosum</i> Vell.	zoo	<i>Vitex agnus-castus</i> L.	zoo
<i>Solanum leucodendron</i> Vell.	zoo	<i>Vitex cymosa</i> Bertero ex Spreng.	zoo

<i>Solanum pseudoquina</i> A.St.-Hil.	zoo	<i>Vitex megapotamica</i> (Spreng.) Moldenke	zoo
<i>Solanum swartzianum</i> Roem. & Schult.	zoo	<i>Vitex polygama</i> Cham.	zoo
<i>Vochysia magnifica</i> Warm.	Anemo	<i>Xylosma prockia</i> (Turcz.) Turcz.	zoo
<i>Vochysia oppugnata</i> (Vell.) Warm.	Anemo	<i>Zanthoxylum fagara</i> (L.) Sarg.	zoo
<i>Vochysia schwackeana</i> Vell.	Anemo	<i>Zanthoxylum monogynum</i> A.St.-Hil.	zoo
<i>Vochysia tucanorum</i> Vell.	Anemo	<i>Zanthoxylum rhoifolium</i> Lam.	zoo
<i>Xylopiá aromática</i> (Lam.) Mart.	Zoo	<i>Zanthoxylum riedelianum</i> Engl.	zoo
<i>Xylopiá brasiliensis</i> Spreng.	Zoo	<i>Zeyheria tuberculosa</i> (Vell.) Bureau ex Verl.	anemo
<i>Xylosma ciliatifolia</i> (Clos) Eichler	Zoo		

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